Regulation of Intracellular Calcium Concentration by Nanosecond Pulsed Electric Fields

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Regulation of intracellular calcium concentration by nanosecond pulsed electric fields

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1. Introduction

Nanosecond pulsed electric fields (nsPEFs) have demonstrated the ability to induce a variety of responses in cells such as increases in intracellular free Ca2+ ([Ca2+]i), caspase activation, phosphatidylserine (PS) externalization, and DNA damage [1–7]. Classical electroporation uses field strengths of several hundred volts per centimeter for microsecond to millisecond durations to allow passage of normally non-permeable molecules across the plasma membrane. In comparison, nsPEF-exposures use much higher field strengths of several hundred kV/cm for durations often shorter or on the order of the charging time of the plasma membrane (typically 100–300 ns). The rise time of conventional electroporation pulses alone is often longer than the entire pulse duration of nsPEFs. For long pulse durations, ions accumulate along the plasma membrane, and a counterfield develops inside the cell that effectively shields the organelles from further exposure to the applied electric field. As a result, only the outer membrane is affected by the prolonged exposure and pores form once the cell-characteristic threshold voltage is reached [8–10]. The time constant for the charging process depends on cell size and conductivity of the suspension media [11,12]. For a spherical cell, 10 μm in diameter, suspended in a physiological buffer, it is on the order of 75 ns. Conversely, within the short duration of nsPEF exposures, organelle membranes will be affected in the same way as the outer cell membrane. For the high electric field strengths applied with nsPEFs, transmembrane voltages reach values of more than 1 V during the exposure and pore formation is also likely along internal membranes [13]. In addition to this direct effect on membrane structure, the intense fields allegedly affect voltage-gated channels and other protein structures in the membrane [14,15]. Ca2+ stores, such as in the endoplasmic reticulum (ER) and mitochondria, are presumably targeted by the same mechanisms, causing the release of Ca2+ into the cytosol [16]. Therefore, changes in [Ca2+]i are among the first physiological responses of the cell to nsPEFs.

In the present study, we explored changes in [Ca2+]i concentrations in Jurkat cells which maintain a baseline [Ca2+]i, of approximately 100 nM [17]. The Ca2+ stored in organelles for signaling events is approximately 30–300 μM [18] and can be released in small amounts, triggering local cellular events, and/or generating Ca2+ waves [19,20]. Cell functions that are Ca2+-mediated are dependent on varying spatial and temporal [Ca2+]i concentrations. Examples of Ca2+-mediated cell events include fertilization, muscle contraction, and apoptosis (programmed cell death) [17,21–23].

Applying nsPEFs increases [Ca2+]i, in a variety of cell types. A field strength-dependent synchronized response was first shown in Jurkat (lymphoid leukemia cell line) and HL-60 (myeloid leukemia cell line) cells [24,25]. It was concluded that both Ca2+-influx and intracellular release contributed to the observed transients. In HL-60 cells, a pulse-induced [Ca2+]i increase mimicked a hormone-stimulated increase in [Ca2+]i, where the magnitude of the response was reduced in the absence of extracellular Ca2+ and the pulse-induced increase in [Ca2+]i reached a peak in approximately 1 s [26].

Jurkat cells loaded with the Ca2+ sensitive fluorescent indicator Calcium Green showed increases in [Ca2+]i, after ten 30-ns pulses of 2.5 kV/cm [4]. The response was recorded with a resolution limit of

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100 ms. The same cells did not lose membrane integrity as confirmed by the lack of propidium iodide uptake, which indicates that the increase in \([\text{Ca}^{2+}]\) was due to internal \([\text{Ca}^{2+}]\) release rather than \([\text{Ca}^{2+}]\) influx. The experiments so far suggest that in the presence of extracellular \([\text{Ca}^{2+}]\), the majority of the increase in \([\text{Ca}^{2+}]\), following nsPEF was due to \([\text{Ca}^{2+}]\) influx. However, in the absence of extracellular \([\text{Ca}^{2+}]\), the increase in \([\text{Ca}^{2+}]\), was generated from intracellular \([\text{Ca}^{2+}]\) stores, most likely by electroportation of intracellular membranes. The influx of \([\text{Ca}^{2+}]\) from the extracellular medium is through non-specific nanopores in the plasma membrane [27] or through the opening of \([\text{Ca}^{2+}]\)-release-activated-channels (CRAC) on the membrane (termed store-operated \([\text{Ca}^{2+}]\) entry) [17].

Thus far, the temporal \([\text{Ca}^{2+}]\) response to nsPEF exposures has been observed on a 100-ms time scale, which was not fast enough to allow observation of the initial \([\text{Ca}^{2+}]\) release [4]. Within this time frame, it was also not possible to determine the source of the pulse-induced \([\text{Ca}^{2+}]\) response. Only instantaneous and uniform increases in \([\text{Ca}^{2+}]\), across the entire cell volume were reported due to nsPEF stimulation [4]. Increasing the temporal and spatial resolution of \([\text{Ca}^{2+}]\), measurements will identify the location of the \([\text{Ca}^{2+}]\) release and the underlying pathways behind the pulse-induced \([\text{Ca}^{2+}]\) response. To this end, our study (1) investigated changes in \([\text{Ca}^{2+}]\), dependent upon field strength and the presence of extracellular \([\text{Ca}^{2+}]\), (2) resolved nsPEF-induced \([\text{Ca}^{2+}]\), responses, comparing the kinetics and magnitude to spontaneous \([\text{Ca}^{2+}]\), transients, and (3) determined the origin of the \([\text{Ca}^{2+}]\) release site.

2. Methods

2.1. Cell culture

Jurkat cells obtained from American Type Culture Collection (ATCC, Manassas, VA) were cultured in 75-cm² flasks in RPMI 1640 medium (Mediatech Cellgro, Herndon, VA) supplemented with 10% fetal bovine serum (Atlanta Biologicals, Norcross, GA), L-glutamine (2.0 mM), penicillin (100 U/ml), streptomycin (100 μg/ml) (Mediatech Cellgro, Herndon, VA) and incubated at 37 °C with 5% CO₂. Cells in log-phase were removed from the culture and re-suspended in a modified Tyrode’s buffer composed of 145.0 mM NaCl, 5.0 mM KCl, 0.4 mM NaH₂PO₄, 1.0 mM MgSO₄, 6.0 mM glucose, 5.0 mM HEPES (4-(2-Hydroxyethyl)-1-piperazineethanesulfonic acid) was added to chelate residual Ca²⁺. An aliquot of stained cells was pipetted between the 80 μm electrode gap. A coverslip was placed on the gap and the slide placed in the pulse chamber set up to deliver a square wave pulse, similar to that described previously [13]. We did not use any agent such as poly-l-lysine to fix the cells to the slide since the cells did not move during the time course of the experiments. The electrode thickness was approximately 15 μm and the diameter of a typical Jurkat cell is approximately 11.5 μm [29].

2.4. Experimental setup

A Blumlein line circuit was used to deliver 60-ns rectangular pulses with a rise time of 2–4 ns to cells in suspension [30]. The pulse generator and the confocal system were synchronized such that a delay generator (Berkeley Nucleonics BCN555, San Rafael, CA) initiated the start of time-lapse movie 10 s before the application of the first pulse. If a second pulse was applied, it was triggered by the delay generator 300 s later. For the application of a single pulse, this time was set to 10 s. The delay generator and image acquisition were controlled by a MATLAB (The MathWorks, Natick, MA) program.

2.5. Fluorescence microscopy and image analysis

Time-lapse images were recorded using the UltraVIEW Rapid Confocal Imager (Perkin Elmer, Waltham, MA), a spinning-disc confocal microscope attached to an Olympus IX71 inverted microscope, capable of taking up to 360 frames per second. The confocal unit was connected to a computer with the Perkin Elmer UltraVIEW ERS software installed, which provides an interface between the excitation laser, camera controls, and the visual display. The Argon/Krypton ion laser (Melles Griot, Carlsbad, CA) 643-Y-A02, attached to the system, lasered at wavelengths 488, 568, and 647 nm. For experiments with a time scale of 1 s per frame with 100 ms exposure, a 60× long-pass air objective was used, but for the smaller time scale of 18 ms per frame with a 5-ms exposure, a 60× oil objective was used. Changes in fluorescence were derived for individual cells (region of interest) from the recorded grey-scale values with the analysis option of UltraVIEW ERS. For the statistical treatment, the results were treated with Microsoft Excel 2003 — Analysis ToolPak (Microsoft Corporation, Redmond, WA).

3. Results

3.1. Physiological \([\text{Ca}^{2+}]\), fluctuations

To characterize the dynamics of the \([\text{Ca}^{2+}]\), response to nsPEF exposure, Jurkat cells were stimulated with 60 ns duration pulses of either 25, 50, or 100 kV/cm field strength in the presence (left column) and absence (middle and right column) of extracellular \([\text{Ca}^{2+}]\) (Fig. 1). Fluorescence intensity changes expressed in grey level were obtained for each cell in the field of view. The recordings were started before the exposure to capture a baseline fluorescence value for normalization and then continued for 10–15 min (Fig. 1). Control cells exhibited spontaneous surges (oscillations) in \([\text{Ca}^{2+}]\), in the presence or absence of extracellular \([\text{Ca}^{2+}]\) (Fig. 1, panels A and B). As shown in Fig. 2, regardless of extracellular \([\text{Ca}^{2+}]\), the maximum increase in fluorescence reached values 1.35–1.40 fold above baseline values. The peak value was reached in approximately 4 s and \([\text{Ca}^{2+}]\), levels returned to values comparable to the initial signal within 1.5–
5 min. The spontaneous response was characterized by an initial moderate increase in [Ca\textsuperscript{2+}] as seen in Fig. 3A. Once the fluorescence signal reached about 10% of the peak value, the rate of change increased rapidly. The time from the onset of this steep rise until the maximum was reached is reported in Fig. 2.

Fig. 1. Typical fluorescence recordings of fluo-4 loaded Jurkat cells stimulated with a 60-ns pulsed electric field. Images of cells were taken every second with a 100-ms exposure time and stimulated with a single pulse at the 10th second (first and second column), and then exposed to an additional pulse at the 300th second (third column). Individual responses vary greatly between cells. (Note considerable deviations from the averages shown in Fig. 2 for outliers in particular for the unstipulated responses of the control groups.)

Similar [Ca\textsuperscript{2+}] transients lasting approximately 100 s, have been described [19,20]. Periodically, the fluorescence intensity, after falling to values around baseline, started to gradually increase again, although at a rate much slower than the initial rise (data not shown). Antigen receptor-stimulated release of inositol triphosphate...
For most cells, the leveled off after several minutes, however, at values higher than baseline after the pulse, which was slightly faster than measured with the nsPEF. The fluorescence signal returned to baseline within 1 min, almost every cell followed this pattern. The time at which cells suddenly lost fluorescence varied, and was observed as early as 3 min after nsPEF exposure. The sudden drop in fluorescence was usually preceded by a small spike in fluorescence.

In the absence of extracellular Ca<sup>2+</sup>, the initial rapid increase in fluorescence was similar to the response of cells exposed to 100 kV/cm in the presence of extracellular Ca<sup>2+</sup>. A peak value 1.9 times higher than the baseline level was reached in approximately 2.4 s. However, this peak value was not followed by a further gradual increase. Consequently the maximum fluorescence changes in the presence and absence of extracellular Ca<sup>2+</sup> for the 100 kV/cm field exposure were significantly different from each other (Fig. 2). The subsequent decrease in fluorescence intensity was again similar for both conditions. In this process, occasionally, the fluorescence intensity of individual cells dropped to values below the pre-exposure values after several minutes. The drop took about 15–20 s in the absence of Ca<sup>2+</sup> and 2–3 times longer in its presence. In the latter case it was preceded by a small spike in fluorescence.

Another distinct difference observed only for exposures of 100 kV/cm was the absence of any spontaneous physiological response following the pulse induced response. When a second pulse of the same field strength was applied, 5 min after the first exposure (Fig. 1, panel B) in Jurkat cells triggers oscillations in [Ca<sup>2+</sup>], [e.g. 31,32] with a slow rise similar to that shown in this study (Fig. 1).

### 3.2. Field strength dependent Ca<sup>2+</sup> response

Cells exposed to an electric field of 12 kV/cm for 60 ns exhibited no measurable increase in [Ca<sup>2+</sup>] (data not shown). The application of 25 kV/cm or more led to an immediate increase in [Ca<sup>2+</sup>] in all exposed cells. For a 25 kV/cm field exposure the peak values were 1.25 times higher than the baseline value and were reached between 2.6 and 3.0 s following stimulation (Fig. 2). The fluorescence signal returned to values measured before the exposure in approximately 30 s. A dependency of the response on extracellular Ca<sup>2+</sup> was not observed. Compared to the spontaneous response, the nsPEF stimulated response had a lower peak value, was reached in a shorter time, and was not preceded by a gradual increase.

Quantitatively, the response to 50 kV/cm was similar to the 25 kV/cm response however the peak was 1.7 times higher than baseline values. The maximum fluorescence response was reached 1.9–2.0 s after the pulse, which was slightly faster than measured with the lower field strength. No dependency on extracellular Ca<sup>2+</sup> was observed. The fluorescence signal returned to baseline within 1 min, which was slower than the return using 25 kV/cm, but still much faster than the physiological spontaneous response. For 25 and 50 kV/cm conditions, the cells showed no lasting effect of the nsPEF exposure since their [Ca<sup>2+</sup>] levels returned to pre-pulse levels within several minutes.

The response to a 100 kV/cm pulse was distinctly different from the exposures at lower field strengths. In the presence of extracellular Ca<sup>2+</sup>, cells exhibited a fast increase in fluorescence intensity within 2 s to values approximately twice as high as the baseline value. This rapid rise was followed by a more gradual increase and a maximum fluorescence value of 2.3 times above the baseline in 23 s. The slow secondary increase in fluorescence was then followed by an equally slow decrease. The fluorescence signal leveled off after several minutes, however, at values higher than baseline values. For most cells, the fluorescence signal then suddenly dropped from this value in 30–60 s to values that were even lower than the initial baseline values. With extracellular Ca<sup>2+</sup> present, almost every cell followed this pattern. The time at which cells suddenly lost fluorescence varied, and was observed as early as 3 min after nsPEF exposure. The sudden drop in fluorescence was usually preceded by a small spike in fluorescence.

In the absence of extracellular Ca<sup>2+</sup>, the initial rapid increase in fluorescence was similar to the response of cells exposed to 100 kV/cm in the presence of extracellular Ca<sup>2+</sup>. A peak value 1.9 times higher than the baseline level was reached in approximately 2.4 s. However, this peak value was not followed by a further gradual increase. Consequently the maximum fluorescence changes in the presence and absence of extracellular Ca<sup>2+</sup> for the 100 kV/cm field exposure were significantly different from each other (Fig. 2). The subsequent decrease in fluorescence intensity was again similar for both conditions. In this process, occasionally, the fluorescence intensity of individual cells dropped to values below the pre-exposure values after several minutes. The drop took about 15–20 s in the absence of Ca<sup>2+</sup> and 2–3 times longer in its presence. In the latter case it was preceded by a small spike in fluorescence.

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3.3. Kinetics of nsPEF induced [Ca\(^{2+}\)] response

The increases in [Ca\(^{2+}\)] levels following the application of nsPEFs were previously reported as occurring “as fast as they can be observed”, which was defined by a temporal resolution of 100 ms [4]. However, recent simulation studies predict the release of Ca\(^{2+}\) from internal stores through pulse-induced pores, reaching saturation thresholds in a few microseconds [33]. To further define the mechanisms leading to increased [Ca\(^{2+}\)], after the exposure to nsPEFs, we enhanced the temporal resolution. Time-lapse movies were recorded with a 5-ms exposure every 18 ms for 5 s. The pulse generator and the recording software were synchronized and a pulse was applied 1 s after recording had started. The experiments were conducted in the absence of extracellular Ca\(^{2+}\). The traces in Fig. 3A show, on an expanded time scale, several individual cell fluorescence signals in control cells similar to the traces presented in Fig. 1B. Each of the data points, taken 1 s apart, represents the fluorescence value collected for a camera exposure of 100 ms. For graphical presentation, the individual signals were shifted in time from their original recording and overlaid, with time zero defined as the time when they have reached 10% of their maximum value. It typically took several seconds until the fluorescence signal increased to this value from baseline. Following this slow onset, the rate of the fluorescence change rapidly increased, but it still took approximately 4 s to reach the peak value (Fig. 2). In comparison, the pulse induced responses were much faster. Examples of responses to different applied electric fields are shown in Fig. 3B, note that the time interval shown in panel B is forty times shorter than the one shown in panel A. A slow rise, as observed with the physiological responses, was not observed. For the applications of 50 kV/cm and 100 kV/cm fields, the signal increased 1.2 times higher than baseline values within approximately 36 ms. Physiological responses took 5000 times longer (3–4 s) to reach similar fluorescence values. Although the rate of fluorescence change was different for a field strength of 25 kV/cm, compared to the higher electric field strengths, it was not significantly different from the slow rise of a physiological response. Altogether, these results demonstrate that a 60 ns pulse with an electric field amplitude at or above 50 kV/cm stimulates a much faster Ca\(^{2+}\) response than any physiological stimulus.

3.4. Origin of pulse induced increase in [Ca\(^{2+}\)]

A series of images taken with a 5 ms exposure every 18 ms allowed us to spatially resolve the increase in Ca\(^{2+}\) concentrations and identify potential intracellular sources. Previous studies were only able to report a homogeneous increase throughout the cell [4]. Staining with the fluorescence indicator brefeldin A, BODIPY® 558/568 [34] indicates the location of the endoplasmic reticulum (ER) (Fig. 4, panel E). In the absence of extracellular Ca\(^{2+}\), there was an increase in [Ca\(^{2+}\)] emanating from the site of the ER at the poles of the cells i.e. along the direction of the electric field. The poles were affected first and more intensely than the equatorial plane. The staining of the ER was co-localized with the fluo-4 fluorescence increase observed after nsPEF stimulation. Subsequently, the fluorescence intensity increased across the cell reaching homogeneous levels within 54 ms (Fig. 4, panel D). In comparison, recordings of spontaneous Ca\(^{2+}\) signals with the same fast resolution show a gradual and even increase throughout the cell, over a longer period of time, with no specific localization for the origin of the Ca\(^{2+}\) signal (data not shown).

3.5. Fluo-4 fluorescence response and membrane integrity

The sudden drop in fluorescence levels observed after the exposure to 100 kV/cm, independent of the presence of extracellular Ca\(^{2+}\) (Fig. 1I and J), indicated a loss of membrane integrity. Cells loaded with fluo-4 and exposed, in the presence of extracellular propidium iodide (PI), to 60-ns pulses of 100 kV/cm did not show an
immediate uptake of this otherwise membrane impermeant dye (Fig. 5). However, when a sudden drop in the fluo-4 fluorescence was observed, PI was observed to be simultaneously and rapidly taken up by the cell (Fig. 5). In the presence of extracellular Ca\(^{2+}\), this delayed rapid decrease was typical for almost every cell after the application of only one pulse. In the absence of extracellular Ca\(^{2+}\), up to five pulses in close succession of 1 to 2 s had to be applied to observe this rapid drop in fluo-4 signal (data not shown). In short, whenever this fluorescence decrease occurred, simultaneous uptake of PI was always observed.

4. Discussion

The data presented show that the nsPEF-induced increase in [Ca\(^{2+}\)]\(_i\), is an immediate, electric field strength-dependent, response (Fig. 1). The presence or absence of extracellular Ca\(^{2+}\) was irrelevant for responses to electric fields of 25 and 50 kV/cm (Fig. 2). The signal rises faster to its peak value than for the spontaneous [Ca\(^{2+}\)]\(_i\) responses and also decreased faster to baseline values. After nsPEF exposure, normal fluctuations in [Ca\(^{2+}\)]\(_i\) occurred again. The cell response was different for a field of 100 kV/cm when compared to lower field strengths and with respect to the presence of extracellular Ca\(^{2+}\). Without extracellular Ca\(^{2+}\), the initial signal was similar to the response for lower fields. However, [Ca\(^{2+}\)]\(_i\) levels did not return to baseline values and no further increase above this value could be observed unless a second, similar pulse was applied. The [Ca\(^{2+}\)]\(_i\) rise in the presence of extracellular Ca\(^{2+}\) reached a considerably higher peak level much later. Another characteristic of the response to the 100-kV/cm stimulus was a delayed (within 15 min) fast drop in fluo-4 fluorescence, which could be seen for almost every cell when extracellular Ca\(^{2+}\) was present, and also for some cells when extracellular Ca\(^{2+}\) was absent.

The [Ca\(^{2+}\)]\(_i\) response, observed in the absence of external Ca\(^{2+}\) and for lower electric fields of 25 and 50 kV/cm, suggests that Ca\(^{2+}\) was released from internal stores. For 25 kV/cm a second pulse seemed to lead to amplitudes that were only slightly lower than for the first exposure. However, a second pulse of 50 kV/cm was not followed by the same increase as the first. This result indicates an inability to release more Ca\(^{2+}\) upon this second stimulus, either because the cell had become less sensitive to the exposure, or Ca\(^{2+}\) stores were depleted. This threshold seems to be overcome by increasing the field strength to 100 kV/cm. The significantly larger signal amplitude, in the presence of extracellular Ca\(^{2+}\), was caused by the influx of Ca\(^{2+}\). This hypothesis was supported by the slower increase to this higher value, which followed the initial rapid development similar to the response observed with no extracellular Ca\(^{2+}\). How Ca\(^{2+}\) entered the cell is not completely understood. A likely explanation is that nsPEF causes the formation of nanopores which are large enough to allow Ca\(^{2+}\) to enter the cell but are too small for the uptake of larger molecules such as PI [27,35] or for fluo-4 to flow through. In our experiments, no observed increase in PI fluorescence while intracellular fluo-4 signals were high, supports this explanation (Fig. 4B). Conversely, the uptake of PI several minutes after 100 kV/cm nsPEF exposure correlated well with a sharp decrease in fluo-4 fluorescence below baseline levels, indicating that large pores had opened in the membrane as a secondary response to the exposure (Fig. 4B). Efflux from the cell and dilution of fluo-4 in the large extracellular volume would account for the decline in fluorescence signal. The lack of spontaneous fluctuations in [Ca\(^{2+}\)]\(_i\), observed after the pulse at this field strength indicates that there was significant damage to the cell that prohibited normal Ca\(^{2+}\) homeostasis events to occur.

We recognize the possibility that the presence of EGTA in the low Ca\(^{2+}\) condition may lessen the ability of nsPEF to electroporate the plasma membrane. Chelators have been shown to alter the stability and rigidity of membranes (this is based on studies performed in red blood cells) [36]. However, since we still observe nsPEF induced increases in [Ca\(^{2+}\)]\(_i\) in the EGTA condition, this implies that the nsPEF is affecting intracellular Ca\(^{2+}\) stores. Even if there was no electroporation of the plasma membrane in the EGTA condition this would not influence our results. Studies in our laboratory showed that by using patch clamp techniques, the permeabilization of GH3, PC-12 and Jurkat cells was not influenced by 10 mM EGTA [14].

In addition to nanopores allowing the influx of Ca\(^{2+}\) following nsPEF-stimulation, an alternative pathway for Ca\(^{2+}\) entry could be the activation of Ca\(^{2+}\)-release-activated-channels (CRAC) in the plasma membrane. This could be achieved with field strengths of 100 kV/cm which are large enough to trigger capacitive Ca\(^{2+}\) entry through the initial release of Ca\(^{2+}\) from internal stores [26,37]. The results for lower field strengths show that the electric fields were affecting internal stores and leading to increased [Ca\(^{2+}\)]. The endoplasmic reticulum was expected to be the most likely source for this Ca\(^{2+}\) increase and our fast time resolution images support this assumption (Fig. 4A).

Spontaneous increases in [Ca\(^{2+}\)]\(_i\), took approximately 4 s to reach a maximum and consequently could easily be resolved with an image taken every 18 ms. It is obviously a response, with extracellular Ca\(^{2+}\) present, that involves the opening of Ca\(^{2+}\) channels in the plasma membrane, which are known to occur on the order of milliseconds [38]. In comparison, the initial temporal response of the pulse-induced Ca\(^{2+}\) release was found to occur much faster. The change in [Ca\(^{2+}\)]\(_i\), for cells pulsed at 50 and 100 kV/cm showed a fast rise to values comparable to spontaneous oscillation peak amplitudes within the first 100 ms of exposure (often within 2–3 frames, i.e., 54 ms). Even on this time scale, the onset of the increase appears to be instantaneous with no indication of a gradual activation of ion channels. For 25 kV/cm the increase in [Ca\(^{2+}\)]\(_i\), was too slow to allow us to determine with statistical significance if the induced response differs from the response of cells in the control. This suggests that either a fast rise in Ca\(^{2+}\) is triggered in less than 18 ms, or fluo-4 cannot bind quickly enough to the pulse-mobilized Ca\(^{2+}\), although fluo-4 operates within milliseconds [39].

These results suggest that the mechanism by which nsPEFs induce increases in [Ca\(^{2+}\)]\(_i\), may not be mediated by IP\(_3\) and may not be following a physiological pathway. However, a contribution to increased [Ca\(^{2+}\)]\(_i\), from internal stores through directly activated channels cannot be excluded. Recent experimental results and simulations suggest that an instantaneous and substantial increase in [Ca\(^{2+}\)]\(_i\), observed on a millisecond timescale is caused by the formation of pores. These pores form as a result of rapid changes in membrane potentials that surpass the membrane threshold due to instantaneous with no indication of a gradual activation of ion channels. For 25 kV/cm the increase in [Ca\(^{2+}\)]\(_i\), was too slow to allow us to determine with statistical significance if the induced response differs from the response of cells in the control. This suggests that either a fast rise in Ca\(^{2+}\) is triggered in less than 18 ms, or fluo-4 cannot bind quickly enough to the pulse-mobilized Ca\(^{2+}\), although fluo-4 operates within milliseconds [39].

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of this trend with our experiments suggests that with the 60-ns pulse, the applied electric field reaches into the cell and that pores are formed at an internal membrane, most likely the ER. This may allow for the passage of Ca\(^{2+}\), which we observe as an increase in fluo-4 fluorescence intensity. Also, the pulse induced Ca\(^{2+}\) release generally shows a faster and more exponential decline, rather than a linear decrease, back to baseline values as seen with the spontaneous increases (e.g. Fig. 1F and G), which suggests some mechanism other than the mere opening and closing of Ca\(^{2+}\) channels.

The creation of pores in the outer membrane also seems to be the primary process for the uptake of Ca\(^{2+}\) from the external medium. Evidence for the formation of pores in Jurkat cells, following nsPEF treatment, was previously observed when the influx of YO-PRO-1 was observed along with externalization of phosphatidylserine [3]. At the same time, no PI entered the cells, which supports the theory that these pores are different than those induced by electroporation pulses, in particular, that they are much smaller and can discriminate between dyes with similar molecular weights. PI has a Connolly solvent exclusion volume of 394 Å\(^3\) which is slightly larger than YO-PRO-1 which is 355 Å\(^3\) (calculated using Chem 3D Ultra version 8.0.3, CambridgeSoft Corp, Cambridge, MA), which may be a sufficient difference to account for their different permeabilities through nanopores. Influx of Ca\(^{2+}\) through nanopores in the plasma membrane together with the outflow of Ca\(^{2+}\) from internal stores appears adequate to activate CRAC channels on the plasma membrane. We believe that this can be seen in the further 45% increase in [Ca\(^{2+}\)] when extracellular Ca\(^{2+}\) was present (Fig. 1I) from a [Ca\(^{2+}\)]\(^+\) value of 2.0 s, which was similar to the maximum seen without extracellular Ca\(^{2+}\) at 2.4 s (Fig. 1J), to a peak level which was slowly reached at 22.9 s (Fig. 2).

In conclusion, our results confirm modeling results that predict, for the exposure to nanosecond pulsed electric fields with field strengths of several tens of kilovolts per centimeter, the formation of pores in the plasma and also internal membranes [27,25]. These nanopores are too small for larger molecules, such as PI, to pass through but permit passage of smaller ions like Ca\(^{2+}\). Longer electrical pulses have been shown to produce primarily large pores in the plasma membrane that will permit the introduction of big molecules such as DNA into cells. However, organelles can be affected as well. Electrical field pulses of 50-µs duration were shown to produce reversible breakdown of plasma membrane and tonoplast membranes as judged by the influx of extracellular Ca\(^{2+}\) and the release of tonoplast Ca\(^{2+}\) in aequorin-transformed tobacco cells [46].

For nsPEFs of different pulse durations and field strengths, combined with multiple exposures, our data suggests the possibility of activating specific cell responses, such as the expression of interleukin-2 in Jurkat cells. To this end, it would be of interest to observe the long-term effects of exposures. For certain pulsed electric field parameters the induction of apoptosis has already been demonstrated [47]. Based on our research, the ability to trigger other Ca\(^{2+}\) controlled responses is likely. For platelets this response has already been exploited as non-ligand agonist for platelet activation, Arch. Biochem. Biophys. 471 (2008) 240–248.


E.S. Buescher, R.R. Smith, K.H. Schoenbach, Submicrosecond intense pulsed electric field generators for the study of subcellular effects, Bioelectromagnetics J. 27 (2006) 1276.


