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FACTORS EFFECTING PHYTOPLANKTON ASSEMBLAGES IN THE LAFAYETTE RIVER ESTUARY

by

Laurie Ann Kalenak B.A. May 1979, Dowling College

A Thesis Submitted to the Faculty of Old Dominion University in Partial Fulfillment of the Requirements for the Degree of

MASTER OF SCIENCE

OCEANOGRAPHY

OLD DOMINION UNIVERSITY September 1982

Approved by:
Harold G. Marshall, Director
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ABSTRACT

FACTORS EFFECTING PHYTOPLANKTON ASSEMBLAGES IN THE LAFAYETTE RIVER ESTUARY

Laurie Ann Kalenak
Old Dominion University, 1982
Director: Dr. Harold G. Marshall

Chemical and physical parameters were measured with phytoplankton species composition and abundance in the Lafayette River from August to October 1981. Stations located in four distinct areas of the river were statistically analyzed to determine data relationships. Environmental factors considered as potentially influencing the presence and numbers of phytoplankton were salinity, temperature, Secchi depth, tidal phase, orthophosphate, combined nitrates and nitrites, ammonia, and reactive silicates.

The River mouth had higher salinity and nutrient values, with lower temperatures than the other river sections. Diatoms were the dominant cells in this section of the River. At mid-river, salinity and nutrient concentrations decreased, with higher temperatures noted. Common to this area were diatoms and a larger number of phytoflagellates. In the two River branches, flagellated cells were dominant, with increasing numbers of chlorophytes and cyanophytes. Environmental conditions associated with

these areas were low salinity, high temperatures and increased nutrient levels.

Discriminant Function and Pearson Correlation analyses were conducted separately on environmental and biological data sets. Adjacent stations were not significantly different environmentally. However, in areas located other than next to one another, clusters were statistically different at the α <.05 level. In analyses using the phytoplankton data set, all areas were statistically different at the α <.005 level of significance.

Visual comparisons between the two sets of analyses showed that stations grouped in the same cluster 75% of the time. Of the remaining fraction, half of the cases were associated with severe storm conditions.

DEDICATION

To my parents: Marion and Terrence Kalenak

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I am eternally grateful to my thesis committee director, boss, and friend, Dr. Harold Marshall. His patience and understanding guided me through many a troubled time. I owe him many thanks, for without his help, this project could not have been completed.

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Chapter 1

INTRODUCTION

The purpose of this study was to determine existing relationships between the phytoplankton composition and environmental parameters of the Lafayette River from August to October 1981. No previous studies have been conducted in this river to associate seasonal phytoplankton assemblages to specific chemical or physical parameters. However, the influence of water chemistry and other factors on phytoplankton populations has been well documented. For instance, the presence and concentrations of various nutrients have been reported to directly influence the growth of phytoplankton (Raymont, 1980).

Seasonal composition and abundance changes in response to environmental fluxes are also common (Riley, 1947;

Pratt, 1965; Frey and Small, 1980). These variations can significantly alter the primary productivity of an area.

Specifically, in the Lafayette River estuary, Purcell (1973) noted a bimodal distribution of cells and their abundance.

Phytoplankton, in terms of numbers, were elevated during spring and autumn. Summer was characterized by a series of pulses, with numbers low in autumn and winter. Correspondingly, Montgomery (1972) found relatively high levels of phosphate, nitrate, and nitrite during spring and summer in the Lafayette River when compared to other local estuarine

systems. He attributed the elevated values to sewage outfall, runoff, and increased use of lawn fertilizer. Lowest concentrations were noted during autumn and winter months.

In many natural water systems, low concentrations of one or more nutrients can limit phytoplankton growth.

Kowever, in the Lafayette River, the major nutrients (phosphorus and nitrogen) have been abundant (Montgomery, 1972).

Nielson and Sturm (1978) attributed these high values partially to wastewater discharged from local sewage treatment plants. This additional input from runoff and sewage discharge, combined with a low flushing rate would increase eutrophication in the Lafayette River. Initially, this may enhance phytoplankton growth, but with the additional production of organic matter and oxygen depletion, this pattern becomes detrimental (Hodges, 1977). Subsequently, phytoplankton concentrations may decrease, reducing the productivity of the area.

In order to determine relationships between phytoplankton and environmental conditions, both spatial and temporal changes must be investigated. Specific environmental differences were noted by White (1972) in his study of the Lafayette River. He divided the River into four sections based on physical parameters and shoreline contour. Each region had characteristic mixing processes, current velocity, and water density. It is surmised that these factors may also influence the types and numbers of phytoplankton in these waters.

Chapter 2

LITERATURE REVIEW

Studies of phytoplankton in the Chesapeake Bay area began with Wolfe et al. (1926). The authors found five types of variation within the community. These consisted of temporal (hourly), vertical, spatial (horizontal), diurnal, and seasonal differences. Peaks in numbers were found to correspond with periods of increased rainfall in late spring and fall. The first peak was characterized by small sized diatoms indicative of good growing conditions. The autumn peak included diatoms, with the dinoflagellate genera Ceratium, Prorocentrum, and Peridinium (Protoperidinium) abundant. Patten, Mulford, and Warinner (1963) found diatoms to be important during colder months, with phytoflagellates dominant in summer. At stations in the lower Chesapeake Bay, the authors found the autumn peak associated with high dissolved orthophosphate concentrations.

Marshall (1980) also noted a bimodal pattern of phytoplankton composition in the lower Chesapeake Bay. He found Skeletonema costatum dominant in autumn and winter. In spring, dominance shifted to other diatoms, with the genera Asterionella, Thalassiosira, Cyclotella, and Ceratulina being common. In summer, the dinoflagellates Gymnodinium spp. and Gyrodinium spp. were abundant. He also noted that population exchanges occurred between the waters of the Bay

and Old Plantation Creek.

Similar growth patterns and species composition have been noted in Virginia coastal waters (Mulford and Norcross, 1971). In addition, Purcell (1973) and Golub (1972) found chain-like diatoms present in high numbers during all but summer months in the Lafayette River. At this time, the phytoflagellates became the dominant cells. Marshall (1966, 1967a, 1967b, and 1968) found many of these same species common in the James River, Willoughby Bay and Hampton Roads, the Elizabeth and Lafayette Rivers. At these sites, small, chain-like diatoms were noted in high numbers between January and August. Phytoflagellates then became dominant during warmer months.

Golub (1972) found temporal changes in dominant species over a year period in the Lafayette River. For most of this time, diatoms were most numerous in his centrally located station. During the summer, however, phytoflagellates increased and became dominant until autumn. Purcell (1973) noted a similar pattern at his stations in the Lafayette River.

Phytoplankton populations are subject to change as influenced by the local physical, chemical, biological, and geological factors. A change in one or more of these variables may initiate a change in the population structure of the phytoplankton and the degree of temporal heterogeneity for an area (Raymont, 1980; Smayda, 1958). Hulburt (1970) noted that different marine systems had different diversities

and abundances of phytoplankton. He found that, in shallow estuaries, the competition for nutrients would often lead to the dominance of one species over others. The elimination of some species would eventually ensue. If the availability of nutrients varies, species composition may also vary. For instance, in the Forge River, Barlow, Lorenzen, and Myren (1963) found added nutrients from runoff promoted a pattern of algal growth. This development also influenced the adjacent population of downstream, coastal waters, where cell concentrations increased.

In nearshore systems, nitrogen has been shown to limit phytoplankton growth (Williams, 1972). Williams found nutrient enrichment caused a shift in species to those adapted to high nutrient levels. Taft, Elliot, and Taylor (1977) determined that the nitrogen fluxes within the Chesapeake Bay were cyclic events. The net flux of ammonia and nitrate-nitrogen was seaward in the winter and landward in spring. Pratt (1965) monitored levels of nitrogen, phosphorus, and silica over several years in the Narragansett Bay. The diatom maximum seen during the spring bloom appeared to be regulated by nitrogen and silica concentrations. Cell numbers increased until the nitrogen sources were exhausted. However, the final number of diatoms was a function of the initial silicate concentration. The maximum nutrient levels were reached about two weeks before bloom conditions were attained.

In nutrient enrichment studies conducted under

laboratory conditions, Thayer (1974) and Frey and Small (1980) found both phosphorus and nitrogen limiting in certain estuarine waters. Thayer found nitrogen the most severely limiting nutrient; however, the most increased growth occurred with additions of both nutrients. Frey and Small found both major and micronutrients limiting, but final yields depended on only initial major nutrient levels.

Many physical parameters of estuarine systems have been related to community structure changes in phytoplankton.

Temperature and salinity are the most discussed factors that potentially alter the presence and numbers of these populations. Harrison and Platt (1980) noted in a Canadian inlet that forty percent of the variation in phytoplankton assimilation number was attributed to temperature. Although the number of cells per liter correlated to changes in water temperature, changes in species composition had no apparent relationship. In similar analyses, Platt, Dickie, and Trites (1970) found high cell numbers associated with high temperatures and high salinities.

In cultures exposed to different salinities, Paasche (1975) reported cell metabolism was altered as the salt content changed. Paasche concluded that salinity was an important factor influencing cell growth, especially in zones between salt and fresh water. In a field study, Ryther et al. (1958) found the dominance of diatoms was inversely related to salinity in a small tidal creek.

Water column depth and Secchi depth values were

important parameters associated with phytoplankton assemblages in the Saint-Lawrence Estuary (Sinclair, Subba Rao, and Couture, 1981). Rainfall and the amount of runoff were also considered important to a lesser degree. However, samples taken at frequent intervals did not reveal what caused changes in these phytoplankton populations. Pingree et al. (1977) found thermocline formation, light penetration, and mixing processes were influential in determining types and abundances of cells. Other factors relating to changes in algal community structures were wind stress (Therriault, Lawrence, and Platt, 1978), rainfall (Reimold and Diaber, 1967), and density fronts (Incze and Yentsch, 1981).

Interactions between phytoplankton and other biota can also help determine the numbers and kinds of cells present. Of these, the most obvious is the relationship between predator and prey. Bainbridge (1953) found zooplankton grazing important in the spatial heterogeneity of algal cells. Steele (1974) based his model of heterogeneity on the continual changes in phytoplankton and zooplankton populations. As copepod numbers increased, phytoplankton decreased in numbers due to grazing. In adjacent areas, grazing pressure was relaxed, and cell growth increased. In St. Margaret's Bay, Nova Scotia, Therriault and Platt (1978) noted differential zooplankton grazing was an important factor in creating and maintaining heterogeneity among phytoplankton.

Interactions between members of the phytoplankton

community can also determine numbers and species present (Haury, McGowan, and Wiebe, 1978). If a reproductive rate is at a maximum, or a species can out-compete all others, that species will become dominant. Each species has a set of environmental conditions that are optimal for growth. As the conditions change, the species present and their numbers correspondingly change. Diurnal migratory patterns, sinking, and competitive exclusion may also influence which species will survive in an area.

The most reliable population studies include a full array of ecological variables that may effect phytoplankton growth and survival. Riley's (1947) model includes both physical and biological parameters noted to affect changes in the phytoplankton community off Georges Bank. Riley takes into consideration the rates of respiration and grazing, nutrient concentrations, and physical parameters. In a review of studies describing factors affecting the distribution of cells, Smayda (1958) indicated most studies included the same measurements as used in the present study. The majority of studies examined various nutrient and physical factors in relation to the changing phytoplankton population. Jeffries (1962) also noted compositional changes in the plankton community of the Raritan Bay with the fluctuation of environmental factors.

Chapter 3

HYDROGRAPHY

The Lafayette River estuary is located in Norfolk, Virginia. It joins the Elizabeth River in entering Hampton Roads, which connects with the lower Chesapeake Bay (Figure 1). The River has a length of approximately 11 km, and has two branches. Water depth at mid channel ranges from 0.3 to 7.0 m, and width from 115 to 638 m. The mean water volume is $2.66 \times 10^7 \, \mathrm{m}^3$ (White, 1972).

The surrounding area is primarily urban, with modest industrial development and several small marinas along its shoreline. Discharge pipes from local storm sewers empty into the main river section and both branches. Commercial and private fishing has been prohibited in the Lafayette due to pollution by industry and local runoff. In addition, several sewage treatment plants (Figure 1) discharge wastes into nearby waters.

Classification of this estuary ranges from B/C (partially mixed) to D, sectionally homogeneous and well-mixed (White, 1972). It is also tidally influenced, with a mean tidal range of 0.82 m. Using White's net non-tidal current velocity measurement of 0.0061 meters per second out of the estuary, a flushing time of 2.10 days is determined. The amount of time needed to totally replace the water on a volume to volume ratio is slightly greater than 2 months,

assuming 100% of the average yearly rainfall. With a reduction to 60% of the average yearly rainfall, this time would increase to over 10 months (White, 1972). Since the River received a rainfall well below this average in 1981, this replacement time would be approximately four months.

Chapter 4

METHODS AND MATERIALS

Weekly cruises were made during the period of study in the Lafayette River, where 15 stations were established (Figure 2). Visitations were made from 17 August to 26 October 1981 on the skiff ODU-3. Cruises began at 1000 hours and ended no later than 1600 hours. Stations were occupied for approximately 15 minutes apiece. Stations were a distance of 1 km apart, Station 1 being located at the mouth of the River. In both branches, the distance between stations was decreased to 0.5 km. Samplings were conducted at the visual center of the River, since cross-sectional homogeneity exists (White, 1972).

At each station salinity, temperature, water transparency, column depth, and tidal phase were recorded.

Salinity and temperatures were measured using a Beckman inductive salinometer, model RS-5. Water transparency was determined using a Secchi disk.

Water samples were collected using 1.5 liter polypropylene NIO water bottles. Surface and bottom samples were taken at those stations where the water depth was 2 m or greater. At stations with depths less than 2 m, one middepth sample was taken. For each depth a 500 ml sample was placed in a polyethylene bottle and preserved with a modified Lugol's solution (Verduin, 1962). Triplicate water

samples were taken for nutrient analysis and placed in 250 ml polyethylene bottles. At stations 1, 4, 9, and 13 triplicate 250 ml samples were taken for use in the analysis of reactive silicates.

Samples for nutrient analyses were filtered with a vacuum pump using Gelman A/E glass fiber filters upon returning to the laboratory. Silicate samples were vacuum filtered using Millipore membrane filters. Nutrient analyses were completed within 48 hours of sampling.

Nutrient analyses for ammonia, nitrates and nitrites, and orthophosphate were done using an auto-analyzer, Scientific Instruments, model 200, with associated manifolds. Reactive silicate analysis was done according to Strickland and Parsons (1977) using a Beckman DU spectrophotometer.

In order to more efficiently evaluate the number of phytoplankton samples, the environmental data were statistically grouped. To standardize these data, numbers obtained were divided by the highest values seen for each variable. In doing this, the high concentrations found for one parameter could be treated the same as the low values for another without the first overshadowing the second. In addition, a log transform was performed on all environmental parameters. A non-hierarchal cluster analysis (Gauch, 1979) was used to group like stations. A program to condense the data for input into this Compelus program (Singer and Gauch, 1979) was also implemented. The clustering method was repeated until the lowest number of clusters was obtained.

The data set was then split into those stations at which silicates had been determined and stations with silicate removed from the analysis. This was employed because the clustering program used did not allow for the missing silicate values. Unfortunately, the latter data set exceeded the capacity of the SPSS programs used and no further analyses could be used. Thus, all analyses referred to include only those stations where reactive silicates were measured. Data from the other set are included in the appendices.

Output from the Compclus program also included a priority listing of factors important to each group. The SPSS version of Discriminant Function Analysis (Nie et al., 1975) was performed on the data to determine whether the clusters found were statistically different. This also indicated if those variables listed in the Compclus output truely distinguished stations. A Pearson Correlation (Nie et al., 1975) was then conducted to determine the relationship between the discriminant functions obtained and the original environmental parameters.

Since the lowest number of collections in any cluster was eight, eight phytoplankton samples from each group were chosen randomly for analysis. Water samples obtained had been allowed to settle and siphoned to a volume between 20 and 40 ml. The concentrate was transferred to a glass vial and labelled. Because of the dense phytoplankton population, a dilution process was necessary to allow for more

reliable identification and enumeration of cells. Phytoplankton were identified using a modified Utermohl method and a Zeiss inverted microscope. Random fields were examined at 312X and 500X. This was repeated until a minimum of 10 fields or 200 cells were observed. Major keys used to identify cells included Cupp (1943), Drouet (1973), Prescott (1951), and Schiller (1933, 1937). Abundance was determined as the number of cells per liter.

Initial clustering of phytoplankton data proved inconclusive because the analysis grouped stations using the species of higher numbers as a priority criterion. However, these species were ubiquitous and were removed from the data set. Also, species that were found less than three times during the sampling period were deleted using a screening program (Gaugh, 1973), for similar reasons. Discriminant and Pearson Correlation Analyses were then performed on the cell counts, as with the environmental parameters. All patterns defined by the analyses of biological and environmental data sets were compared visually, as no statistical test is available.

Chapter 5

RESULTS

The physical data from the four stations that were sampled per cruise and used in the statistical analyses are listed in Table 1. Collections occurred during all phases of the tidal cycle. Water column depth ranged from 4.5 m at the River mouth to 0.5 m in the River branches. Secchi depth readings decreased up river, indicating increased turbidity. Temperature ranged over the study from a high of 27.58°C to a low of 15.27°C, with water temperatures always highest at stations No. 9 and 13, the stations furthest up river. Temperature decreased as sampling continued into autumn. Salinity ranged from 14.23 to 23.40% oo, with lowest values in the branches of the Lafayette River. Values below 160/∞ represent periods of increased rainfall and fresh water input into the system. During this period of time, the Norfolk area was subject to severe storm action and increased precipitation. At these times, salinity was markedly decreased, particularly in the branches.

The results of chemical analyses are given in Table 2 for stations 1, 4, 9, and 13. These are shown graphically in Figures 3-6. Ammonia concentrations ranged from 0.28 to 226.81 µg N/liter. These amounts varied with the location of the River sampled. High values were noted during heavy rain conditions at stations where storm sewers were abundant

along river banks. These conditions existed at stations 1 and 9. Lowest values were observed at station 13. Combined nitrate and nitrite values ranged from 100 to 1900 µg N/liter. Highest concentrations were at the mouth of the Lafayette River, and were possibly influenced by discharge from local sewage treatment plants. Lowest values were at stations located in both river branches. Orthophosphate concentrations ranged from 64.08 to 1697.96 µg P/liter. Highest values were observed at the mouth of the River, and may also be associated with sewage effluents mixing with these waters. Lowest values were recorded at station 4, located midway up the River, with concentrations increasing in the branches. Reactive silicate values ranged from 8.85 to 84.00 µg Si/liter. These values followed the same pattern as nitrate/nitrite values, being high at the River mouth and lower in the branches.

Four different groups representing the four areas of the River sampled were obtained from the Compolus clustering program. These are indicated by the separate histograms in Figure 7. Factors used in separating these stations are listed in order of importance in Table 3. In the River mouth, high nitrogen and phosphate nutrient values distinguished this area from others. At station 4, reactive silicates, salinity, and temperature were important in identifying this region. In the southern branch (station 9), low nitrogen and high phosphate values were characteristic, whereas the northern branch (station 13) had moderate

concentrations of the major nutrients.

Discriminant Analysis results are given in Figure 7. The first discriminant function formed from this program separated groups along the X-axis. Since this was the only function of three that was statistically significant, it is the only one considered in further analyses. This explained nearly 80% of the variation in the data. The histograms represent the frequency of discriminant scores of each cluster. Groups 1 and 3, 1 and 4, 2 and 4, and 3 and 4 were significantly different at the α =.05 level. However, groups 1 and 2, and 2 and 3 were not statistically different. indicates a continuum, rather than separated, distinct areas. Results of the Pearson Correlation are listed on the X-axis of the graph in Figure 7. The variables listed represent environmental parameters important in determining the discriminant function and are listed in order of importance. An asterisk indicates the variable was statistically important in the Pearson Correlation Analysis. Reactive silicate was not included in the first discriminant function, but was significant in the Pearson Correlation, presumably because it followed the same pattern as another variable. Salinity was noted to be the factor explaining most of the variance between stations. Temperature, tide, combined nitrates and nitrites, ammonia, Secchi depth, and orthophosphate respectively were of decreasing importance in explaining differences in the data set.

The bars above the initial clusters represent the 95%

confidence regions calculated according to Sokol and Rohlf (1981). The X's indicate locations of groups centroids (means). The bars represent the probability that 95% of the time the group centroids would fall within these ranges if the entire experiment were repeated. Those bars that overlap on the X-axis indicate areas of the River not clearly separated by the discriminant functions.

Results from the phytoplankton identification and enumeration are listed in Tables 4-35. These are shown graphically in Figures 8-11. Ubiquitous species were Cylindrotheca closterium, Cryptomonas sp., Leptocylindrus minimus, Pyramimonas sp., and Gymnodinium nelsonii. In addition, an unidentified nanoplankton fraction was present at all stations. These cells, noted as "Green Spheres" in the tables, were separated by size.

Cells important to the four clusters are listed in Table 36. High concentrations of diatoms were characteristic of the stations forming the first cluster. These stations, located at the mouth of the Lafayette River, also had low numbers of phytoflagellates. Common species were Cylindrotheca closterium, Leptocylindrus minimus, Skeletonema costatum, and Thalassiosira nana. At mid-river stations, diatoms remained dominant, with phytoflagellates increasing in number. Actinoptychus senarius, Nitzschia delicatissima, Plagiogramma staurophorum, Amphidinium acutum, and Calycomonas spp. were abundant at these stations. In the southern branch of the River (station 13), low diatom

and high phytoflagellate concentrations were noted.

Gymnodinium nelsonii, Prorocentrum minimum and Scrippsiella tricoidea increased in number, with the euglenoid Eutreptia lanowii also important. Although not statistically important to this cluster of stations, an increase of blue-green cells was noted. At station 9, located in the northern branch of the Lafayette, phytoflagellates and an unidentified chlorophyte were reported. Common species included Amphidinium acutum, Prorocentrum minimum, and Scrippsiella tricoidea.

In terms of numbers, total cells per liter increased up river. This was mainly due to the unidentified nanoplankton fraction. Total phytoplankton concentration ranged from an average of nearly 5 million cells per liter at the River mouth to 13.5 million in River branches.

The results of all statistical analyses on cell counts are given in Figure 12. The numbers 1 through 4 represent clusters formed by the Compclus program. The X-axis separates these groups according to the first discriminant function formed. The second discriminant function is shown on the Y-axis. Together, these explain 99.9% of the variation in the data set. All groups were statistically different at the α <.005 level. The results of the Pearson Correlation are those factors significantly related to the discriminant functions. These are listed along the axes. Those species important to the Pearson Correlation Analysis are indicated by an asterisk. Cylindrotheca closterium,

Eutreptia lanowii, Tabellaria fenestrata, Prorocentrum micans, and the unknown centrales (>20 μm) were not included in the Discriminant Analysis, but significant in the Pearson Correlation. It is presumed these species follow a pattern similar to species significant in the Discriminant Analysis, but do not separate groups as well.

Confidence regions at the α =.05 significance level are represented by the ellipses enclosing discriminant scores of like collections (Sokal and Rohlf, 1981). These represent the probability that 95% of centroids (means) from any group would fall within its respective region if the experiment were repeated. For example, in a similar experimental procedure, 95% of the time the centroid of the discriminant scores from Group 1 would fall within the ellipse for group 1.

Table 37 represents the comparison between stations based on environmental and biological variables. A total of 75% of all stations were grouped alike for both data sets. Table 38 represents the classification of all clusters, indicating where incorrectly grouped collections were found. Of the stations clustered differently, half were associated with severe storm surges. High winds and excess rainfall may have altered either the environmental or biological characteristics of the various areas of the Lafayette River.

Chapter 6

DISCUSSION

Spatial differences were found between all stations analyzed. The four regions of the Lafayette River were separated by both environmental and biological factors. These areas consisted of the River mouth, mid-river, and each of two branches. Stations located in any of these areas consistently had a characteristic set of variables over time.

The River mouth was defined as an area of high salinities, with measurements near 230/00 being common. Water temperature was low compared to stations upriver. Major nutrient concentrations were highest at Station 1. Reactive silicates had lower concentrations in this region. Characteristic of the mid-river area were lower salinities (19-210/00) and higher temperatures. Nitrogen and phosphorus levels were slightly decreased from concentrations in the mouth of the River. Silicate values were elevated from 10-20 µg Si/1 at the mouth to 20-30 µg Si/1 at Station 4 in the second division of the River. Salinity decreased as sampling continued into both River branches. Values between 16 and 190/00 were common, with the water temperature highest in the branches. High levels of nutrients were

consistently noted at stations in the two branches, indicating input from up river.

These results agree with past studies conducted in the Lafayette River. Montgomery (1972) found high phosphate concentrations at stations in the River mouth and again in the branches. Ranges agreed with those found in the present study. Mid-river had lower concentrations, also indicating inputs from both sources. High values at the mouth were attributed to sewage wastewater mixing with local estuarine waters. The increasing values noted in the branches may have been caused by either industrial waste or runoff containing fertilizer. Neilson and Sturm (1978) also noted that nitrogen and phosphorus concentrations were high in the Lafayette River. However, their nutrient ranges were lower than those in the present study. One reason for this difference could be the sampling period. Neilson and Sturm sampled only once, whereas weekly samplings were taken to obtain a range of values over time in this study. Neilson and Sturm could have missed high values with one time sampling. Neilson (1975) notes long residence times for pollutants in adjacent areas. This condition, combined with a low flushing rate, would concentrate these substances in the River. Since the pollutants noted by Neilson contain both nitrogen and phosphorus, these elements may be accumulating over time, accounting for increased concentrations.

Biologically, the River mouth was characterized by a dominance of diatoms, with low concentrations of

phytoflagellates. At Station 4, mid-river, diatoms remained abundant, with an increasing number of phytoflagellates. In both River branches, phytoflagellates were dominant, with diatoms low in number. Chlorophytes and cyanophytes had high concentration at Stations 9 and 13, the southern and northern branches. In terms of numbers per liter, abundance increased up river. This was mainly due to an increase in numbers of the unidentified nanoplankton component.

Purcell (1973) also noted high diatom concentrations at his station in the mouth of the River. The diatoms preferred cooler, more saline waters. At Purcell's Station No. 2, located between mid-river and the branches, an increase in phytoflagellates was noted, but diatoms remained dominant. The same pattern was observed in this study. Flagellated cells appeared to flourish in warmer, less saline waters. The large amount of species overlap between stations was attributed to the closeness of stations. However, abundance of phytoplankton in terms of numbers was very different in the two studies. This may be due to the different methods used for enumeration of cells. Purcell counted cells at 350%. In this study, random fields at 500% enabled the enumeration of a nanoplankton component not accounted for in previous studies.

Although adjacent areas of the Lafayette River were separated by the cluster analysis, these were not statistically different environmentally. However, the discriminant function analysis did distinguish stations not adjacent

to each other. One reason for this may be a continuum exists. For instance, stations located in the River mouth may be different from those at mid-river, but some overlap occurs. This was particularly evident during periods of increased rainfall and high winds. These factors may have horizontally mixed waters to establish more uniform conditions. Another reason this would be if a parameter not measured in this study effected the area enough to distinguish these stations. In this case, the factors measured separated areas enough to form different clusters, but did not form statistically significant groups. For example, White (1972) divided the River into the same four areas by shoreline contour, eddy currents, and mixing processes. It is possible these parameters may separate areas significantly if taken into consideration.

Although various environmental factors were important in separating clusters, overall separation, as indicated by the Discriminant Function and Pearson Correlation analyses, found salinity the most significant factor distinguishing stations. Temperature was of secondary importance. These two factors varied consistently throughout the sampling period. The River mouth had high salinities and low temperatures as compared to stations up river. These results agree with Platt, Dickie, and Trites (1970) who found a direct correlation between chlorophyll a values and salinity. However, Paasche (1975) found an inverse relationship between salinity and phytoplankton growth. Differences in these results may

be attributed to the measurement of different aspects of phytoplankton dynamics or the fact that other variables may be effecting cells.

Nutrient concentrations were of lesser importance in separating areas of the Lafayette River. Since major nutrients were elevated in concentration throughout the River, these are unlikely to be limiting in the system. With this abundant nutrient supply, the small variations noted in regions may not distinguish stations as well as salinity and temperature.

Stations separated by presence and numbers of phytoplankton corresponded to those separated by environmental parameters. However, all areas of the River were significantly different, including adjacent stations. species important in forming each cluster were determined to be statistically significant in Discriminant Function and Pearson Correlation analyses. These included 18 diatoms, 6 dinoflagellates, 2 euglenoids, 1 chlorophyte, 1 cyanophyte, and 2 other phytoflagellates. Diatoms significant at the a<.05 level were Cylindrotheca closterium, Gyrosigma</pre> fasciola, Navicula sp., Pleurosigma sp., Tabellaria fenestrata, and an unidentified centrales between 20 and 100 µm. Flagellated cells also statistically important at the same a level were Eutreptia lanowii, E. viridis, Prorocentrum minimum, Pyramimonas sp., and Scrippsiella tricoidea. The presence and numbers of these species explain 99.9% of the variation between stations in the

Lafayette River.

In comparing the two sets of completed analyses, stations were classified in the same cluster 75% of the time. During cruises where stations grouped similarly for both environmental and biological parameters, weather conditions were relatively calm. Of the remaining 25%, those stations clustering in different groups, severe weather conditions prevailed. It may be that measured parameters explain phytoplankton population differences under fair weather conditions, while severe storm action, although sporadic, may be of overriding importance. Another reason these stations did not group together could be a lack of distinguishing parameters. Those factors accounted for in the present study may separate these areas most of the time, but other factors not considered may be necessary to explain 100% of the variation between stations. Zooplankton presence and numbers, rainfall, currents, mixing processes, pollutant concentrations, and other factors may account for the remaining variation in the data set.

Although a cause and effect relationship is indicated by comparison, this cannot be shown by the methods used. It appears that a certain set of environmental conditions is associated with specific phytoplankton assemblages, but whether these conditions cause the presence and numbers of cells remains a question. Further study of the Lafayette River system, involving the frequent monitoring of changing environmental and biological conditions, as well as the

incorporation of other variables, may advance our knowledge of this and other similar systems.

During the sampling period, a temporal variation in the data set was expected. This was because samplings began during late summer and continued into early fall. At least two groups, defined by these time frames, seemed apparent (Golub, 1975). However, changes in stations over time were not observed during this study. Initial clustering of both data sets showed no change in either environmental or biological water composition at any one area of the River over time. This may be because the sampling period was not long enough to show such differences. Also, since 1981 was a year of low rainfall, the changing parameters associated with the oncoming autumn season may not have been as easily detected.

Changes in phytoplankton community structure may be influenced by changes in various physical and chemical factors. Although a causative relationship cannot be shown, it is indicated that given a particular set of environmental conditions, the presence of certain cells can be expected. Variations from the relationships seen may be caused by other factors not measured, such as pollutant toxicity or oxygen content of the waters. These potentially alter phytoplankton growth and productivity and warrant further study in the Lafayette River estuarine system.

Chapter 7

CONCLUSION

Areas of the Lafayette River were separated in space by both environmental and biological factors. The River mouth was characterized by high salinities and nutrient concentrations, with low temperatures. Eigh numbers of diatoms were noted, with low phytoflagellate concentrations. Midriver areas had decreased salinity and nutrient concentrations, and increasing temperatures. Although diatoms remained dominant, an increase in flagellated cells was noted. Branches had fairly high nutrient levels and temperatures, with low salinities. The northern Branch had high phytoflagellate concentrations, with an increasing occurrence of chlorophytes. The southern branch also had high numbers of flagellates, with cyanophytes common.

Statistically, stations not adjacent to one another were significantly different in all analyses conducted.

However, adjacent areas of the River were statistically different when considering the phytoplankton data base only.

Factors not accounted for in this study may be necessary to separate stations on the basis of environmental factors alone. It is also possible that a continuum exists along the River, with conditions being different enough to be

associated with different phytoplankton communities.

In comparing results, similar clusters were obtained for 75% of the stations sampled. Stations grouped in different environmental and biological clusters were often associated with increased rainfall and high winds. These factors may be of greater importance than those measured in separating areas of the Lafayette River. It is also possible these storms alter the environmental conditions too rapidly for the phytoplankton population to react accordingly before sampling took place.

Although a cause and effect relationship cannot be shown, it appears that each set of environmental conditions is associated with a certain phytoplankton community structure. Further study is needed to determine if other variables are important to cells within the system.

LITERATURE CITED

- Bainbridge, R. 1953. Studies on the interrelationships of zooplankton and phytoplankton. J. Mar. Biol. Ass. U.K. 32: 385-452.
- Barlow, J. P., C. J. Lorenzen, and R. T. Myren. 1963. Eutrophication of a tidal estuary. Limnol. Oceanogr. 8: 251-262.
- Cupp, E. E. 1943. Marine Plankton Diatoms of the West Coast of North America. University of California Press, Los Angeles. 237 pp.
- Drouet, F. 1973. Revision of the Nostocaceae with Cylindrical Trichomes. Hafner Press, New York. 292 pp.
- Frey, B. E. and L. F. Small. 1980. Effects of micronutrients and major nutrients on natural phytoplankton populations. J. Plankton Res. 2: 1-22.
- Gauch, H. G., Jr. 1973. Cornell Ecology Program 8A: Data screening for species importances martrices. Cornell University. Ithaca, New York. 23 pp.
- Gauch, H. G., Jr. 1979. COMPCLUS -- A fortran program for rapid initial clustering of large data sets. Cornell University. Ithaca, New York. 59 pp.
- Golub, B. M. 1972. Diurnal studies at a station on an urban estuary in Norfolk, Virginia. Masters Thesis. Biological Sciences. Old Dominion University. Norfolk, Virginia.
- Harrison, W. G. and T. Platt. 1980. Variations in assimilation number of coastal marine phytoplankton: Effects of environmental co-variates. J. Plankton Res. 2: 249-260.
- Haury, L. R., J. A. McGowan, and P. H. Wiebe. 1978.

 Patterns and processes in the time-space scales of plankton distributions. In: Spatial Pattern in Plankton Communities, J. H. Steele, ed. Plenum Press, New York. p. 277-328.

- Hodges, L. 1977. Environmental Pollution. Holt, Rinehart, and Winston, New York. 496 pp.
- Hulburt, E. M. 1970. Competition for nutrients by marine phytoplankton in oceanic, coastal, and estuarine regions. Ecology 51: 475-484.
- Incze, L. S. and C. M. Yentsch. 1981. Stable density fronts and dinoflagellate patches in a tidal estuary. Estuarine, Coastal, and Shelf Sci. 13: 547-556.
- Jeffries, H. P. 1962. Environmental characteristics of Raritan Bay, a polluted estuary. Limnol. Oceanogr. 7: 21-31.
- Marshall, H. G. 1966. Diurnal distribution of phytoplankton from a single station at the mouth of the James River. Ohio J. Sci. 66: 253-255.
- Marshall, H. G. 1967a. Plankton in James River estuary, Virginia. I. Phytoplankton in Willoughby Bay and Hampton Roads. Ches. Sci. 8: 90-101.
- Marshall, H. G. 1967b. Plankton in James River estuary, Virginia. II. Phytoplankton in the Elizabeth River. Virginia J. Sci. 18: 105-109.
- Marshall, H. G. 1968. Plankton in James River estuary, Virginia. III. Phytoplankton in the Lafayette and Elizabeth Rivers (western and eastern branches). Castanea 33: 255-258.
- Marshall, H. G. 1980. Seasonal phytoplankton composition in the lower Chesapeake Bay and Old Plantation Creek, Cape Charles, Virginia. Estuaries 3: 207-216.
- Montgomery, J. R. 1972. Determination of nutrient levels and proposed predictive models for phosphate in the Lafayette River, Norfolk, Virginia. Masters Thesis. Oceanography. Old Dominion University. Norfolk, Virginia.
- Mulford, R. A. and J. J. Norcross. 1971. Species composition and abundance of net phytoplankton in Virginia coastal waters, 1963-1964. Ches. Sci. 12: 142-155.
- Neilson, B. J. 1975. A water quality study of the Elizabeth River: The effects of the Army base and Lambert Point STP effluents. Special Report No. 75. Virginia Institute of Marine Science. Gloucester Point, Virginia. 133 pp.

- Neilson, B. J. and S. C. Sturm. 1978. Elizabeth River water quality report. Special Report No. 134. Virginia Institute of Marine Science, Gloucester Point, Virginia. 45 pp.
- Nie, N. H., C. H. Hull, J. G. Jenkins, K. Steinbrenner, and D. H. Bent. 1975. SPSS: Statistical Package for the Social Sciences. McGraw-Hill Book Company, New York. 675 pp.
- Paasche, E. 1975. The influence of salinity on the growth of some plankton diatoms from brackish water. Norw. J. Bot. 22: 209-215.
- Patten, B. C., R. A. Mulford, and J. E. Warinner. 1963.

 An annual phytoplankton cycle in the lower Chesapeake
 Bay. Ches. Sci. 4: 1-20.
- Pingree, R. D., P. M. Holligan, G. T. Margell, and R. N. Head. 1977. The influence of physical stability on spring, summer, and autumn phytoplankton blooms in the Celtic Sea. J. Mar. Biol. Ass. U.K. 56: 845-873.
- Platt, T., L. M. Dickie, and R. W. Trites. 1970. Spatial heterogeneity in a near-shore environment. J. Fish Bd. Canada 27: 1453-1473.
- Pratt, D. M. 1965. The winter-spring diatom flowering in Narragansett Bay. Limnol. Oceanogr. 10: 173-184.
- Prescott, G. W. 1951. Algae of the western Great Lakes area. Cranbrook Institution of Science Bulletin No. 31. 946 pp.
- Purcell, T. W., III. 1973. Phytoplankton succession in the Lafayette River estuary, Norfolk, Virginia. Masters Thesis. Biological Sciences. Old Dominion University. Norfolk, Virginia.
- Raymont, J. E. G. 1980. Plankton and Productivity in the Oceans. Pergamon Press, New York. 489 pp.
- Reimold, R. J. and F. C. Daiber. 1967. Eutrophication of estuarine areas by rainwater. Ches. Sci. 8: 132-133.
- Riley, G. A. 1947. Seasonal fluctuations of the phytoplankton population in New England coastal waters. J. Mar. Res. 6: 114-125.
- Ryther, J. H., C. S. Yentsch, E. M. Hulburt, and R. F. Vaccaro. 1958. The dynamics of a diatom bloom. Biol. Bull. 115: 257-268.

- Schiller, J. 1933. In: Dr. L. Rabenhorsts' Kryptogamen-Flora von Deutschland, Osterreich und der Schweiz 10(3)(1). 617 pp.
- Schiller, J. 1937. In: Dr. L. Rabenhorsts' Kryptogamen-Flora von Deutschland, Osterreich und der Schweiz 10(3)(2). 590 pp.
- Sinclair, M., D. V. Subba Rao, and R. Couture. 1981.
 Phytoplankton temporal distributions in estuaries.
 Oceanologica Acta 4: 239-245.
- Singer, S. B. and E. G. Gauch, Jr. 1979. CONDENSE -Convert data matrices from any ORDIFLEX format into a
 condensed format by samples. Cornell University.
 Ithaca, New York. 7 pp.
- Smayda, T. J. 1958. Biogeographical studies on marine phytoplankton. Oikos 19: 158-191.
- Sokol, R. R. and F. J. Rohlf. 1981. Biometry. W. H. Freeman and Company, San Francisco. 859 pp.
- Steele, J. H. 1974. Spatial heterogeneity and population stability. Nature 248: 83.
- Strickland, J. D. H. and T. R. Parsons. 1977. A practical handbook of seawater analysis. Fisheries Res. Bd. Canada Bull. 167. 310 pp.
- Taft, J. L., A. J. Elliot, and W. R. Taylor. 1977. Box model analysis of Chesapeake Bay ammonium and nitrate fluxes. In: Estuarine Interactions, M. L. Wiley, ed. Academic Press, New York. p. 115-130.
- Thayer, G. W. 1974. Identity and regulation of nutrients limiting phytoplankton production in the shallow estuaries near Beaufort, N.C. Oecologia 14: 75-92.
- Therriault, J-C. and T. Platt. 1978. Spatial heterogeneity of phytoplankton biomass and related factors in the near-shore waters of an exposed coastal embayment. Limnol. Oceanogr. 23: 888-899.
- Therriault, J-C., D. J. Lawrence, and T. Platt. 1978.

 Spatial variability of phytoplankton turnover in relation to physical processes in a coastal environment.

 Limnol. Oceanogr. 23: 900-911.
- Verduin, J. 1962. In: Great Lakes Basin, H. J. Pincus, ed. The Horn-Shafer Company, Baltimore. p. 107-121.

- White, E. G. 1972. A physical hydrographic study of the Lafayette River. Masters Thesis. Oceanography. Old Dominion University. Norfolk, Virginia.
- Williams, R. B. 1972. Nutrient levels and phytoplankton in the estuary. In: Proceedings of the Coastal Marsh and Estuary Management Symposium, R. H. Chabreck, ed. Louisiana State University Press, Baton Rouge. p. 59-89.
- Wolfe, J. S., B. Cunningham, N. Wilkerson, and J. Barnes. 1926. An investigation of the microplankton of the Chesapeake Bay. J. Elisha Mitchell Sci. Soc. 42: 25-54.

TABLES

Physical data from Stations 1, 4, 9, and 13 for all cruise dates. Table 1.

Cruise Date	Tidal Phase	Station	Water Depth (m)	Secchi Depth (m)	Salinity (%00)	Temperature (OC)
17 August 1981	Storm Spring High	1 1 13	1111	0.46 0.15 0.61 0.30	22.85 21.96 20.89 21.12	26.84 27.22 27.58 27.31
24 August 1981	Storm Neap Low	1 9 4 1	. 2 4 4 0	1.22 0.46 0.30 0.15	22.12 19.30 14.23	23.40 23.05 23.14 23.14
31 August 1981	Lower Spring High	1 1 1 3	4.2.2.0 0.0.0	0.46 0.46 0.30 0.23	22.13 21.58 21.01 20.42	25.38 26.80 26.83 27.30
7 September 1981	Higher Neap Low	1 4 13	3.0 1.0	0.76 0.30 0.61 0.15	22.35 22.00 21.02 21.30	25.48 25.57 25.95 26.45
14 September 1981	Higher Spring High	1 13 13	1 3 3 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5	0.91 0.46 0.61 0.30	21.35 20.92 17.52 18.63	26.19 26.19 27.20 26.78

Table 1. (continued)

Cruise Date	Tidal Phase	Station	Water Depth (m)	Secchi Depth (m)	Salinity (%oo)	Temperature (^O C)
21 September 1981	Storm Neap	니 작	4°0	0.76	21.04	21.62
	Low		•	9.	8.5	1.0
		13	•	0.30	9.2	2.0
28 September 1981		1	•	6	1.5	1.5
	Spring	7	3.0	0.30	21.48	21.88
	High	ത	•	7.	0.4	2.5
		13	•	Ď.	0.5	1.8
5 October 1981	Higher	Н	•	.7	1.4	7.5
	Neap	4	2.5	0.30	21.10	17.50
	Low	თ	•	9	9.6	7.5
		13	•	.	9.9	8.1
12 October 1981	Storm	H	•	0.	3.2	6.5
	Spring	4	•	4.	2.7	6.1
	High	6	2.0	0.91	20.75	15.58
		13	•	• 4	0.8	5,3
19 October 1981	Lower	Н	•	3	3.4	5.8
	Neap	か	2.5	0.46	23.08	16.00
	Low	σ	•	9.	2.1	6.3
		13	•	.	2.2	6.3

Table 1. (continued)

Cruise Date	Tidal Phase	Station	Water Depth (m)	Secchi Depth (m)	Salinity (⁰ /00)	Temperature (^O C)
26 October 1981	Lower	1	4.0	1.07	22.39	15.27
	Spring	4	2.5	0.46	21.42	15.35
	High	9	1.5	0.46	14.84	16.30
	_	13	1.5	0.15	12.38	17.15

Table 2. Chemic	Chemical data from	n stations l,	4, 9, and	13 for all	cruise dates.	
Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites ug N/1	Ortho- phosphate ug P/1	Reactive Silicates ug Si/l
17 August 1981	1	3 3 3	117.01 51.94 32.82	210.00 240.00 220.00	508.34 728.59 636.74	22.18 22.79 22.51
	4	351	32.82 28.98 32.94	190.00 190.00 210.00	718.46 741.83 940.26	40.43 36.01 41.05
	ത	351	1.23 0.35 0.28	360.00 260.00 220.00	1308.01 1453.93 1430.59	84.00 80.08 78.40
	13	351	0.28	240.00 160.00 180.00	1465.62 1453.93 1395.56	48.05 52.81 59.36
24 August 1981	H	H 27 E	126.94 120.85 116.28	120.00 120.00 120.00	318.49 389.51 346.89	26.94 29.01 28.34
	4	351	111.72 102.59 111.72	120.00 120.00 120.00	375.32 432.14 460.50	33.71 42.22 41.55
	ெ	H 02 E	111.72 113.25 105.64	440.00 300.00 400.00	460.54 417.94 460.54	49.50 45.08 40.32
	13	H 07 EP	126.94 120.85 113.25	300.00 300.00 260.00	659.37 630.97 630.97	26.26 25.20 24.36

Table 2. (continued)

				Nitrates		
			oni	and trit	ho- pha	Reactive Silicates
Cruise Date	Station	Replicate	ng N/1	μg N/1	μg P/1	ug Si/1
31 August 1981	п	-1	62.2	60.0		1.6
		7	136,72	260.00	240.31	11.03
		ო	41.8	20.0	60.9	0.7
	7	Н	19.6	40.0	40.3	5.1
		7	114.56	160.00	116.37	
		က	24.8	80.0	99.0	5.5
	6	-	2.4	20.0	'n	7.9
		7	97.51	120.00	95	28.84
		က	5.8	20.0	.7	6.5
	13	H	9.2	20.0	37.0	5.9
		7	107.74	120.00	116.37	30.07
		က	00.9	20.0	37.0	9.3
7 September 1981	Н	-	26.9	40.0	511.44	∞.
		7	115.43		8.6	4
		ო	22.3	000	23.4	۲.
	4	- -i	7.0	0	02.5	8.9
		7	101.64	300.00	802.50	18,93
		က	0.9	0.	13.6	0.7
	6	Т	٣,	60.0	418.1	9.0
		7	126.92	260.00	1406.97	19.66
		က	20.0	70.0	440.5	7.7
	13	-1	70.6	20.0	686.7	2.6
		7	189.00	170.00	1697.96	10.53
		m	89.0	20.0	686.7	1.0

Table 2. (continued)

Cruise Date		Station	Replicate	Ammonia uq N/1	Nitrates and Nitrites ug N/1	Ortho- phosphate	Reactive Silicates
14 September 1981	1981	Н	3 2 1	196.62 186.76 191.69	000	80.3 35.8 80.3	· ~ - ~
		4	357	107.81 122.61 102.89	1160.00 1040.00 1270.00	280.39 280.39 280.39	10.36 10.42 9.86
		ത	351	117.68 97.94 88.09	1380.00 1040.00 930.00	502.35 502.35 465.34	11.42 10.42 11.37
		13	H 20 E	83.15 102.89 93.02	930.00 930.00 710.00	502.35 483.85 502.35	15.29 15.23 15.90
21 September]	1981	1	357	226.81 208.84 199.85	630.00 630.00 720.00	518.16 480.19 461.19	19.54 17.69 17.92
		4	406	208.84 202.85 208.84	190.00 360.00 190.00	423.21 461.19 461.19	13.16 14.34 15.68
		on .	H 07 E	178.88 172.89 175.88	280.00 190.00 100.00	347.23 309.26 309.26	39.26 37.74 35.06
		1.3	3 2 1	190.86 193.86 178.88	100.00	347.23 309.26 309.26	27.05 23.30 23.80

Table 2. (continued)

Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate µg P/1	Reactive Silicates µg Si/l
28 September 1981	-1	351	98.84 89.38 89.38	1160.00 1260.00 1160.00	7.00	20.7 20.4 19.7
	4	H 0 E	94.11 84.66 79.93	640.00 530.00 530.00	767.00 767.00 767.00	31.30 33.04 32.48
	თ	351	30.27 27.89 32.62	320.00 320.00 220.00	932.88 767.00 767.00	28.67 28.22 26.71
	13	H 0 E	27.89 25.54 32.62	120.00 320.00 220.00	601.09 767.00 767.00	32.09 28.67 30.07
5 October 1981	ч	H 2 E	92.06 92.06 95.91	1060.00 1060.00 950.00	404.05 438.28 438.28	28.78 25.20 25.87
	4	3 2 1	40.01 40.01 41.94	640.00 740.00 840.00	221.53 221.53 221.53	26.54 24.30 25.31
	თ	354	55.44 57.36 51.58	1900.00 1370.00 1680.00	164.45 164.45 153.05	37.46 37.02 37.02
	13	351	55.44 51.58 57.36	1480.00 1370.00 1480.00	118.82 118.82 118.82	28.06 26.21 25.20

Table 2. (continued)

				Nitrates and	Ortho-	Reactive
Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrites µg N/1	phosphate µg P/1	Silicates ng Si/l
12 October 1981	H	Т	30.0	280.0	6.96	28.00
		7	130.06	1380.00	196.97	29.62
		ო	30.0	480.0	96.9	2.4
	4	٦	0.4	00	08.3	5.6
		7	က	0.0	8	34.72
		ო	ស	00	4.0	4.8
	O	Н	7	610.00	08.	5.4
		7	•	•	108.38	
		ĸ	9.1	20	08.	8.3
	13	-1	8.9	20.	8.3	7.6
		7	90.19	520.00	86.24	40.00
		ო	7.8	20.	8.3	7.4
19 October 1981	H	г	.3	460.0	126.6	2.7
		7	90.19	1380.00	1116.15	35.50
		ო	7.8	460.0	116.1	4.6
	4	П	0.1	70.0	032.0	4.6
		7	90.19	840.00	1042.59	35.34
		m	0.1	70.0	032.0	7.3
	თ	г	7.	60.0	58.5	6.3
		7	94.86	380.00	958.52	38.42
		က	7	0.09	48.0	6.2
	13	- -i	4.8	10.	27.3	~
		7	86.37	310.00	716.81	44.58
		m	4.8	10,	27.3	6.2

Table 2. (continued)

Ortho-Reactive phosphate Silicates ug P/1 ug Si/1	352.04 14.39 352.04 14.28 374.17 15.90	263.44 32.59 241.30 33.54 241.30 29.12	185.91 48.27 208.07 55.16 230.21 59.92	329.90 37.86
Nitrates and Nitrites ug N/1	850.00 850.00 850.00	670.00 670.00 670.00	300.00	300.00
Ammonia µg N/1	147.81 143.84 118.01	121.97 121.97 119.99	145.82 139.86 139.86	181.58
Replicate	3 5 1	3 2 1	351	r-1 (
Station	1	4	ത	13
Cruise Date	26 October 1981			

Table 3. Environmental factors important in distinguishing areas of the River from the Compclus program.

Cluster 1. River Mouth

Ammonia Orthophosphate Nitrates and Nitrites Tidal Phase Salinity

Cluster 3. Southern Branch

Ammonia Nitrates and Nitrites Tidal Phase Orthophosphate Temperature

Cluster 2. Mid-River

Reactive Silicates Salinity Tidal Phase Secchi Depth Temperature

Cluster 4. Northern Branch

Nitrates and Nitrites Orthophosphate Ammonia Tidal Phase Salinity

Table 4. Phytoplankton composition and abundance from Station 1 on 17 August 1981.

	Number Cells per Liter
Bacillariophyceae	
Acanthes sp. Actinoptychus senarius Ehrenberg Amphora sp. Cylindrotheca closterium (Ehrenberg)	7,769 15,537 128
Reimann and Lewin Gyrosigma fasciola (Ehrenberg) Cleve	38,843 128
Leptocylindrus minimus Gran Navicula cancellata Donkin	186,447 23,306
Paralia <u>sulcata</u> (Ehrenberg) Cleve <u>Pleurosigma elongatum</u> W. Smith	512 256
Pleurosigma sp. Rhizosolenia delicatula Cleve	15,537 15,537
Skeletonema costatum (Greville) Cleve Synedra sp.	15,537 256
<u>Thalassionema</u> <u>nitzschioides</u> Hustedt <u>Thalassiosira</u> <u>eccentrica</u> (Ehrenberg) Cleve	
Thalassiosira nana Lohmann Unknown centrales (<20 µm)	1,920 62,149
Unknown pennales #2 (>20 μm)	23,306
Dinophyceae	
Prorocentrum minimum (Pavillard) Schiller	23,306
Chlorophyceae	
Unknown Chlorophyte	15,537
Cyanophyceae	
Oscillatoria erythraea (Ehrenberg) Kutzing	256
Euglenophyceae	
<u>Eutreptia lanowii</u> Steuer <u>Eutreptia viridis</u> Perty	147,604 31,074
Others	
Calycomonas ovalis Wulff Calycomonas wulfii Conrad and Kufferath Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m) Pyramimonas sp.	285,683 25,971 955,540 3,220,429 2,129,628 623,309 467,482
Total Cells per Liter	8,341,012

Table 5. Phytoplankton composition and abundance from Station 1 on 31 August 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Asterionella glacialis Castracane Chaetoceros compressum Lauder Chaetoceros constrictum Gran Chaetoceros gracile Schutt Chaetoceros sp. Coscinodiscus centralis Ehrenberg Coscinodiscus sp. Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Ditylum brightwellii (West) Grunow Eucampia zoodiacus Ehrenberg Gyrosigma fasciola (Ehrenberg) Cleve Leptocylindrus danicus Cleve Leptocylindrus minimus Gran Navicula cancellata Donkin Nitzschia pungens Grunow Paralia sulcata (Ehrenberg) Cleve Plagiogramma staurophorum (Gregory) Heilberg	
Pleurosigma angulatum (Quekett) W. Smith Pleurosigma sp. Rhizosolenia calcar-avis Schultze Rhizosolenia delicatula Cleve Skeletonema costatum (Greville) Cleve Streptotheca thamensis Shrubsole Tabellaria fenestrata (Lyngbye) Kutzing Thalassionema nitzschioides Hustedt Thalassiosira eccentrica (Ehrenberg) Cleve Thalassiosira gravida Cleve Thalassiosira nana Lohmann Unknown centrales (<20 µm) Unknown centrales (>20 µm) Unknown pennales #4 (>20 µm) Unknown pennales #4 (>20 µm)	5,050 256 192 704 313,075 512 256 25,248 64 256 100,992 55,546 64 10,099 15,149
Dinophyceae Ceratium lineatum (Ehrenberg) Cleve Gymnodinium nelsonii Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller	64 5,050 15,149 15,149
Chlorophyceae Unknown Chlorophyte	20,198

Table 5. (continued)

	Number Cells per Liter
Euglenophyceae	
Eutreptia lanowii Steuer	15,149
Others	
Calycomonas ovalis Wulff Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m) Pyramimonas sp.	12,986 212,083 1,688,128 1,064,819 337,626 623,309
Total Cells per Liter	5,153,040

Table 6. Phytoplankton composition and abundance from Station11 on 7 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Acanthes sp. Actinoptychus senarius Ehrenberg Amphora sp. Cylindrotheca closterium (Ehrenberg)	64 31,074 64
Reimann and Lewin Ditylum brightwellii (West) Grunow	50,496 7,769
Gyrosigma fasciola (Ehrenberg) Cleve Leptocylindrus minimus Gran Navicula cancellata Donkin	64 271,902 64
Pleurosigma sp. Skeletonema costatum (Greville) Cleve Streptotheca thamensis Shrubsole	64 2,112 320
Tabellaria fenestrata (Lyngbye) Kutzing Thalassiosira nana Lohmann Unknown centrales (<20 µm) Unknown pennales #2 (>20 µm)	768 62,149 11,653 3,884
Unknown pennales #4 (>20 μm) Unknown pennales #5 (>20 μm)	3,884 11,653
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller	3,884 3,884 7,769
Chlorophyceae	
Unknown Chlorophyte	7,769
Euglenophyceae	
Eutreptia lanowii Steuer Eutreptia viridis Perty	15,537 11,653
Others	
Cryptomonas sp. Green spheres (<3 μm) Green spheres (3-5 μm) Green spheres (5-10 μm) Pyramimonas sp.	295,207 1,506,330 779,136 181,798 129,856
Total Cells per Liter	3,400,807

Table 7. Phytoplankton composition and abundance from Station 1 on 14 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Coscinodiscus centralis Ehrenberg Coscinodiscus marginatus Ehrenberg Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Ditylum brightwellii (West) Grunow Gyrosigma fasciola (Ehrenberg) Cleve Leptocylindrus minimus Gran Navicula cancellata Donkin Plagiogramma staurophorum (Gregory) Heilberg Pleurosigma angulatum (Quekett) W. Smith Pleurosigma sp. Skeletonema costatum (Greville) Cleve Tabellaria fenestrata (Lyngbye) Kutzing Thalassiosira nana Lohmann Unknown centrales (<20 µm) Unknown pennales #2 (>20 µm) Unknown pennales #5 (>20 µm)	20,198 6,733 384 128 20,198 128 128 740,608 6,733 6,733 6,733 384 7,936 384 53,862 67,328 6,733 6,733
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium sp.	128 6,733
Chlorophyceae	
Unknown Chlorophytes	6,733
Euglenophyceae	
Eutreptia lanowii Steuer	20,198
Others	
Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	484,762 3,116,544 1,636,186 441,510 25,971
Total Cells per Liter	6,690,829

Table 8. Phytoplankton composition and abundance from Station 1 on 21 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Chaetoceros gracile Schutt Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Gyrosigma fasciola (Ehrenberg) Cleve Leptocylindrus minimus Gran Navicula cancellata Donkin Nitzschia delicatissima Cleve Nitzschia pungens Grunow Pleurosigma sp. Skeletonema costatum (Greville) Cleve Unknown centrales (<20 μm) Unknown pennales #2 (>20 μm) Unknown pennales #4 (>20 μm) Unknown pennales #4 (>20 μm)	7,481 18,702 11,221 256 93,511 384 321,678 11,221 11,221 7,481 7,481 89,771 14,962 3,740 18,702 18,702 14,962
Dinophyceae	·
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Prorocentrum ovum (Schiller) Dodge	11,221 512 3,740
Euglenophyceae	
Eutreptia viridis Perty	128
Others	
Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m) Pyramimonas sp.	112,213 3,350,285 1,636,186 467,482 103,885
Total Cells per Liter	6,337,128

Table 9. Phytoplankton composition and abundance from Station 1 on 5 October 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Biddulphia aurita (Lyngbye) Brebisson Coscinodiscus sp. Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Ditylum brightwellii (West) Grunow Gyrosigma fasciola (Ehrenberg) Cleve Leptocylindrus minimus Gran Navicula cancellata Donkin Navicula sp. Plagiogramma staurophorum (Gregory) Heilberg Pleurosigma sp. Skeletonema costatum (Greville) Cleve Thalassionema nitzschioides Hustedt Thalassiosira eccentrica (Ehrenberg) Cleve Unknown centrales (<20 µm) Unknown pennales #2 (>20 µm) Unknown pennales #4 (>20 µm) Unknown pennales #6 (>20 µm)	2,405 4,809 2,405 4,873 38,473 192 64 62,519 4,809 128 2,176 12,023 64 33,664 9,618 9,618 9,618
Dinophyceae	
Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum sp.	7,214 4,809 64
Chlorophyceae	
Unknown Chlorophyte	48,091
Euglenophyceae	
Eutreptia lanowii Steuer Eutreptia viridis Perty	4,809 2,405
Others	
Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	293,358 1,973,811 1,207,661 168,813 38,957
Total Cells per Liter	3,957,132

Table 10. Phytoplankton composition and abundance from Station 1 on 26 October 1981.

	mber Cells er Liter
Bacillariophyceae	
Acanthes sp. Amphora sp.	2,295 4,590 100,992 32 4,590 288 11,476 64 512 2,295 160 128
Reimann and Lewin Ditylum brightwellii (West) Grunow Eucampia zoodiacus Ehrenberg Leptocylindrus minimus Gran Navicula cancellata Donkin Plagiogramma staurophorum (Gregory) Heilberg Pleurosigma angulatum (Quekett) W. Smith Pleurosigma sp. Rhaphoneis amphiceros Ehrenberg Rhizosolenia delicatula Cleve Rhizosolenia stolterfothii Peragallo Schroederella delicatula (Peragallo) Pavillard Skeletonema costatum (Greville) Cleve Streptotheca thamensis Shrubsole Tabellaria fenestrata (Lyngbye) Kutzing Thalassionema nitzschioides Hustedt Thalassiosira eccentrica (Ehrenberg) Cleve Unknown centrales (<20 µm) Unknown pennales #2 (>20 µm) Unknown pennales #4 (>20 µm) Unknown pennales #4 (>20 µm)	4,590 288 22,953 128 128 96 2,327 2,295 256 224
Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum compressum (Bailey) Abe Prorocentrum dentatum Stein Prorocentrum micans Ehrenberg Prorocentrum minimum (Pavillard) Schiller Protoperidinium sp.	96 32 32 2,295 2,295 2,295 32

Table 10. (continued)

	Number Cells per Liter
Euglenophyceae	
Eutreptia lanowii Steuer Eutreptia viridis Perty	2,295 2,295
Others	
Calciosolenia granii Schiller Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm)	2,295 201,984 876,528 623,309 19,478
Total Cells per Liter	1,970,291

Table 11. Phytoplankton composition and abundance from Station 4 on 17 August 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Cylindrotheca closterium (Ehrenberg)	512
Reimann and Lewin	20,198
Diploneis crabro Ehrenberg	256
<u>Diploneis crabro</u> Ehrenberg <u>Gyrosigma fasciola</u> (Ehrenberg) Cleve	128
Leptocylindrus minimus Gran	403,968
Paralia sulcata (Ehrenberg) Cleve	1,024
Pleurosigma elongatum W. Smith	256
Pleurosigma sp.	10,099
Skeletonema costatum (Greville) Cleve	4,352
Thalassiosira nana Lohmann	2,304
Unknown centrales (<20 µm)	60,595
Unknown pennales #2 (>20 μm)	20,198
Unknown pennales #6 (>20 μm)	20,198
Dinophyceae	
Amphidinium acutum Lohmann	128
Gymnodinium nelsonii Martin	424,166
Prorocentrum compressum (Bailey) Abe	128
Prorocentrum minimum (Pavillard) Schiller	30,298
Chlorophyceae	
Unknown Chlorophyte	50,496
Euglenophyceae	
Eutreptia lanowii Steuer	20,198
Eutreptia viridis Perty	20,198
Others	
Calycomonas ovalis Wulff	25,971
Calycomonas wulfii Conrad and Kufferath	25,971
Cryptomonas sp.	1,070,515
Green spheres (<3 µm)	5,324,096
Green spheres (3-5 µm)	2,726,976
Green spheres (5-10 µm)	1,792,013
Pyramimonas sp.	259,712
	-
Total Cells per Liter	12,314,954

Table 12. Phytoplankton composition and abundance from Station 4 on 31 August 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Chaetoceros constrictum Gran Cylindrotheca closterium (Ehrenberg)	15,149 512
Reimann and Lewin	60,595
<u>Ditylum brightwellii</u> (West) Grunow Gyrosigma fasciola (Ehrenberg) Cleve	832 5,050
Leptocylindrus minimus Gran	388,819
Navicula sp.	64 g 15,149
Plagiogramma staurophorum (Gregory) Heilberg Pleurosigma angulatum (Quekett) W. Smith	192
Skeletonema costatum (Greville) Cleve	156,538
Streptotheca thamensis Shrubsole	128
Tabellaria fenestrata (Lyngbye) Kutzing Thalassiosira eccentrica (Ehrenberg) Cleve	20,198 64
Unknown centrales (<20 µm)	25,248
Unknown pennales #4 (>20 µm)	15,149
Unknown pennales #5 (>20 μm)	15,149
Dinophyceae	
Amphidinium acutum Lohmann	5,050
Gymnodinium nelsonii Martin	5,050
Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller	5,050 15,149
Scrippsiella tricoidea (Stein) Loeblich III	
Chlorophyceae	
Unknown Chlorophyte	5,050
Euglenophyceae	
Eutreptia lanowii Steuer	40,397
Others	
Cryptomonas sp.	414,067
Green spheres (<3 µm)	2,908,774
Green spheres (3-5 μm) Green spheres (5-10 μm)	1,610,214 662,266
Pyramimonas sp.	1,506,330
	7 016 421
Total Cells per Liter	7,916,431

Table 13. Phytoplankton composition and abundance from Station 4 on 7 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Chaetoceros decipiens Cleve Chaetoceros gracilis Schutt Cylindrotheca closterium (Ehrenberg)	3,607 320 3,607
Reimann and Lewin Ditylum brightwellii (West) Grunow Grammatophora sp. Gyrosigma fasciola (Ehrenberg) Cleve	25,248 192 7,214 64
Leptocylindrus minimus Gran Nitzschia delicatissima Cleve Plagiogramma staurophorum (Gregory) Heilberg	371,506 28,855
Pleurosigma sp. Skeletonema costatum (Greville) Cleve Unknown centrales (<20 µm) Unknown pennales #6 (>20 µm)	256 2,624 3,607 3,607
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum micans Ehrenberg Prorocentrum minimum (Pavillard) Schiller	128 320 10,821 3,607 3,607
Euglenophyceae	
Eutreptia lanowii Steuer	14,427
Others	
Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	248,873 1,207,661 1,999,782 168,813 25,971
Total Cells per Liter	4,141,931

Table 14. Phytoplankton composition and abundance from Station 4 on 14 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Ditylum brightwellii (West) Grunow Leptocylindrus minimus Gran Melosira moniliformis (Muller) Agardh Navicula cancellata Donkin Paralia sulcata (Ehrenberg) Cleve Plagiogramma staurophorum (Gregory) Heilberg Pleurosigma sp. Skeletonema costatum (Greville) Cleve Thalassiosira nana Lohmann Unknown centrales (<20 µm) Unknown pennales #4 (>20 µm) Unknown pennales #4 (>20 µm) Unknown pennales #5 (>20 µm)	128 256 128 50,496 256 504,960 640 8,416 768 16,832 512 5,888 14,208 67,328 128 8,416 16,832
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller	25,248 16,832 16,832 25,248
Euglenophyceae	
Eutreptia lanowii Steuer	42,080
Others	
Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	934,176 4,051,507 2,856,832 337,626 77,914
Total Cells per Liter	9,080,487

Table 15. Phytoplankton composition and abundance from Station 4 on 28 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Acanthes sp. Actinoptychus senarius Ehrenberg Biddulphia aurita (Lyngbye) Brebisson Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Leptocylindrus minimus Gran Navicula cancellata Donkin Paralia sulcata (Ehrenberg) Cleve Plagiogramma staurophorum (Gregory) Heilberg Pleurosigma sp. Skeletonema costatum (Greville) Cleve Thalassiosira nana Lohmann Unknown centrales (<20 µm) Unknown centrales (>20 µm) Unknown pennales #2 (>20 µm) Unknown pennales #4 (>20 µm) Unknown pennales #4 (>20 µm)	10,099 768 128 302,976 333,274 10,099 640 30,298 384 242,381 7,936 60,595 128 10,099 60,595 121,190
Dinophyceae	
Gymnodinium nelsonii Martin Prorocentrum minimum (Pavillard) Schiller	60,595 128
Chlorophyceae	
Unknown Chlorophyte	30,298
Euglenophyceae	
Eutreptia lanowii Steuer	30,298
Others	
Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m)	1,282,598 5,324,096 3,506,112 805,107
Total Cells per Liter	L2,230,822

Table 16. Phytoplankton composition and abundance from Station 4 on 12 October 1981.

	Number Cells per Liter
Bacillariophyceae	
Amphora sp. Chaetoceros constrictum Gran Coscinodiscus sp. Cylindrotheca closterium (Ehrenberg)	15,946 1,152 10,631
Reimann and Lewin Gyrosigma fasciola (Ehrenberg) Cleve	122,253 384
Leptocylindrus minimus Gran	10,631
Nitzschia delicatissima Cleve Plagiogramma staurophorum (Gregory) Heilberg Pleurosigma sp.	10,631 10,631 128
Skeletonema costatum (Greville) Cleve Thalassionema nitzschioides Hustedt	15,946 10,631
Unknown pennales #4 (>20 μm) Unknown pennales #6 (>20 μm)	5,315 15,946
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin	10,631 26,577
Cyanophyceae	
Oscillatoria erythraea (Ehrenberg) Kutzing	5,315
Euglenophyceae	
Eutreptia viridis Perty	10,631
Others	
Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	818,567 4,570,931 2,181,581 363,597 51,942
Total Cells per Liter	8,269,997

Table 17. Phytoplankton composition and abundance from Station 4 on 26 October 1981.

	Number Cells per Liter
Bacillariophyceae	
Acanthes sp. Actinoptychus senarius Ehrenberg Amphora sp. Asterionella glacialis Castracane Bacillaria paxillifer (Muller) Hendey Chaetoceros decipiens Cleve Chaetoceros gracile Schutt Chaetoceros sp. Coscinodiscus lineatus Ehrenberg Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Ditylum brightwellii (West) Grunow Gyrosigma fasciola (Ehrenberg) Cleve Leptocylindrus minimus Gran Paralia sulcata (Ehrenberg) Cleve Plagiogramma staurophorum (Gregory) Heilberg Pleurosigma elongatum W. Smith Rhizosolenia stolterfothii Peragallo Schroederella delicatula (Peragallo) Pavilla Skeletonema costatum (Greville) Cleve Tabellaria fenestrata (Lyngbye) Kutzing Thalassionema nitzschioides Hustedt Thalassiosira eccentrica (Ehrenberg) Cleve	64 64
Unknown centrales (>20 μ m) Unknown pennales #6 (>20 μ m)	2,525 2,525
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum micans Ehrenberg Prorocentrum minimum (Pavillard) Schiller	5,050 7,574 30,298 32 5,050
Cyanophyceae	
Agmenellum thermale (Kutz) Drouet and Daily Oscillatoria erythraea (Ehrenberg) Kutzing	5,248 32
Euglenophyceae	
Eutreptia lanowii Steuer	2,525
Others	
Cryptomonas sp. Green spheres (<3 µm)	416,592 954,442

Table 17. (continued)

	Number Cells per Liter
Green spheres (3-5 μm)	564,874
Green spheres (5-10 µm)	38,957
Pyramimonas sp.	38,957
Total Cells per Liter	2,238,508

Table 18. Phytoplankton composition and abundance from Station 9 on 17 Autust 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Cylindrotheca closterium (Ehrenberg)	30,298 256
Reimann and Lewin Navicula cancellata Donkin Pleurosigma angulatum (Quekett) W. Smith	30,298 10,099 256
Pleurosigma sp. Skeletonema costatum (Greville) Cleve	20,198 4,096
Thalassiosira nana Lohmann Unknown centrales (<20 µm) Unknown pennales #2 (>20 µm)	768 60,595 128
Unknown pennales #5 (>20 μm)	30,298
Dinophyceae	_
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin	20,198 1,020,019
Prorocentrum minimum (Pavillard) Schiller Scrippsiella tricoidea (Stein) Loeblich III	40,397
Chlorophyceae	
Unknown Chlorophyte	20,198
Euglenophyceae	
Eutreptia viridis Perty	40,397
Others	
Calycomonas ovalis Wulff Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	25,971 1,181,606 7,142,080 5,038,413 1,298,560 103,885
Total Cells per Liter	16,139,212

Table 19. Phytoplankton composition and abundance from Station 9 on 31 August 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Chaetoceros constrictum Gran Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Gyrosigma fasciola (Ehrenberg) Cleve Leptocylindrus minimus Gran Navicula cancellata Donkin Navicula sp. Paralia sulcata (Ehrenberg) Cleve Pleurosigma angulatum (Quekett) W. Smith Pleurosigma elongatum W. Smith Skeletonema costatum (Greville) Cleve Thalassiosira nana Lohmann	256 128 640 18,362 128 128,535 9,181 128 896 256 128 10,496 36,724
Unknown centrales (>20 μ m) Unknown pennales #2 (>20 μ m) Unknown pennales #4 (>20 μ m) Unknown pennales #5 (>20 μ m)	18,362 128 18,362 18,362
Dinophyceae	
Ceratium fusus (Ehrenberg) Dujardin Gymnodinium nelsonii Martin Gymnodinium sp Prorocentrum minimum (Pavillard) Schiller Scrippsiella tricoidea (Stein) Loeblich III	
Chlorophyceae	
Crucigenia fenestrata Schmidle Unknown Chlorophyte	9,181 18,362
Euglenophyceae	
Eutreptia lanowii Steuer	55,087
Others	
	51,942 103,885 1,221,085 6,103,232 4,181,363 986,906 986,906
Total Cells per Liter	14,741,179

Table 20. Phytoplankton composition and abundance from Station 9 on 14 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Coscinodiscus sp. Cylindrotheca closterium (Ehrenberg)	20,198 128
Reimann and Lewin	10,099
Leptocylindrus minimus Gran	515,059
Skeletonema costatum (Greville) Cleve	3,840
Unknown pennales #5 (>20 μm)	512
Unknown pennales #6 (>20 μ m)	60,595
Dinophyceae	
Amphidinium acutum Lohmann	100,992
Gymnodinium nelsonii Martin	777,638
Prorocentrum minimum (Pavillard) Schiller	70,694
Scrippsiella tricoidea (Stein) Loeblich III	10,099
Chlorophyceae	
Crucigenia fenestrata Schmidle	20,198
Euglenophyceae	·
Eutreptia lanowii Steuer	40,397
Others	·
Cryptomonas sp.	1,161,408
Green spheres (<3 μm)	7,479,706
Green spheres (3-5 µm)	4,778,701
Green spheres (5-10 μm)	701,222
Pyramimonas sp.	363,597
Total Cells per Liter	16,115,083

Table 21. Phytoplankton composition and abundance from Station 9 on 28 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Coscinodiscus sp. Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Leptocylindrus minimus Gran Navicula cancellata Donkin Pleurosigma sp. Skeletonema costatum (Greville) Cleve Thalassiosira nana Lohmann Unknown pennales #4 (>20 µm)	384 10,099 1,454,285 2,029,939 128 3,072 40,397 70,694 40,397 141,389
Unknown pennales #5 (>20 μm)	141,389
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin	640 777 , 638
Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller	10,099 10,099
Chlorophyceae	
Unknown Chlorophyte	272,678
Euglenophyceae	
Eutreptia lanowii Steuer	10,099
Others	
Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m) Pyramimonas sp.	1,181,606 6,804,454 4,467,046 1,298,560 181,798
Total Cells per Liter	18,805,501

Table 22. Phytoplankton composition and abundance from Station 9 on 5 October 1981.

	Number Cells per Liter
Bacillariophyceae	
Acanthes sp. Actinoptychus senarius Ehrenberg Coscinodiscus lineatus Ehrenberg Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Gyrosigma fasciola (Ehrenberg) Cleve Nitzschia delicatissima Cleve Pleurosigma elongatum W. Smith Pleurosigma sp. Thalassiosira eccentrica (Ehrenberg) Cleve Unknown pennales #6 (>20 µm)	256 128 256 624,314 384 36,724 18,362 384 256 768
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller Protoperidinium sp. Scrippsiella tricoidea (Stein) Loeblich III	82,630 229,527 9,181 55,087 128 9,181
Others	
Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	835,479 3,635,968 2,519,206 805,107 51,942
Total Cells per Liter	8,915,268

Table 23. Phytoplankton composition and abundance from Station 9 on 12 October 1981.

	Number Cells per Liter
Bacillariophyceae	
Acanthes sp. Actinoptychus senarius Ehrenberg Cylindrotheca closterium (Ehrenberg)	5,050 5,050
Reimann and Lewin Gyrosigma fasciola (Ehrenberg) Cleve	287 , 827 64
Leptocylindrus minimus Gran Melosira moniliformis (Muller) Agardh	10,099 192
Navicula cancellata Donkin	5,050 128
Pleurosigma sp. Thalassionema nitzschioides Hustedt	5,050 256
Unknown centrales (>20 μm) Unknown pennales #5 (*20 μm)	5,050 5,050
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Gymnodinium sp. Scrippsiella tricoidea (Stein) Loeblich III	757,440 50,496 10,099 5,050
Chlorophyceae	
Scenedesmus quadricauda (Turpin) Brebisson	20,198
Others	
Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m) Pyramimonas sp.	701,894 2,804,890 1,701,114 194,784 25,971
Total Cells per Liter	5,069,802

Table 24. Phytoplankton composition and abundance from Station 9 on 26 October 1981.

	Number Cells per Liter
Bacillariophyceae	
Acanthes sp. Actinoptychus senarius Ehrenberg Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Gyrosigma fasciola (Ehrenberg) Cleve Navicula forcipata Greville Pleurosigma sp. Rhizosolenia fragilissima Bergon Skeletonema costatum (Greville) Cleve Unknown pennales #2 (>20 µm) Unknown pennales #6 (>20 µm)	128 128 151,488 128 128 20,198 640 30,298 128 20,198
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Gymnodinium sp.	10,099 1,287 20,198
Others	
Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	2,666,189 8,518,554 5,038,413 337,626 155,827
Total Cells per Liter	16,971,655

Table 25. Phytoplankton composition and abundance from Station 13 on 17 August 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Coscinodiscus sp. Cylindrotheca closterium (Ehrenberg)	2,432 256 128
Reimann and Lewin Ditylum brightwellii (West) Grunow	121,190 128
Leptocylindrus minimus Gran Pleurosigma elongatum W. Smith Pleurosigma sp.	40,397 256 512
Skeletonema costatum (Greville) Cleve Thalassionema nitzschioides Hustedt	5,376 256
Thalassiosira nana Lohmann Unknown pennales #4 (>20 μm) Unknown pennales #5 (>20 μm)	40,397 10,099 10,099
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller Scrippsiella tricoidea (Stein) Loeblich III	
Chlorophyceae	
Unknown Chlorophyte	20,198
Euglenophyceae	
Eutreptia lanowii Steuer Eutreptia viridis Perty	10,099 2,304
Others	
Calycomonas ovalis Wulff Calycomonas wulfii Conrad and Kufferath Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	103,885 25,971 1,282,598 8,051,072 4,830,643 1,298,560 51,942
Total Cells per Liter	16,595,544

Table 26. Phytoplankton composition and abundance from Station 13 on 24 August 1981.

	Number Cells per Liter
Bacillariophyceae	
Acanthes sp. Actinoptychus senarius Ehrenberg Amphora sp. Coscinodiscus sp. Cylindrotheca closterium (Ehrenberg) Reimann and Lewin	256 10,099 256 128 30,298
Leptocylindrus minimus Gran Melosira moniliformis (Muller) Agardh Pleurosigma sp. Skeletonema costatum (Greville) Cleve Unknown centrales (<20 µm) Unknown pennales #4 (>20 µm) Unknown pennales #6 (>20 µm)	20,198 512 128 1,024 30,298 20,198 20,198
Dinophyceae	
Gymnodinium nelsonii Martin Prorocentrum dentatum Stein Scrippsiella tricoidea (Stein) Loeblich Dinoflagellate cysts	4,261,862 20,198 III 40,397 30,298
Cyanophyceae	
Oscillatoria splendida Greville	272,678
Euglenophyceae	
Eutreptia viridis Perty	212,083
Others	
Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m) Pyramimonas sp.	575,654 8,570,496 4,467,046 883,021 129,856
Total Cells per Liter	19,597,182

Table 27. Phytoplankton composition and abundance from Station 13 on 31 August 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Coscinodiscus sp. Cylindrotheca closterium (Ehrenberg)	128 7,769 128
Reimann and Lewin Leptocylindrus minimus Gran Nitzschia delicatissima Cleve Pleurosigma sp. Rhaphoneis surirella (Ehrenberg) Grunow	27,190 73,802 15,537 3,884
Skeletonema costatum (Greville) Cleve Thalassionema nitzschioides Hustedt Unknown centrales (>20 µm)	38,843 5,888 384 3,884
Dinophyceae	
Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller	341,819 66,033 7,769
Chlorophyceae	
Unknown Chlorophyte	11,653
Cyanophyceae	
Nostoc commune Vauch	23,306
Euglenophyceae	
Eutreptia lanowii Steuer	11,653
Others	
	77,914 167,025 3,584,026 2,519,206 883,021 15,537
Total Cells per Liter	7,886,399

Table 28. Phytoplankton composition and abundance from Station 13 on 7 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Amphora sp. Cylindrotheca closterium (Ehrenberg)	128
Reimann and Lewin	15,537
Leptocylindrus minimus Gran	31,074
Thalassiosira eccentrica (Ehrenberg) Cleve Unknown centrales (<20 µm)	128 23,306
Unknown pennales #2 (>20 µm)	7,769
Unknown pennales #5 (>20 μm)	7,769
Unknown pennales #6 (>20 μm)	15,537
Dinophyceae	
Amphidinium acutum Lohmann	69,918
Gymnodinium nelsonii Martin	77,686
Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller	15,537 31,074
Frorocentrum minimum (Faviriard) Schiller	31,074
Chlorophyceae	
Crucigenia fenestrata Schmidle	7,769
Unknown Chlorophyte	7,769
Euglenophyceae	
Eutreptia lanowii Steuer	69,918
Others	
Calycomonas wulfii Conrad and Kufferath	25,971
Cryptomonas sp.	1,196,367
Green spheres (<3 µm)	5,220,211 4,467,046
Green spheres (3-5 μm) Green spheres (5-10 μm)	1,116,762
Pyramimonas sp.	857,050
Total Cells per Liter	13,264,326

Table 29. Phytoplankton composition and abundance from Station 13 on 14 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Amphora sp. Cylindrotheca closterium (Ehrenberg)	128
Reimann and Lewin Grammatophora sp. Leptocylindrus minimus Gran Navicula cancellata Donkin Pleurosigma sp. Thalassiosira eccentrica (Ehrenberg) Cleve Thalassiosira nana Lohmann	10,099 20,198 353,472 256 384 128
Unknown centrales (>20 μm) Unknown pennales #5 (>20 μm) Unknown pennales #6 (>20 μm)	10,099 768 10,099
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller	201,984 232,282 10,099 40,397
Chlorophyceae	
Unknown Chlorophyte	10,099
Euglenophyceae	
Eutreptia lanowii Steuer	10,099
Others	
Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	1,141,210 6,155,174 4,077,478 805,107 623,309
Total Cells per Liter	13,714,405

Table 30. Phytoplankton composition and abundance from Station 13 on 21 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Coscinodiscus marginatus Ehrenberg Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Leptocylindrus minimus Gran Pleurosigma sp. Skeletonema costatum (Greville) Cleve Thalassiosira nana Lohmann Unknown pennales #4 (>20 µm) Unknown pennales #5 (>20 µm)	256 128 10,099 10,099 90,893 80,794 896 4,096 3,840 10,099 60,595
Dinophyceae	
Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller Scrippsiella trichoidea (Stein) Loeblich II	2,272,370 10,099 640 II 20,198
Chlorophyceae	
Unknown Chlorophyte	70,694
Euglenophyceae	
Eutreptia lanowii Steuer	70,694
Others	
Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m) Pyramimonas sp.	1,322,995 6,518,771 3,765,824 571,366 207,770
Total Cells per Liter	15,103,216

Table 31. Phytoplankton composition and abundance from Station 13 on 28 September 1981.

	Number Cells per Liter
Bacillariophyceae	
Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Leptocylindrus minimus Gran Navicula sp. Pleurosigma elongatum W. Smith Skeletonema costatum (Greville) Cleve Thalassiosira nana Lohmann Thalassiosira sp. Unknown centrales (<20 µm) Unknown centrales (>20 µm) Unknown pennales #4 (>20 µm) Unknown pennales #5 (>20 µm) Unknown pennales #6 (>20 µm)	1,282,598 3,241,843 128 30,298 141,389 30,298 512 30,298 10,099 20,198 212,083 10,099
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller Prorocentrum sp. Scrippsiella tricoidea (Stein) Loeblich III	80,794 1,413,888 20,198 20,198 20,198 20,198
Chlorophyceae	
Unknown Chlorophyte	90,893
Others	
Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m) Pyramimonas sp.	424,166 4,025,536 3,428,198 1,168,704 233,741
Total Cells per Liter	15,946,555

Table 32. Phytoplankton composition and abundance from Station 13 on 5 October 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Amphora sp. Coscinodiscus marginatus Ehrenberg Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Gyrosigma fasciola (Ehrenberg) Cleve Leptocylindrus minimus Gran Navicula sp. Pleurosigma angulatum (Quekett) W. Smith Pleurosigma sp. Rhizosolenia setigera Brightwell Skeletonema costatum (Greville) Cleve Unknown pennales #4 (>20 µm) Unknown pennales #5 (>20 µm)	10,099 10,099 384 444,365 128 20,198 128 256 10,099 384 20,198 20,198 20,198
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Gymnodinium sp. Prorocentrum minimum (Pavillard) Schiller Scrippsiella tricoidea (Stein) Loeblich III	141,389 595,853 20,198 40,397 10,099
Chlorophyceae	
Unknown Chlorophyte	60,595
Euglenophyceae	
Eutreptia lanowii Steuer	60,595
Others	
Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m) Pyramimonas sp.	1,151,309 5,324,096 3,480,141 493,453 129,856
Total Cells per Liter	12,064,715

Table 33. Phytoplankton composition and abundance from Station 13 on 12 October 1981.

	Number Cells per Liter
Bacillariophyceae	
Actinoptychus senarius Ehrenberg Cylindrotheca closterium (Ehrenberg)	256
Reimann and Lewin	383,770
Leptocylindrus minimus Gran	121,190
Pleurosigma elongatum W. Smith	128
Pleurosigma sp.	896
Skeletonema costatum (Greville) Cleve	512
Unknown pennales (<20 µm)	10,099
Unknown pennales #6 (>20 µm)	10,099
Dinophyceae	
Amphidinium acutum Lohmann	252,480
Gymnodinium nelsonii Martin	60,595
Gymnodinium sp.	30,298
Others	
Cryptomonas sp.	1,141,210
Green spheres (<3 μm)	7,713,446
Green spheres (3-5 μm)	3,843,738
Green spheres (5-10 μm)	779,136
Pyramimonas sp.	129,856
Total Cells per Liter	14,477,709

Table 34. Phytoplankton composition and abundance from Station 13 on 19 October 1981.

	Number Cells per Liter
Bacillariophyceae	
Acanthes sp. Chaetoceros gracile Schutt Coscinodiscus lineatus Ehrenberg Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Ditylum brightwellii (West) Grunow Leptocylindrus minimus Gran Nitzschia delicatissima Cleve Paralia sulcata (Ehrenberg) Cleve Pleurosigma elongatum W. Smith Pleurosigma sp. Skeletonema costatum (Greville) Cleve Unknown centrales (<20 µm) Unknown pennales #6 (>20 µm)	10,099 384 128 252,480 256 80,794 20,198 256 128 10,099 20,198 128 10,099
Dinophyceae	
Amphidinium acutum Lohmann Gymnodinium nelsonii Martin Prorocentrum minimum (Pavillard) Schiller	30,298 20,198 10,099
Others	
Cryptomonas sp. Green spheres (<3 µm) Green spheres (3-5 µm) Green spheres (5-10 µm) Pyramimonas sp.	1,797,658 4,337,190 2,701,005 519,424 25,971
Total Cells per Liter	9,847,090

Table 35. Phytoplankton composition and abundance from Station 13 on 26 October 1981.

	Number Cells per Liter
Bacillariophyceae	
Cylindrotheca closterium (Ehrenberg) Reimann and Lewin Gyrosigma fasciola (Ehrenberg) Cleve Leptocylindrus minimus Gran Navicula cancellata Donkin Nitzschia obtusa W. Smith Nitzschia pungens Grunow Paralia sulcata (Ehrenberg) Cleve Pleurosigma angulatum (Quekett) W. Smith Rhizosolenia hebetata f. semispina (Hensen) Gran Unknown pennales #6 (>20 µm)	158,702 7,214 57,710 256 384 384 7,214 128 28,855
Dinophyceae	
Amphidinium schroederi Schiller Gymnodinium nelsonii Martin Gymnodinium sp.	7,214 28,855 7,214
Cyanophyceae	
Oscillatoria erythraea (Ehrenberg) Kutzing	768
Others	
Cryptomonas sp. Green spheres (<3 μ m) Green spheres (3-5 μ m) Green spheres (5-10 μ m) Pyramimonas sp.	1,183,049 2,662,048 1,796,341 151,499 14,427
Total Cells per Liter	6,112,646

Table 36. Phytoplankton species important in distinguishing areas of the River from the COMPCLUS program.

Cluster l - River Mouth

Cylindrotheca closterium Leptocylindrus minimus Skeletonema costatum Thalassiosira nana Amphidinium acutum Gymnodinium nelsonii Gymnodinium sp. Prorocentrum minimum Scrippsiella tricoidea Unknown Chlorophyte Eutreptia lanowii Calycomonas ovalis Pyramimonas sp.

Cluster 2 - Mid-River

Actinoptychus senarius
Cylindrotheca closterium
Leptocylindrus minimus
Nitzschia delicatissima
Plagiogramma staurophorum
Skeletonema costatum
Unknown centrales (<20 µm)
Unknown pennales #2 (>20 µm)
Amphidinium acutum
Gymnodinium nelsonii
Scrippsiella tricoidea
Unknown Chlorophyte
Calycomonas ovalis
Calycomonas wulfii
Pyramimonas sp.

Cluster 3 - Southern Branch

Actinoptychus senarius Cylindrotheca closterium Leptocylindrus minimus Skeletonema costatum Unknown centrales (<20 µm) Unknown pennales #5 (>20 µm) Amphidinium acutum Gymnodinium nelsonii Gymnodinium sp. Prorocentrum minimum Scrippsiella tricoidea Eutreptia lanowii Calycomonas ovalis Pyramimonas sp.

Cluster 4 - Northern Branch

Acanthes sp.

Cylindrotheca closterium
Leptocylindrus minimus
Plagiogramma staurophorum
Pleurosigma sp.
Skeletonema costatum
Amphidinium acutum
Gymnodinium nelsonii
Gymnodinium sp.
Prorocentrum minimum
Scrippsiella tricoidea
Unknown Chlorophyte
Calycomonas ovalis
Calycomonas wulfii
Pyramimonas sp.

Table 37. Comparison of environmental and biological clusters for Stations 1, 4, 9, and 13.

Cruise Date	Station	Environmental Cluster	Biological Cluster
17 August 1981	1	1	1
	4	2	2
	9	4	2
	13	3	3
24 August 1981	13	3	3
31 August 1981	1	1	1
	4	2	2
	9	4	4
	13	3	3
7 September 1981	1	1	1
	4	1	1
	13	3	3
14 September 1981	1	1	1
	4	2	1
	9	4	2
	13	3	3
21 September 1981	1 13	1 3	1 3
28 September 1981	4	3	1
	9	4	2
	13	2	2
5 October 1981	1	1	1
	9	2	3
	13	2	2
12 October 1981	4	2	1
	9	4	4
	13	4	4
19 October 1981	13	3	4
26 October 1981	1	1	1
	4	2	2
	9	4	4
	13	4	4

Table 38. Summary of cluster classifications for both environmental and biological data sets, indicating percentages belonging to each category. Underlined values indicate correctly classified collections.

Environmental Cluster	Biological Cluster			uster
	1	2	3	4
1	100.0	0.0	0.0	0.0
2	25.0	62.5	12.5	0.0
3	12.5	0.0	75.0	12.5
4	0.0	37.5	0.0	62.5

FIGURES

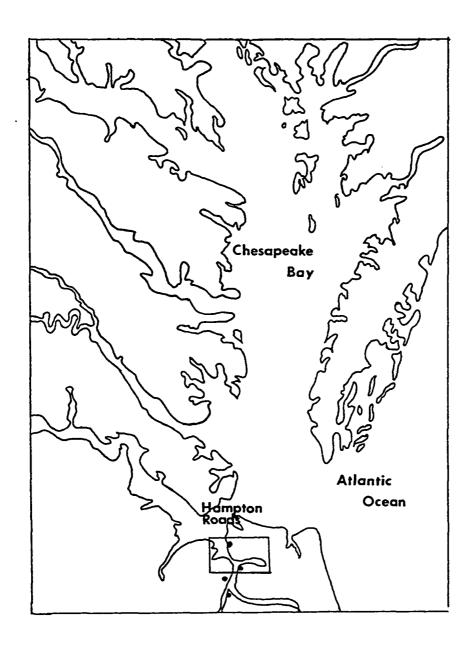


Figure 1. Chesapeake Bay area showing the Lafayette River. Circles indicate locations of sewage treatment plants.

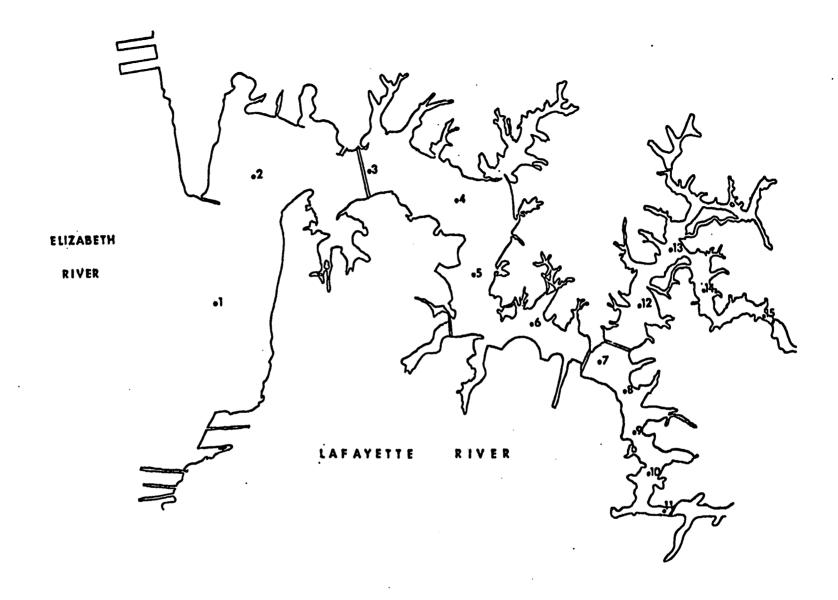


Figure 2. The Lafayette River showing station locations.

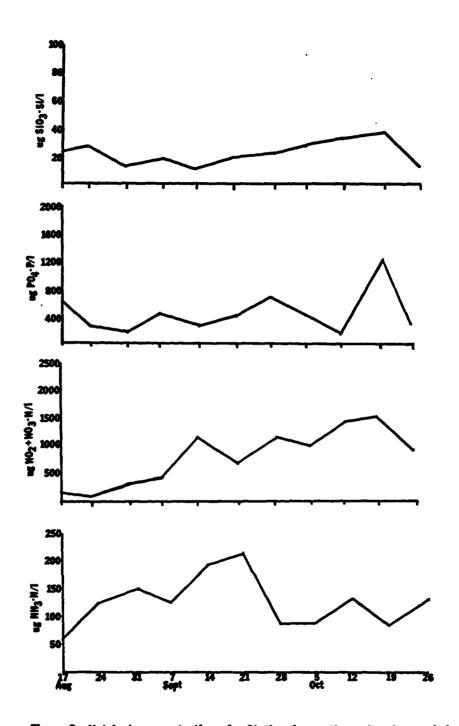


Figure 3. Nutrient concentrations for Station 1 over the collection period.

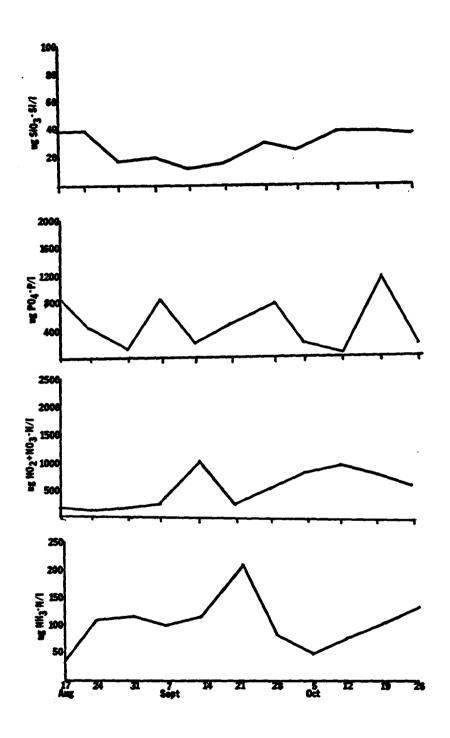


Figure 4. Nutrient concentrations for Station 4 over the collection period.

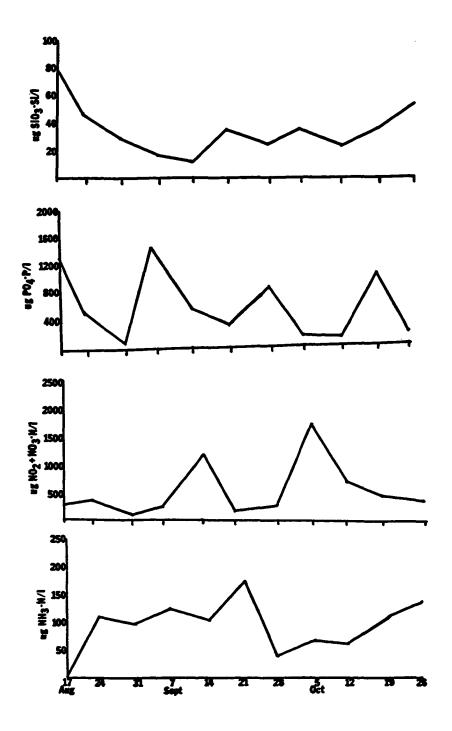


Figure 5. Nutrient concentrations for Station 9 over the collection period.

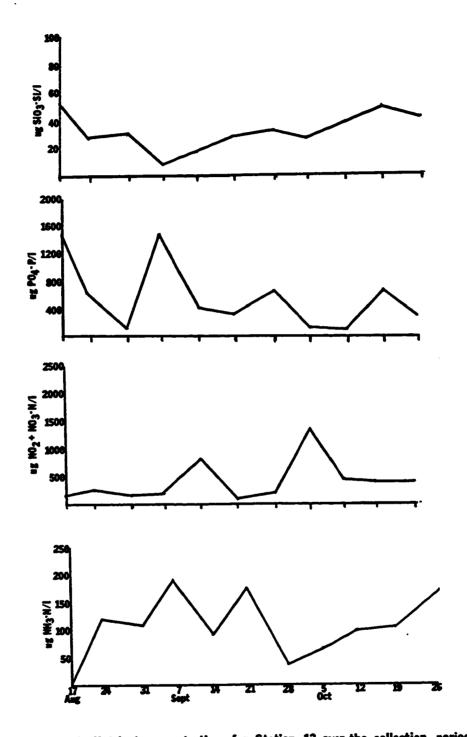


Figure 6. Nutrient concentrations for Station 13 over the collection period.

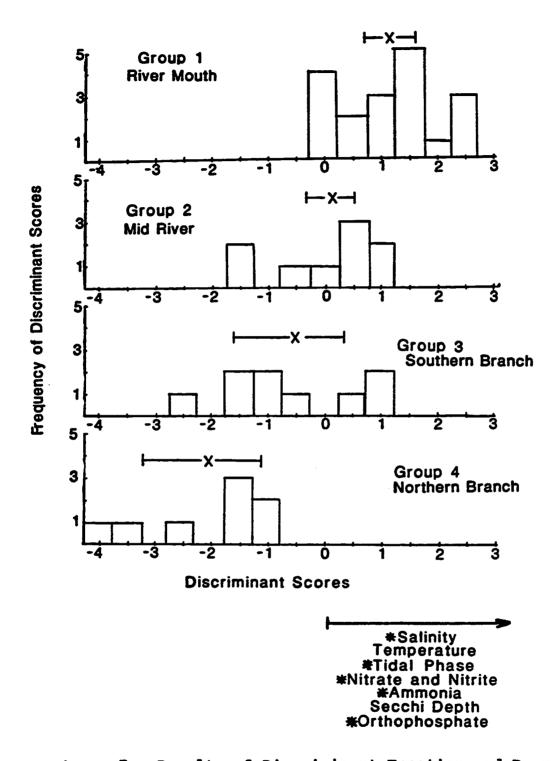


Figure 7. Results of Discriminant Function and Pearson Correlation Analyses on Environmental data.

Arrow indicates a positive relationship between discriminant function scores and original variables.

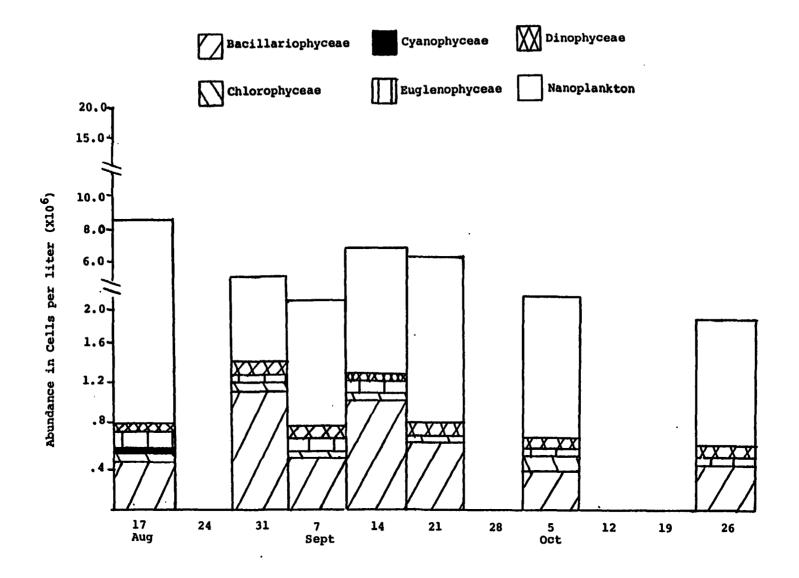


Figure 8. Cell abundances over time for Station 1. Divisions indicate phylogenetic groups.

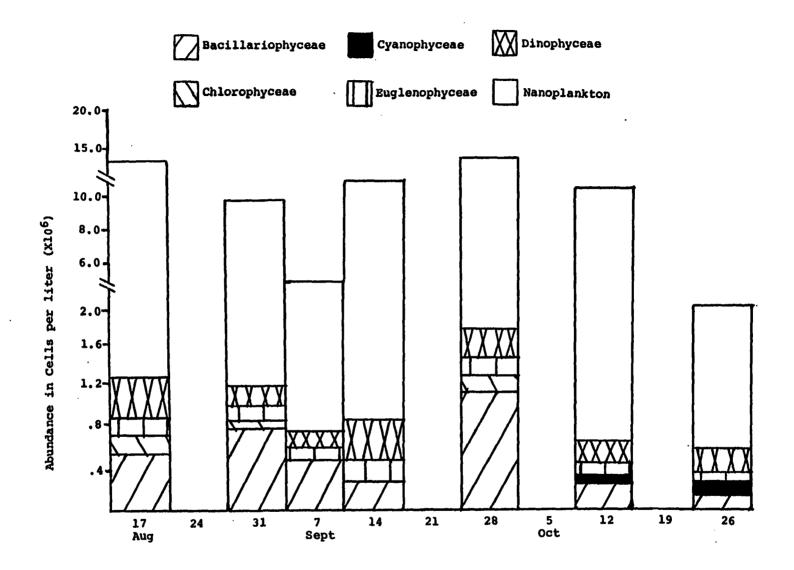


Figure 9. Cell abundances over time for Station 4. Divisions indicate phylogenetic groups.

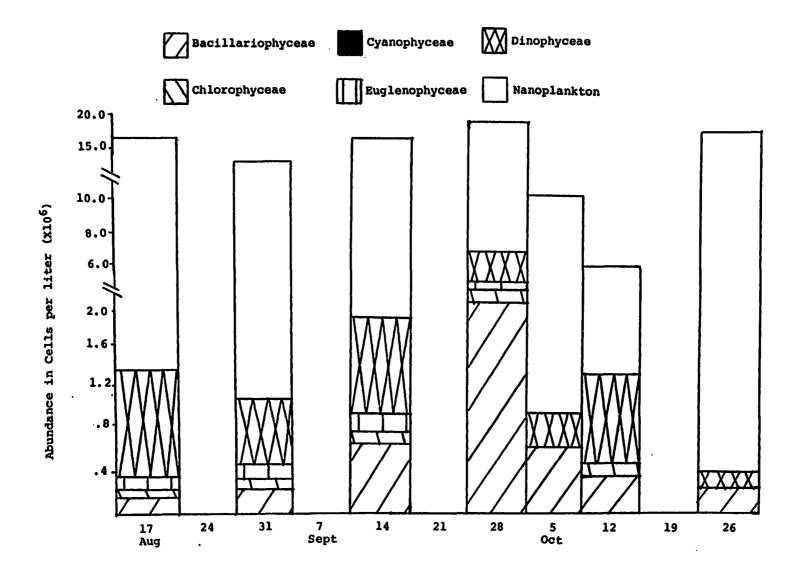


Figure 10. Cell abundances over time for Station 9. Divisions indicate phylogenetic groups.

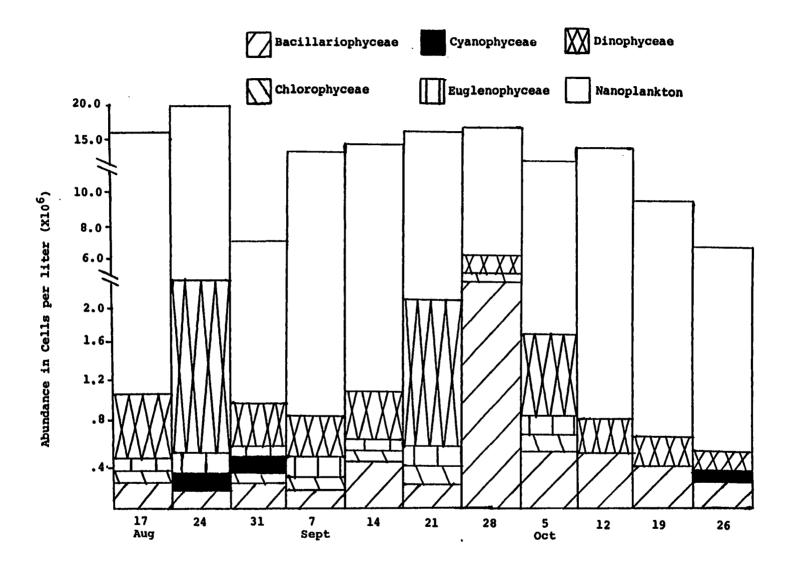


Figure 11. Cell abundances over time for Station 13. Divisions indicate phylogenetic groups.

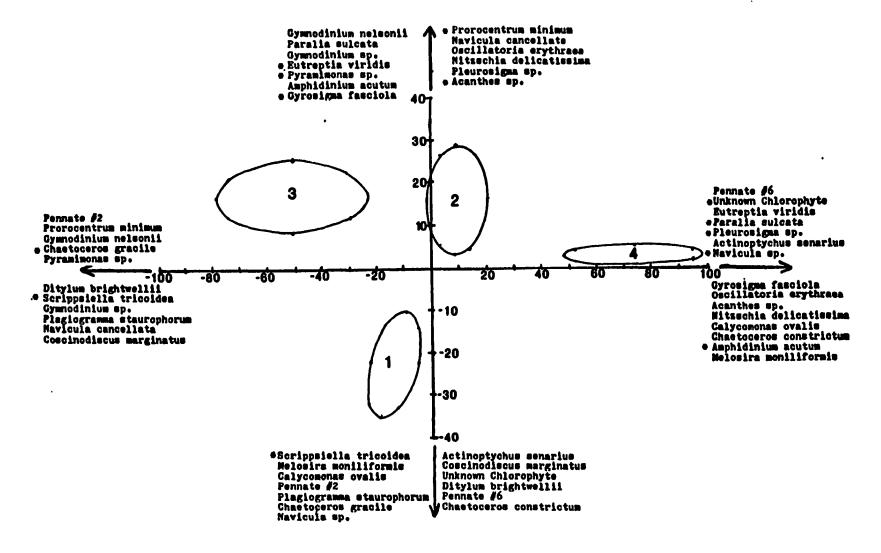


Figure 12. Results of Discriminant Function and Pearson Correlation Analyses on Phytoplankton data. X-axis is the first Discriminant function. Y-axis is the second. Ellipses indicate 95% confidence intervals around group centroids. Arrows indicate phytoplankton populations correlated in the direction of the axes.

APPENDICES

Appendix A. Supportive physical data for all stations, surface and bottom.

Cruise Date	Tidal Phase	Station	Water Depth (m)	Secchi Depth (m)	Salinity (^O /∞)	Temperature (°C)
17 August 1981	Storm	ls	1.5	0.46	22.85	26.84
5	Spring	2 S	1.0	0.46	22.85	26.80
	High	3S	5.0	0.61	22.07	27.12
	_	3B			23.03	26.53
		4 S	0.5	0.30	21.96	27.22
		5S	2.0	0.30	21.65	27.49
		5B			21.84	27.31
		6S	3.0	0.30	21.39	27.69
		6B			21.65	27.25
		7S	3.0	0.46	20.85	27.92
		7B			21.12	27.68
		8S	2.0	0.30	21.20	27.26
		9S	2.0	0.30	20.89	27.58
		10S	2.0	0.30	20.54	27.74
		11S	0.8	0.15	20.15	27.72
		12 S	3.0	0.46	21.18	27.80
		13S	1.0	0.46	21.12	27.31
		14S	1.0	0.30	20.65	26.25
		15S	1.0	0.30	20.71	27.19
24 August 1981	Storm	ls	4.0	1.22	22.12	23.40
-	Neap	1B			22.20	23.42
	Low	2S	3.0	0.91	22.00	23.29
		2B			21.85	23.24
		3S	3.5	0.91	21.02	23.20
		3B			21.28	23.10
		4 S	1.5	0.76	19.30	23.05
		5S	2.0	0.61	17.64	23.62
		5B			20.84	22.84
		6S	3.5	0.46	17.30	22.85

Appendix A. (continued)

Cruise Date	Tidal Phase	Station	Water Depth (m)	Secchi Depth (m)	Salinity (⁰/∞)	Temperature (^O C)
24 August 1981		6B	i	İ	20.18	
(continuea)		ກ ແ	•	4 . ~) °	η γ.ς
		3 S	0.0	0.15	4.2	3. T
		0	•	د .	4.6	3.2
		2	•	٣,	4.7	3.2
		1.38	•	.	4.4	3.1
		4	•	.	2.6	3,3
31 August 1981	Lower	18	•	7	2.1	5.3
	Spring	2S	1.0	1.00	2.0	5.5
	High	38	•	ο.	1.7	6.9
		3B	!!!	 	2.2	5.1
		4S	1.5	0.76	1.5	6.8
		4B	!!!	1 1	2.1	5.4
		58	3.0	0.76	1.3	7.1
		5B	! !	! ! !	1.6	6.1
		89	4.0	0.61	21,13	27.03
		6В	1	1	1.5	6.3
		78	2.0	0.61	1.0	6.2
		7.8	Ī	i	1.0	6.1
		88 8	•	9	1.1	6.8
		98	•	•	1.0	6.8
		108	1.0	0.61	0.7	6.8
		118	•	4.	0.1	7.3
		125	•	9.	0.7	7.0
		138	•	9	0.4	7.3
		14S	•	4.	0.2	7.4
		158	•	₽•	0.4	7.4

Appendix A. (continued)

Cruise Date	Tidal Phase	Station	Water Depth (m)	Secchi Depth (m)	Salinity (%)	Temperature (^O C)
7 September 1981	Higher	1.5	2.5	0.91	2.3	5.4
4	Neap	18	ı į	1	2.3	5.1
	Low	28	1.0	0.76	22.26	25.67
		38	4.5	0.76	2.2	5.6
		3B	1	1 1 1	2.0	5.2
		4s	•	0.61	2.0	5.5
		58	1.5	0.46	1.8	6.0
		5B	!!!	1 1	1.8	5.5
		89	3.5	0.46	1.6	6.1
		6В	1 1	1	1.5	5.5
		78	2.5	0.46	1.5	6.3
		7B	!	1 1 1	1.5	5.9
		88	1.0	•	1.2	6.1
		36		0.30	1.0	5.9
		108	1.0	•	0.9	6.3
		118	•	•	9.0	6.4
		128	•	•	1.5	6.0
		138	•	•	1,3	6.4
		14S	•	•	1.1	9.9
		158	•	0.30	0.9	9•9
14 September 1981		18	3.0	0.76	H	6.1
1	Spring	113	! ! !	1 1	1:	5.7
	High	28	•	•	i.	6.3
	1	38	3.0	1.07	0	6.9
		3B	1 1	1 1	ij	5.4
		45	1.5	1.07	0	6.1
		58	•	1.07	19.91	27.06
		5B	1 1	1 1 1	0	5.9

Appendix A. (continued)

Cruise Date	Tidal Phase	Station	Water Depth (m)	Secchi Depth (m)	Salinity (0/00)	Temperature (^O C)
14 September 1981		6S	•	0.	19.45	27.59
(continued)		6B	1	İ	0.0	7.0
		7S	2.5	0.91	9.2	7.7
		7B	İ	l	8.5	9.9
		88	•	9	9.2	6.4
		98	•	6	7.5	7.2
		0	•	و.	8.4	7.2
			•	9	7.6	6.9
		$^{\circ}$	•	9	9.2	6.5
		138	1.0	0.46	8.6	6.7
		4	•	4.	8.5	6.8
		വ	•	4.	7.9	6.9
21 September 1981	Storm	18	2.5	1.22	1.0	1.6
ì	Neap	1B	!	!!!	9.1	3.0
	Low	28	2.5	1.22	1.1	1.6
		2B	! !	 	6.3	3.0
		38	4.0	1.07	20.50	22.04
		3B	!!	1 1 1	0.1	2.3
		48	•	9	0.5	1.5
		58	1.5	0.91	0.0	1.3
		89	•	.7	9.7	2.3
		6B	1	1	8.2	1.8
		78	3.5	0.76	9.5	2.1
		7B	! ! !	1	6.4	1.8
		88	•	•	9.1	1.3
		98	2.0	0.46	8.5	1.0
		10S	•	• 4	8.5	0.9
		118	•	۴.	7.6	1.6

Appendix A. (Continued)

Cruise Date	Tidal Phase	Station	Water Depth (m)	Secchi Depth (m)	Salinity (0/00)	Temperature (OC)
21 September 1981 (continued)		12S 13S	0.8	0.46	19.45	21.92
		4	•	۳,	9.0	2.0
		ഗ	•	.	9.8	2.0
28 September 1981	Lower	18	3.0	1.22	1.5	1.5
	Spring	1B	!!!	Ì	1.5	1.5
	High	2 S	2.5	0.91	21.55	21.55
	•	2B	! !	1 1 1	1.5	1.5
		38	5.5	1.22	1.5	1.5
		3В	Ĺ		1.5	1.5
		4S	1.0	0.91	1.4	1.8
		58	•	1.22	1.3	2.1
		5B	!!!	ŀ	1.3	1.3
		89	4.5	9	1.0	2.1
		6B	1	ļ	8.3	1.6
		78	2.5	0.91	0.9	2.3
		7B	1	i	1.2	1.7
		88	•	.7	0.9	1.9
		9S	•	9	0.4	2.2
		10S	•	4.	9.0	1.8
		118	•	4.	0.0	2.0
		128	•	.7	0.7	2.0
v-r		138	1.5	0.61	0.5	1.8
		148	•	9	0.2	2.5
		158	•	• 4	0.4	2.4
5 October 1981	Higher	18	2.5	0.91	1.4	7.5
	Neap	118		1	20.00	18.28

Appendix A. (continued)

Cruise Date	Tidal Phase	Station	Water Depth (m)	Secchi Depth (m)	Salinity (0/00)	Temperature (^O C)
5 October 1981	LOW	28	1.0		'ــ ا	7 7
(continued)		38		0.91	21.29	17.44
		3B	1 1	1 1 1	0	8.0
		4S	1.0	0.76	ä	7.5
		58	2.0	0.76	•	7.5
		5B	1 1 1	1 1 1	•	7.3
		89	2.5	0.61	0	8.0
		6B	ı	 	٠ ۵	7.6
		7S	3.5	0.61		7.7
		7B	1 1	! ! !	0	7.8
		88	•	4.	•	7.7
		9S	•	₽•	9	7.5
		108	1.0	0.46	٠ ق	7.6
		118	•	4.	8	8.3
		12S	•	9•		7.6
		138	•	9.	6	8.1
		14S	•	4.	•	8.4
		158	•	. 4	9	8.2
12 October 1981	Storm	18	3.5	1.52	3.2	6.5
	Spring	13	!!	!!!	3.4	6.9
	High	2S	1.0	0.91	3.1	6.4
		38	•	1.52	2.8	6.0
		3B	1 1 1	! ! !	23.41	16.76
		4 S	1.5	1.37	2.7	6.1
		58	•	•	2.3	6.2
		5B	ı	ļ	2.2	6.2
		es S	4.5	1.22	1.9	6.5
		6B	!	t ! !	2.1	6.2

Appendix A. (continued)

			Water	Secchi		
Cruise Date	Tidal Phase	Station	Depth (m)	Depth (m)	Salinity (0/00)	Temperature (^O C)
12 October 1981		7.8	3.5	0.91	21.62	6.1
(continued)		7B	1 1	! ! !	1.8	6.0
		88	•	.7	20.81	15.65
		98	•	9	0.7	5.5
		108	2.5	0.46	0.7	5.6
		118	•	4.	0.8	5.8
		128	•	9•	1,3	5.8
		138	•	4.	0.8	5.3
		148	•	4.	6.0	5.6
		158	•	. 4	0.5	5.1
19 October 1981	Lower	18	4.5	1.52	3.4	5.8
	Neap	1B	1	1 1 1	3,3	5.8
	Low	2S	3.0	1.37	3.5	5.9
		2B	1]	3.5	5.9
		38	4.5	1.07	23.45	15.71
		3B	1	! ! !	3.4	5.8
		4 S	1.5	.7	3.0	6.0
		58	•	0.76	2.7	5.9
		5B	!	! ! !	2.8	6.1
		89	4.5	0.91	2.6	5.9
		6 B	! ! !	!!!	2.6	5.9
		7S	3.5	0.61	2.3	6.0
		7B	!!	1 1 1	2.5	5.9
		88 8	•	.7	2.2	6.1
		98 86	•	.7	2.1	6.3
		108	•	4.	2.0	6.5
		118	1.5	0.46	1.8	6.5
		128	•	7	2.3	6.0

Appendix A. (continued)

Truise Date	Tidal Phase	Station	Water Depth (m)	Secchi Depth (m)	Salinity (0/00)	Temperature (OC)
19 October 1981		138	1.0	0.61	2	ς,
(continued)		14S	1.0	• 6	22.06	9
		158	1.0		1.	• 3
26 October 1981	Lower	18	3.5	1.22	•	5.2
	Spring	118	i	! ! !	•	5.1
	High	28	3.0	1.22	•	5.5
	•	2B	!!!	1 1 1	•	5.4
		38	4.5	1.22	21.56	15,34
		3B	!!!	1 1	•	4.9
		4 S	1.5	0.76	•	5.3
		58	2.0	0.61	•	5.9
		5B	i	1 1	•	5.2
		89	3.0	0.61	•	5.8
		6B	!!!	!!!	•	5.2
		78	2.5	0.76	•	5.9
		7B	!!!	ł ! !	•	5.4
		88	•	0.46	•	5.5
		9S	•	•	•	6.3
		108	•	0.46	•	6.5
		118	1.5	0.30	•	6.7
		128	•	•	•	6.5
		138	•	•	•	7.1
		148	•	۳,	•	7
		158	0.3	0.30	•	

ndix B.	Supportive	chemical	data from	a11	stations,	surface and	and bottom.
se Date	Station		Replicate		Ammonia µg N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
ugust 1981	ч	മ	H 03 FB		117.01 51.94 32.82	210.00 210.00 190.00	508.34 728.59 636.74
	28	മ	H 07 E		27.08 9.84 19.40	240.00 240.00 240.00	555.52 846.86 683.43
	38	ω	H 07 EC		23.24 42.38 13.66	280.00 280.00 280.00	730.14 741.83 800.17
	3B	æ	357		27.08 40.46 17.50	260.00 280.00 240.00	566.71 648.43 683.46
	48	Ø	357		32.82 28.98 32.94	240.00 310.00 310.00	718.46 741.83 940.26
	ŭ	മ	3 2 1		28.98 19.42 21.34	280.00 240.00 240.00	963.60 893.58 788.52
	5B	m	3 2 1		32.82 17.50 15.60	260.00 240.00 220.00	846.89 963.60 858.58

Appendix B. (con	(continued)				
Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites uq N/1	Ortho- phosphate
17 August 1981 (continued)	89	321	30.90 21.32 25.16	10.0	1220.44 1267.16 1313.84
	6 B	351	21.34 17.50 19.40	210.00 210.00 170.00	1173.75 1173.75 1115.38
	78	351	28.98 11.76 19.40	190.00 180.00 180.00	1430.59 1360.53 1395.56
	7B	351	30.90 21.34 17.50	170.00 180.00 210.00	1407.21 1383.87 1453.93
	88 8	H 20 E	6.97 3.14 8.89	170.00 220.00 190.00	1389.70 1407.21 1407.21
	86	3 2 1	1.23 0.35 0.28	170.00 170.00 170.00	1308.01 1453.93 1430.59
	108	321	3.15 4.10	170.00 170.00 170.00	1488.93 1518.13 1506.44
	118	H 0.6	4.10	170.00	1734.11

phosphate 1488.93 1850.82 1757.45 1757.45 1909.23 1909.23 318.49 389.52 346.89 403.71 460.54 488.93 460.54 Ortho-1465.62 1453.93 1442.24 1395.50 1395.56 1407.21 ug P/1 Nitrates Nitrites 510.00 540.00 560.00 540.00 610.00 510.00 170.00 190.00 170.00 160.00 160.00 190.00 170.00 540.00 540.00 620.00 170.00 160.00 170.00 140.00 ug N/1 and 0.28 0.28 126.94 120.85 110.21 108.68 111.72 114.77 4.10 4.10 4.10 0.28 0.28 0.28 Ammonia 107.16 116.28 ug N/1 Replicate Station 1s138 **14S** 158 **1B** 28 (continued) 17 August 1981 (continued) 24 August 1981 Appendix B. Cruise Date

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia ug N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
24 August 1981 (continued)	2B	321	113.25 122.36 113.25	520.00 440.00 510.00	432.14 460.54 446.34
	38	3 5 H	110.21 114.77 111.72	460.00 440.00 420.00	531.56 531.56 559.98
	38	H 02 E	113.25 120.85 102.56	370.00 420.00 380.00	517.36 574.18 545.79
	4S	. H 70 E	111.72 102.59 111.72	320.00 360.00 320.00	375.32 432.14 460.50
·	5.S	H 20 E	119.34 119.34 113.25	240.00 260.00 280.00	446.34 375.29 446.34
	5B	H 00 E	105.64 107.16 107.16	310.00 300.00 300.00	460.54 517.36 517.36
	s9	327	110.21 119.34 116.28	300.00 310.00 320.00	559.98 517.36 488.93

659.37 630.97 630.97

190.00

126.94 120.85 113.25

403.71 531.56 531.56

210.00 220.00 190.00

> 116.28 113.25

12S

138

Nitrates Nitrites 240.00 260.00 240.00 220.00 210.00 210.00 210.00 220.00 220.00 320.00 300.00 210.00 ug N/1 116.28 116.28 122.37 105.64 113.25 110.21 114.77 111.72 Ammonia 113.25 119.34 101.08 105.64 ng N/1 Replicate 327 Station 10s(continued) 24 August 1981 Appendix B. Cruise Date (continued)

phosphate

µg P/1

Ortho-

503.13

446.34

460.54 488.93 474.73 488.93 545.75 503.13 460.54 417.94 460.54 403.71 389.51 432.14

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Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia ug N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
24 August 1981 (continued)	148	H 27 E	113.25 114.77 108.68	190.00 210.00 210.00	716.22 702.03 616.78
31 August 1981	18	351	162.29 136.72 141.85	360.00 260.00 220.00	178.37 240.31 260.99
	2S	321	126.50 136.72 138.43	330.00 220.00 240.00	178.37 178.37 157.73
	38	357	150.36 131.60 128.20	200.00 320.00 220.00	219.67 157.73 199.02
	3B	355	128.20 136.72 129.91	270.00 180.00 180.00	157.73 219.67 199.02
	48	351	119.67 114.56 124.80	240.00 160.00 180.00	240.31 116.37 199.02
	4B	H 27 °	123.09	200.00	178.37 219.67

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate µg P/1
31 August 1981 (continued)	55	3 2 1	117.96 117.96 109.45		178.37 178.37 157.70
	5B	426	95.82 99.22 100.91	180.00 120.00 150.00	137.05 178.37 178.37
	89	H 27 E	102.62 106.04 95.82	180.00 180.00 180.00	116.37 116.37 95.73
	6B	975	107.74 102.62 111.15	120.00 120.00 120.00	199.33 219.67 219.67
	7S	H 27 E	107.74 104.33 100.93	120.00 120.00 120.00	219.67 178.37 178.37
	7B	H 27 E	109.45 102.62 94.17	120.00 120.00 120.00	260.99 178.44 178.44
	8 8	H 27 m	104.33	120.00	137.05

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia ng N/1	Nitrates and Nitrites	Ortho- phosphate
31 August 1981 (continued)	1 🔿	3 2 3	92.40 97.51 95.82	000	16. 95.
	108	3 2 1	87.28 97.51 100.93	120.00 120.00 120.00	95.73 95.73 137.05
	118	3 2 1	119.67 116.27 107.74	120.00 120.00 120.00	75.08 75.08 54.44
	12S	3 2 1	104.33 95.82 97.51	120.00 120.00 120.00	157.73 95.73 95.73
	138	3 2 1	99.22 107.74 100.91	120.00 120.00 120.00	137.05 116.37 137.05
	14S	357	114.56 114.56 128.20	120.00 120.00 120.00	302.31 302.31 343.60
	158	H 02 FI	114.56 121.38 117.96	120.00	219.67 178.37 199.02

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites yg N/1	Ortho- phosphate ug P/1
7 September 1981	1.5	351	126.92 115.43 122.33	440.00 300.00 400.00	511.44 589.81 623.38
	118	H 21 E	117.74 117.74 113.13	350.00 350.00 400.00	679.36 690.56 668.17
	2S	H 26 E	101.64 110.84 106.23	400.00 350.00 350.00	656.98 724.13 724.13
	38	H 24 E	115.43 124.63 106.23	350.00 400.00 350.00	724.13 768.92 780.08
	. 3B	351	94.75 115.44 106.23	400.00 350.00 260.00	813.69 858.45 858.45
	4S	351	97.03 101.64 80.95	300.00 300.00 260.00	802.50 802.50 813.69
	ភន	H 0 F	87.84 87.84 92.44	440.00 260.00 260.00	970.39 959.20 981.58

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia ug N/1	Nitrates and Nitrites ug N/1	Ortho- phosphate ug P/1
7 September 1981 (continued)	5B	3 2 1	110.84 106.23 106.23	220.00 260.00 170.00	1048.73 1104.72 1071.11
	89	351	99.34 108.53 99.34	260.00 220.00 260.00	1048.73 1071.14 1127.10
	6 B	357	101.64 92.44 97.03	220.00 170.00 120.00	1149.48 1194.27 1194.24
	78	H 27 E	108.53 110.84 110.84	170.00 170.00 170.00	1160.67 1194.27 1205.47
	78	351	113.13 113.13 110.82	220.00 260.00 170.00	1306.22 1283.83 1283.80
	88	H 07 EP	115.43 113.12 120.04	220.00 260.00 170.00	1339.76 1373.36 1339.76
	S6	H 02 FI	122.33 126.92 120.04	260.00 260.00 170.00	1418.13 1406.97 1440.51

Appendix B. (continued	inued)				
Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites	Ortho- phosphate
7 September 1981 (continued)	108	321	133.83 133.83 143.02	000	1541.26 1552.48 1518.88
	118	H 27 E	129.22 143.02 136.12	170.00 80.00 80.00	1563.64 1563.64 1563.64
	12S	321	124.63 124.63 126.92	80.00 80.00 80.00	1642.01 1630.82 1642.01
	138	321	170.60 189.00 189.00	120.00 170.00 120.00	1686.77 1697.96 1686.77
	14S	351	267.16 267.16 269.47	80.00 80.00 80.00	1776.33 1776.33 1731.54
	158	3.2.1	271.77 278.66 271.77	80.00 80.00 80.00	1440.51 1451.73 1440.51
14 September 1981	18	3 2 1	196.62 186.76 191.69	1270.00 1160.00 1160.00	280.39 335.88 280.39
	1.8	-12 6	167.02 142.63 147.28	1270.00 1380.00 1270.00	354.39 280.39 354.39

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites ug N/1	Ortho- phosphate ug P/1
14 September 1981 (continued)	2S	-1 O E	152.21 147.28 132.48	1380.00 1380.00 1380.00	280.39 354.39 354.39
	38	- 1 2 E	107.81 68.35 93.02	1160.00 1270.00 1160.00	280.39 354.39 280.39
	3B	126	122.61 152.21 152.22	1160.00 1160.00 930.00	317.38 335.88 372.87
	48	428	107.81 122.61 102.89	1160.00 1040.00 1270.00	280.39 280.39 280.39
	ន	-1 2 E	122.61 122.61 117.68	1160.00 1270.00 1270.00	317.38 280.39 335.88
	5B	321	122.61 142.35 132.48	1160.00 1270.00 1270.00	372.87 354.39 391.37
	89	പ ପ ന	102.89 93.02 93.02	1160.00 1040.00 930.00	372.87 354.39 354.39

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites ug N/l	Ortho- phosphate ug P/1
14 September 1981 (continued)	6В	3 2 1	122.61 122.61 137.42	1160.00 1040.00 1270.00	391.37 317.38 391.37
	78	3.2.1	112.76 132.48 112.76	1270.00 1270.00 1160.00	317.38 391.37 391.37
	7B	3.2.1	112.76 122.61 93.02	1040.00 1040.00 820.00	317.38 391.37 317.38
	88	3.2.1	112.76 122.61 122.61	1270.00 1270.00 1270.00	428.36 465.34 446.83
	86	3 2 1	117.68 97.94 88.09	1380.00 1040.00 930.00	502.35 502.35 465.34
	108	H 02 E	132.48 142.35 132.48	930.00 930.00 1040.00	428.36 465.34 465.34
	118	H 02 FF	147.28 142.35 132.48	930.00 820.00 930.00	502.35 465.34 465.34

537.17 499.16 480.19

280.00 190.00 280.00

193.86 184.87 169.90

518.16 480.19 461.19

630.00

557.81 557.81 576.32

480.19 480.19 499.16

450.00 450.00

187.88 193.86 190.86

1B

28

540.00

Nitrates Nitrites 1040.00 930.00 930.00 710.00 820.00 930.00 600.00 630.00 720.00 820.00 710.00 ug N/1 191.69 176.88 93.02 78.20 83.16 83.15 226.81 208.84 199.85 Ammonia ug N/1 122.61 102.89 132.48 83,15 191.69 Replicate Station 13 **12**S 138 14S 158 (continued) 14 September 1981 September 1981 Appendix B. Cruise Date (continued)

phosphate

µg P/1

Ortho-

and

483.85 502.35 502.35

930.00

502.35

502.35

539.34 557.81 576.32

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Appendix B. (continued)

se Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
eptember 1981 tinued)	2B	H 20 E	181.87 160.90 160.92	280.00 450.00 360.00	442.18 423.21 423.21
	38	୮୯୯	154.92 160.92 151.93	450.00 450.00 280.00	404.21 404.21 442.18
	3B	H 27 E	166.91 172.89 160.92	360.00 360.00 450.00	461.19 480.19 461.19
	4S	3.2.1	208.84 202.85 208.84	190.00 360.00 190.00	423.21 461.19 461.19
	5 8	321	178.89 160.92 157.92	100.00 190.00 190.00	423.21 442.18 423.21
	8 9	351	172.89 169.90 172.89	100.00	423.21 423.21 423.21
	6В	-1 2%	142.92 148.93	100.00	385.21 385.21 423.21

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
21 September 1981 (continued)	7.8	351	148.93 148.93 148.93	360.00 450.00 630.00	366.23 347.23 347.23
	7B	351	160.90 160.90 160.90	720.00 720.00 630.00	404.21 366.23 385.21
	88 8	351	172.89 172.89 172.89	540.00 450.00 280.00	385.21 423.21 347.23
	S6	H 21 E	178.88 172.89 175.88	280.00 190.00 100.00	347.23 309.26 309.26
	108	351	184.87 184.87 184.87	190.00 100.00 280.00	252.28 214.27 233.27
	118	-1 2 6	181.87 193.86 184.87	100.00	214.27 233.27 233.27
	128	H 01 m	181.87 172.89 166.89	100.00	385.21 385.21 366.23

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
21 September 1981 (continued)	1 138	3 2 1	190.86 193.86 178.88	100.00 100.00 100.00	347.23 309.26 309.26
	148	3 2 1	196.85 196.85 190.86	100.00 100.00 100.00	309.26 271.25 271.25
	158	321	208.84 202.85 208.84	100.00 100.00 100.00	271.25 309.26 309.26
28 September 1981	1 18	3 2 H	98.84 89.38 89.38	1160.00 1260.00 1160.00	767.00 767.00 767.00
	18	3 2 1	84.64 87.02 79.93	1160.00 950.00 1060.00	932.88 932.88 849.96
	28	351	53.91 42.08 42.08	950.00 1160.00 950.00	932.88 932.88 767.00
	2B	H 07 K	51.55 39.72 51.55	950.00 830.00 740.00	849.90 849.90 932.88

767.00 684.05 767.00

27.89 18.42 13.69

321

eb

phosphate 849.90 1015.81 932.88 684.05 684.05 684.05 849.90 849.90 932.88 684.05 684.05 849.90 767.00 849.90 767.00 Orthoµg P/1 Nitrates Nitrites 740.00 830.00 530.00 530.00 640.00 530.00 530.00 420.00 530.00 530.00 640.00 530.00 530.00 530.00 ng N/1 830.00 530.00 640.00 and 51.55 42.08 51.55 49.18 51.55 51.55 56.28 84.6679.93 37.35 37.35 32.62 13.71 18.42 13.71 Ammonia 44.45 56.28 ng N/1 Replicate Station 38 3B 48 55 **5B 6**S (continued) 28 September 1981 Appendix B. Cruise Date (continued)

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Appendix B.	(cont	(continued)				
Cruise Date		Station	Replicate	Ammonia ug N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
28 September (continued)	1981	7.8	3 2 1	23.17 25.54 27.89	530.00 420.00 420.00	684.05 601.09 601.09
		7B	351	27.89 27.89 27.89	530.00 420.00 320.00	684.05 601.09 601.09
		88 8	351	8.97 8.97 8.97	320.00 420.00 530.00	601.09 684.05 601.09
		86	3 2 1	30.27 27.89 32.62	320.00 320.00 220.00	932.88 767.00 767.00
		108	3 2 1	23.16 23.16 30.24	120.00 120.00 320.00	601.09 601.09 601.09
		118	351	30.25 23.16 23.16	220.00 120.00 320.00	767.00 767.00 849.90
		128	H 02 FF	35.00 35.00 35.00	420.00 320.00 320.00	849.90 767.00 767.00

phosphate 932.88 932.88 932.88 438.28 438.28 267.16 301.38 278.57 601.09 767.00 767.00 767.00 767.00 767.00 358.42 392.65 381.34 438.28 438.28 104.05 Ortho-126.87 µg P/1 Nitrates Nitrites 1060.00 1060.00 950.00 120.00 320.00 220.00 220.00 320.00 220.00 320.00 120.00 950.00 950.00 740.00 640.00 840.00 840.00 740.00 120.00 1/N gu and 25.54 27.90 27.90 92.06 92.06 95.91 61.22 25.54 25.54 27.89 61.22 55.44 49.66 27.89 25.54 32.62 47.73 41.94 41.94 Ammonia nd N/1 Replicate 32 Station **1B** 38 158 138 14S 13 (continued) 28 September 1981 5 October 1981 Cruise Date Appendix B. (continued)

phosphate 210.12 221.53 232.93 175.86 187.27 187.27 301.38 267.16 244.34 221.53 221.53 221.53 175.86 175.86 175.86 198.68 221.53 221.53 198.68 175.86 187.27 Ortho-Nitrates Nitrites 740.00 740.00 950.00 1060.00 1060.00 740.00 840.00 950.00 640.00 740.00 840.00 1160.00 950.00 1160.00 950.00 µg N/1 950.00 1060.00 1160.00 and 49.66 51.58 43.88 45.81 40.01 41.94 45.79 40.01 41.94 Ammonia 40.01 45.81 40.01 40.01 38.08 41.94 40.01 ud N/1 Replicate 327 Station **6**B 78 58 **5B 89** (continued) 5 October 1981 Appendix B. Cruise Date (continued)

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites ug N/1	Ortho- phosphate ug P/1
5 October 1981 (continued)	7B	3 2 1	40.01 43.88 43.88	1060.00 1060.00 1160.00	198.68 198.68 198.68
	88	3 2 1	47.73 47.73 47.73	1260.00 1260.00 1160.00	175.86 187.27 187.27
	S6	3.2.1	55.44 57.36 51.58	1900.00 1370.00 1680.00	164.45 164.45 153.05
	108	321	59.29 59.29 57.37	1580.00 1580.00 1580.00	130.23 153.06 130.23
	118	3 2 1	59.29 55.44 61.22	1580.00 1580.00 1580.00	118.82 118.82 118.82
	12S	H 20 E	47.73 45.81 43.88	1160.00 1260.00 1370.00	175.86 175.86 153.05
	138	H 02 K	55.44 51.58	1480.00	118.82

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia uq N/1	Nitrates and Nitrites uq N/1	Ortho- phosphate
5 October 1981	148	1	9.2	0	73.19
(continued)		0 M	61.22 57.36	1790.00 1580.00	73.19 73.19
	158	н (3.1	580.	1.7
		N 10	69.29 61.22	1580.00	61.78
12 October 1981	18	ч	30.0	•	96.9
		N M	130.06 130.06	1380.00 1480.00	196.97 196.97
	18	1	32.8	280.	96.9
		0.0	115.86	1380.00	174.84
		m	T0.T	380.	52.6
	28	1	15.8	380.	52.6
		0 1 W	117.29 112.62	1280.00 1280.00	152.67 108.38
	38	H	4		4.0
		7	98	1000.00	64.
		m	. 5	•	ლ დ
	3B	Н	.3	•	4
		7 1 M	107.35	1000.00	64.08 64.08

Appendix B. (continued)

70.46 900.0 73.30 800.0 77.56 800.0 64.79 900.0 64.79 800.0 64.79 800.0 61.95 800.0 61.95 800.0 61.95 800.0 81.82 800.0 61.95 900.0 81.82 800.0 81.82 800.0 61.95 900.0	Ammonia		Ortho- phosphate
1981 45 1 70.46 900 2 73.30 800 3 77.56 800 5 2 64.79 900 5 3 77.36 800 5 8 1 50.60 800 6 8 1 50.60 800 6 8 1 50.60 800 6 8 1 50.60 610 6 8 1 50.60 610 7 8 1 80.39 700		/N bd	1/4 Bd
1 50.60 710 2 64.79 900 3 77.36 800 2 64.79 800 3 64.79 800 3 64.79 800 3 61.95 800 3 61.95 900 2 81.82 800 3 61.95 700 2 39.27 700 2 39.27 700 2 84.91 700 2 84.66 610	70	46 900.	108.38
3 77.56 800 2 64.79 900 3 77.36 800 2 64.79 800 2 64.79 800 3 64.79 800 2 81.82 800 2 81.82 800 3 61.95 900 2 39.27 700 2 39.27 700 2 84.66 610	73	30 800.	r,
1 50.60 710 2 64.79 900 1 50.60 800 2 64.79 800 3 64.79 800 2 81.82 800 2 81.95 900 1 50.60 610 2 39.27 700 2 39.27 700 2 84.66 610	77	56 800.	0.
2 64.79 900 3 77.36 800 2 64.79 800 3 64.79 800 1 61.95 800 2 81.82 800 3 61.95 900 1 50.60 610 2 39.27 700 2 39.27 700 2 84.66 610	C.R.	012	20
1 50.60 800 2 64.79 800 3 64.79 800 1 61.95 800 3 61.95 900 1 50.60 610 2 39.27 700 2 80.39 700	V	70	000
1 50.60 800 2 64.79 800 3 64.79 800 2 81.82 800 3 61.95 800 1 50.60 610 2 39.27 700 2 39.27 700 2 39.27 700	7 (2)	.006	7.00
1 50.60 800 2 64.79 800 3 64.79 800 1 61.95 800 3 61.95 900 1 50.60 610 2 39.27 700 2 39.27 700 2 84.91 700		36 800.	4. O
2 64.79 800 3 64.79 800 2 81.82 800 3 61.95 900 1 50.60 610 2 39.27 700 1 80.39 700 2 84.66 610	50	. 008 09	8.3
3 64.79 800 2 81.82 800 3 61.95 800 1 50.60 610 2 39.27 700 2 39.27 700 2 84.66 610	64	.008 800	86.2
1 61.95 800 2 81.82 800 3 61.95 900 1 50.60 610 2 39.27 700 3 44.91 700 2 84.66 610	64	.008 67	ω
2 81.82 800 3 61.95 900 1 50.60 610 2 39.27 700 3 44.91 700 2 84.66 610	61	95 800	ۍ ص
3 61.95 900 1 50.60 610 2 39.27 700 3 44.91 700 1 80.39 700 2 84.66 610	81	82 800	86
1 50.60 610 2 39.27 700 3 44.91 700 1 80.39 700 2 84.66 610	19	95 900	8.3
2 39.27 700 3 44.91 700 1 80.39 700 2 84.66 610	50	60 610	08.3
3 44.91 700 S 1 80.39 700 2 84.66 610	39	27 700	0.5
s 1 80.39 700. 2 84.66 610.	44	91 700	30
84.66 610.	80	39 700.	08.
	84	66 610.	108.38
76.15 520.	92	15 520.	08.
50.60 610	50	019 09	
2 43.51 700.00	43	51 700	108.38

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia ug N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
12 October 1981 (continued)	88	H 02 E	35.00 54.85 50.60	520.00 520.00 610.00	108.38 108.38 108.38
	86	351	47.77 50.60 49.18	610.00 700.00 520.00	108.38 108.38 108.38
	108	321	76.13 73.30 70.46	610.00 420.00 420.00	130.51 130.51 108.38
	11.8	321	67.62 70.46 64.79	420.00 420.00 520.00	108.38 130.51 130.51
	128	354	93.17 96.01 93.16	320.00 320.00 520.00	130.51 130.51 130.51
	138	321	78.99 76.13 67.62	520.00 520.00 420.00	108.38 86.24 108.38
	14S	H 20 FF	76.13 78.97 80.39	520.00 420.00 420.00	108.38

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
12 October 1981 (continued)	158	321	64.79 67.62 61.95	520.00 420.00 320.00	108.38 108.38 108.38
19 October 1981	18	351	109.34 90.19 97.85	1460.00 1380.00 1460.00	1126.66 1116.15 1116.15
	1.13	351	78.71 78.71 86.37	1380.00 1230.00 1080.00	1105.65 1095.14 1095.14
	2S	321	55.72 59.56 74.87	1080.00 1000.00 1,000.00	1084.63 1063.61 1063.61
	2B	321	59.56 76.78 59.56	920.00 840.00 1000.00	1053.10 1063.61 1053.10
	38	3 2 1	69.13 55.72 49.98	920.00 840.00 920.00	1063.61 1063.61 1074.12
	3B	7 7 7	120.82	840.00 840.00	1063.61

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia ug N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
19 October 1981 (continued)	4 S	3 2 1	90.19 90.19 90.19	770.00 840.00 770.00	1032.08 1042.59 1032.08
	ខន	321	53.82 44.24 51.90	770.00 700.00 700.00	1053.10 1053.10 1063.61
	5B	321	65.30 65.30 65.30	620.00 620.00 620.00	1042.59 1042.59
	S 9	321	44.24 44.24 36.58	620.00 620.00 770.00	1042.59 1042.59
	6B	321	28.92 42.32 28.92	460.00 460.00 460.00	1042.59 1032.08 1032.08
	78	H 02 FS	113.16 120.82 113.16	620.00 460.00 620.00	1032.08 1021.57 1021.57
	7B	- 70 F	84.45 78.71	460.00	1032.08

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia ug N/1	Nitrates and Nitrites ug N/1	Ortho- phosphate µg P/1
19 October 1981 (continued)	88 8	351	48.06 51.88 51.88	380.00 310.00 460.00	979.54 969.03 1000.56
	86	406	97.86 97.85 97.86	460.00 380.00 460.00	858.52 858.52 948.01
	108	351	76.78 86.35 84.45	380.00 380.00 380.00	979.54 1000.56 979.54
	118	H 02 E	32.75 55.72 55.72	310.00 310.00 310.00	842.92 853.43 863.94
	128	H 28 E	132.31 132.31 132.31	310.00 310.00 310.00	842.92 821.90 790.38
	138	H 0 E	74.87 86.37 74.87	310.00 310.00 310.00	727.32 716.81 727.32
	145	H 07 FF	120.82	310.00	706.30 706.30 685.29

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia µg N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
19 October 1981 (continued)	158	3 2 1	92.11 94.02 90.19	310.00 230.00 230.00	622.23 622.23 622.23
26 October 1981	18	H 26 E	147.81 143.84 118.01	850.00 850.00 850.00	352.04 352.04 374.17
	1.8	351	139.86 118.01 139.86	850.00 850.00 850.00	329.87 352.04 363.10
	2S	351	108.07 100.13 86.21	670.00 670.00 670.00	329.87 340.94 307.74
	2B	H 72 FC	96.15 100.13 104.09	670.00 670.00 670.00	285.57 285.57 318.80
	38	H 28 E	118.01 110.05 112.04	670.00 670.00 670.00	296.67 285.57 285.57
	3B	- 7 ° °	102.10	670.00	296.67 307.74

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia ug N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
26 October 1981 (continued)	48	3 2 1	121.97 121.97 119.99	670.00 670.00 670.00	263.44 241.30 241.30
	ន	351	133.90 121.97 112.04	480.00 480.00 480.00	241.27 241.27 241.27
	5B	H 0 E	133.90 137.87 127.93	480.00 480.00 480.00	241.27 230.21 219.14
	s ₉	351	125.94 121.98 127.93	480.00 480.00 480.00	174.84 174.84 174.84
	6B	351	133.90 137.87 137.87	480.00 480.00 480.00	196.97 196.97 196.97
	78	351	133.90 125.94 133.90	480.00 480.00 480.00	230.21 196.97 196.97
	7B	H 20 °C	133.90	480.00	196.97

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia ug N/1	Nitrates and Nitrites µg N/1	Ortho- phosphate ug P/1
26 October 1981 (continued)	88	3 2 1	129.92 145.82 133.90	300.00 300.00 300.00	219.14 208.07 219.11
	ន	нию	145.82 139.86 139.86	300.00	185.91 208.07 230.21
	108	351	149.80 149.80 149.80	300.00 300.00 300.00	208.07 230.21 208.07
	118	351	177.60 177.60 181.58	300.00	296.67 307.74 329.87
	128	351	163.70 159.73 171.65	300.00	196.97 208.07 196.97
	138	H 02 E	181.58 175.62 175.62	300.00	329.90 385.27 407.40
	148	351	179.59 187.54 185.56	300.00	440.63 462.80 462.80

Appendix B. (continued)

Cruise Date	Station	Replicate	Ammonia μg N/l	Nitrates and Nitrites µg N/l	Ortho- phosphate µg P/1
26 October 1981	158	1	171.64	300.00	484.93
		2	185.56	300.00	462.80
		3	175.62	300.00	462.80