

Spring 1984

A Molecular Phylogeny of the Grunts (Perciformes: Haemulidae) Inferred from Nuclear RAG1 Gene Sequences

Millicent D. Sanciangco
Old Dominion University

Follow this and additional works at: https://digitalcommons.odu.edu/biology_etds



Part of the [Evolution Commons](#), and the [Marine Biology Commons](#)

Recommended Citation

Sanciangco, Millicent D.. "A Molecular Phylogeny of the Grunts (Perciformes: Haemulidae) Inferred from Nuclear RAG1 Gene Sequences" (1984). Master of Science (MS), Thesis, Biological Sciences, Old Dominion University, DOI: 10.25777/n9hh-9v94
https://digitalcommons.odu.edu/biology_etds/257

This Thesis is brought to you for free and open access by the Biological Sciences at ODU Digital Commons. It has been accepted for inclusion in Biological Sciences Theses & Dissertations by an authorized administrator of ODU Digital Commons. For more information, please contact digitalcommons@odu.edu.

**A MOLECULAR PHYLOGENY OF THE GRUNTS (PERCIFORMES:
HAEMULIDAE) INFERRED FROM NUCLEAR RAG1 GENE SEQUENCES**

by

Millicent D. Sanciangco
B.S. Zoology, 2002, University of the Philippines Los Baños

A Thesis Submitted to the Faculty of
Old Dominion University in Partial Fulfillment of the
Requirement for the Degree of

MASTER OF SCIENCE

BIOLOGY

OLD DOMINION UNIVERSITY

May 2007

Approved by:

Kent Carpenter (Director)

John Holsinger (Member)

Christopher Osgood (Member)

ABSTRACT

A MOLECULAR PHYLOGENY OF THE GRUNTS (PERCIFORMES: HAEMULIDAE) INFERRED FROM NUCLEAR RAG1 GENE SEQUENCES

Millicent D. Sanciangco
Old Dominion University, 2007
Director: Dr. Kent Carpenter

Species and genera of Haemulidae have undergone various taxonomic revisions, however, there is no study that infers the phylogeny of the haemulid genera using morphological or molecular data. The purpose of this study was to use approximately 1386 base pairs of the nuclear Recombination Activation Gene-1 (RAG1) from 35 haemulid species representing 13 genera, one species of the closely related Inermiidae, and two species of the outgroup Sparidae to infer an intrafamilial phylogeny of Haemulidae. This analysis is corroborated using approximately 650 base pairs of the mitochondrial Cytochrome Oxidase I (COI) gene and RAG1-COI combined gene analyses of 27 haemulids, an inermiid, and two sparids. Results show strong support for a monophyletic Haemulidae. However, the placement of Inermiidae within the proposed superfamily Haemuloidea remains unresolved. The subfamilies Haemulinae and Plectorhinchinae are recovered from both maximum parsimony and maximum likelihood analyses using RAG1, COI, and RAG1-COI genes combined. These analyses also recovered similar clade components within these subfamilies, with some exemptions. The RAG1 gene phylogeny combined with distribution data also revealed a biogeographic pattern that suggests a specific radiation of haemulids. There was strong support for a basal paraphyletic Old World (coastal Eurasia, Africa, Australia, and

western central Pacific) group, a derived monophyletic New World (coastal Americas) group, and an intermediate Old World-New World group, which can be accounted for by the closing Tethys Sea and Atlantic Ocean widening vicariant events. In addition, molecular data using RAG1 and COI genes also highlighted potential problems regarding the validity of several haemulid genera and suggest a re-evaluation of these genera. Finally, this study indicates that the nuclear RAG1 gene is useful for inferring phylogeny at the intrafamilial level for this percoid family of fishes.

This thesis is dedicated to Jonnell and Andre.

ACKNOWLEDGMENTS

I would like to thank Dr. Kent Carpenter for his unending support and guidance throughout the duration of this project and throughout the duration of my Master's degree. I would also like to thank my committee members, Dr. John Holsinger and Dr. Christopher Osgood for their comments and advice. I am grateful to Andrew Mahon for all the help in the lab, constant support, and providing some of the RAG1 sequences. I thank the BSSF staff at ODU for assisting with my sequencing reactions and Dr. Robert Ratzlaff for letting me use the thermal cycler in his lab. I also thank Karin Berling and the Molecular Systematics Laboratory group for help and knowledge. I thank Dr. Thomas Orrell for providing help and suggestions on my analyses. Thanks also to Dr. Yukio Iwatsuki, Dr. Paul Hastings, and H.J. Walker for providing some haemulid tissues and Dr. Guillermo Orti for the two haemulid RAG1 sequences. I am grateful to Dr. Giacomo Bernardi for providing an article and suggestions. I also would like to thank the FishBOL Project for providing us with haemulid COI sequences. I am indebted to my parents, in-laws, Ate Len, and Ate Ivy for unending support and encouragement. My Master's program was funded by the Fulbright Philippine Agriculture Scholarship Program and in part by a graduate research assistantship through an IUCN (World Conservation Union) project at Old Dominion University.

TABLE OF CONTENTS

	Page
LIST OF TABLES	vii
LIST OF FIGURES	viii
INTRODUCTION	1
Recombination Activating Gene-1 (RAG1)	9
MATERIALS AND METHODS.....	12
Chemicals.....	12
Equipment	12
Specimen Samples	13
DNA Extraction	13
Gel Electrophoresis.....	16
DNA Amplification	17
PCR Purification	18
Sequencing.....	19
Data Analyses	20
RESULTS	22
DISCUSSION	37
CONCLUSION.....	44
REFERENCES	45
APPENDICES	52
1. RAG1 sequences of haemulids, including outgroups.....	52
2. COI sequences of haemulids, including outgroups.....	68
3. RAG1-COI concatenated sequences of haemulids, including outgroups.....	74
VITA.....	92

LIST OF TABLES

Table	Page
1. Distribution and Biogeographic (Old World vs New World) Designation of Species Used in this Study.....	2
2. Chemicals Used and Product Information	12
3. List of Species and the Accession Number of Haemulid Specimens	14

LIST OF FIGURES

Figure	Page
1. Total Number of Substitutions Plotted as a Function of Percent Sequence Divergence of Pooled Transitions for RAG1 Sequence Data.....	24
2. Total Number of Substitutions Plotted as a Function of Percent Sequence Divergence of Pooled Transversions for RAG1 Sequence Data	24
3. Total Number of Substitutions Plotted as a Function of Percent Sequence Divergence of Pooled Transitions for COI Sequence Data	25
4. Total Number of Substitutions Plotted as a Function of Percent Sequence Divergence of Pooled Transversions for COI Sequence Data.....	25
5. Total Number of Substitutions Plotted as a Function of Percent Sequence Divergence of Pooled Transitions for RAG1-COI Sequence Data	26
6. Total Number of Substitutions Plotted as a Function of Percent Sequence Divergence of Pooled Transversions for RAG1-COI Sequence Data	26
7. Maximum Parsimony Cladogram with Nodes Collapsed with Less Than 50% Bootstrap Support and Bootstrap Values of 50 and Above Indicated at the Respective Nodes for RAG1 Sequence Data.....	27
8. Maximum Likelihood Cladogram with Nodes Collapsed with Less Than 50% Bootstrap Support and Bootstrap Values of 50 and Above Indicated at the Respective Nodes for RAG1 Sequence Data.....	28
9. Maximum Parsimony Cladogram with Nodes Collapsed with Less Than 50% Bootstrap Support and Bootstrap Values of 50 and Above Indicated at the Respective Nodes for COI Sequence Data	29
10. Maximum Likelihood Cladogram with Nodes Collapsed with Less Than 50% Bootstrap Support and Bootstrap Values of 50 and Above Indicated at the Respective Nodes for COI Sequence Data	30
11. Maximum Parsimony Cladogram with Nodes Collapsed with Less Than 50% Bootstrap Support and Bootstrap Values of 50 and Above Indicated at the Respective Nodes for RAG1-COI Sequence Data.....	31
12. Maximum Likelihood Cladogram with Nodes Collapsed with Less Than 50% Bootstrap Support and Bootstrap Values of 50 and Above Indicated at the Respective Nodes for RAG1-COI Sequence Data.....	32

Figure	Page
13. Proposed Radiation of the Members of Haemulidae Using Maximum Likelihood Analysis for RAG1 Sequence Data	33

INTRODUCTION

Perciformes is the largest order of vertebrates with over 10,000 species (Nelson, 2006). The classification of this order is tentative and its members thought to be polyphyletic. Perciformes contains 20 suborders, which includes the suborder Percoidei. There are 79 families in this suborder, one of which is Haemulidae. This family is one of the most ecologically and commercially important groups of nearshore fishes. They are typically shallow-water species inhabiting corals reefs, rocky bottoms, seagrass beds, sand flats, and mud-bottoms in coastal and estuarine waters. As one of the most speciose families of percoids, they are circumtropically distributed in the Atlantic, Indian, and the Pacific Oceans (Table 1). Nelson (2006) recognized about 145 haemulid species in 17 to 18 genera. Several species and generic taxonomic revisions have been made of this family, and a number of articles on the systematics and distribution of individual species exist (Konchina, 1976; Courtenay, 1961; Nelson, 1994, 2006; Lindeman and Toxey, 2003; McKay, 1984, 2001). There is no systematic study yet conducted on haemulids employing molecular or morphological techniques to infer a phylogeny of the genera and species within Haemulidae.

There are no hypotheses of outgroups for haemulids based on systematic studies, but some of the suggested outgroups include Inermiidae, Lutjanidae, Nemipteridae, Sciaenidae, and Sparidae (Lindeman and Toxey, 2003). These belong in superfamilies proposed by Johnson (1980): Lutjanoidea, Sparoidea, and Haemuloidea, who found no conclusive evidence to suggest relationships by immediate common ancestry between

Table 1
Distribution and biogeographic (Old World vs New World) designation of species used in this study

Species	Distribution	Old World/ New World
<i>Anisotremus dovii</i>	Eastern Pacific: southern margins of Gulf of California to Peru.	New World
<i>Anisotremus interruptus</i>	Eastern Pacific: Gulf of California to Peru, including the Galapagos Islands.	New World
<i>Anisotremus pacifici</i>	Eastern Pacific: Mexico to Peru.	New World
<i>Anisotremus virginicus</i>	From Bahamas and Florida throughout much of the area, extending southward to Brazil. In Bahamas, recorded primarily from the ventral and northern islands.	New World
<i>Brachydeuterus auritus</i>	Along the West African coast from Mauritania (exceptionally Morocco) to Angola.	Old World
<i>Conodon serrifer</i>	Eastern Pacific: Gulf of California to Peru.	New World
<i>Diagramma pictum</i>	Indo-West Pacific from south Natal to southern Japan and Fiji, excluding northern Australia and most of southern New Guinea. (Includes tropical coasts of Australia).	Old World
<i>Genyatremus luteus</i>	Southern Lesser Antilles and northern coast of South America from eastern Colombia to Brazil.	New World
<i>Haemulon aurolineatum</i>	Western Atlantic from Chesapeake Bay and Bermuda southward throughout much of the area to Brazil.	New World
<i>Haemulon flaviguttatum</i>	From Chesapeake Bay and Gulf of Mexico, southward throughout much of the area to Brazil. Eastern Pacific: Mexico to Ecuador.	New World
<i>Haemulon plumieri</i>	Western Atlantic: Chesapeake Bay, through the Gulf of Mexico and Caribbean southward to Brazil, including Antilles	New World
<i>Haemulon sciurus</i>	Western Atlantic from the lower Gulf of Mexico, South Carolina and the Bahamas southward throughout much of the area to Brazil; also in Bermuda.	New World
<i>Haemulon scudderii</i>	Eastern Pacific: Mexico to Ecuador, including the Galapagos Islands.	New World
<i>Haemulon steindachneri</i>	Juveniles recorded from Guatemala. Adults recorded from Panama along the coast of South America, Brazil. Also recorded from the tropical eastern Pacific from the sea of Cortez to Peru. Western Atlantic: Panama to Rio de Janeiro, Brazil.	New World
<i>Haemulopsis axillaris</i>	Eastern Central Pacific: Mazatlan, Mexico to Ecuador.	New World
<i>Haemulopsis elongatus</i>	Eastern Central Pacific: Mazatlan, Mexico to Panama.	New World
<i>Haemulopsis leuciscus</i>	Eastern Pacific: Gulf of California to Peru.	New World
<i>Haemulopsis nitidus</i>	Eastern Pacific: Gulf of California to Peru.	New World
<i>Inermia vittata</i>	Western Atlantic from Bermuda, Florida to Bahamas, Belize, and northern South America. Also Caribbean, including Antilles	New World
<i>Orthopristis chalceus</i>	Eastern Pacific: Mexico to Panama, including the Galapagos Islands.	New World
<i>Parapristipoma octolineatum</i>	West African coast including islands, from the Straits of Gibraltar to Angola; northward extending into the Western Mediterranean and along the coasts of Portugal and Spain.	Old World
<i>Parapristipoma trilineatum</i>	Indo-West Pacific: southern Japan, East China Sea and Taiwan.	Old World
<i>Plectorhinchus chaetodonoides</i>	Indo-West Pacific from East Africa to Ryukyu Islands and wide ranging throughout South Pacific eastwards to Rapa. In the area, recorded only from Mauritius. Elsewhere, East Indies to the Western Pacific.	Old World
<i>Plectorhinchus lessonii</i>	Zanzibar, Red Sea to Sri Lanka, Indonesia, Queensland, Polynesia to Philippines, Taiwan Province of China, and southern Japan.	Old World
<i>Plectorhinchus macrolepis</i>	West African coast, from Senegal to Congo.	Old World
<i>Plectorhinchus schotaf</i>	Indo-West Pacific from Port St. Johns, Transkei, East Africa to northern Australia, the Philippines, and southern Japan. Red Sea population differs slightly in coloration.	Old World
<i>Plectorhinchus sordidus</i>	Found in the Red Sea, off Mozambique and the west coast of Madagascar and Mauritius.	Old World
<i>Plectorhinchus vittatus</i>	Indo-West Pacific from East Africa to Indonesia, northern Australia, Thailand, Philippines, and Polynesia.	Old World
<i>Pomadasyys argenteus</i>	Found in Red Sea to southern Japan, including northern Australia from Exmouth Gulf of Clarence River, New South Wales. Elsewhere, eastward extending to the Philippines.	Old World

Table 1 (*continued*)

Species	Distribution	Old World/ New World
<i>Pomadasys argyreus</i>	India, Sri Lanka to Papua New Guinea, excluding northern Australia. In the area, found in Pakistan to Sri Lanka. Elsewhere, eastward extending to the Philippines.	Old World
<i>Pomadasys branickii</i>	Eastern Pacific: Mexico to Peru.	New World
<i>Pomadasys kaakan</i>	Indo-West Pacific from Transkei to Red Sea, Persian Gulf, India, Indonesia to China (including Taiwan Province), including northern Australia from Exmouth Gulf to Moreton Bay. In the area, found along the east coast of Africa (including Madagascar, the Seychelles, Farquhar and Comoro islands).	Old World
<i>Pomadasys maculatus</i>	East coast of Africa, Madagascar, Red Sea, Gulf of Aden, Persian Gulf, Pakistan, India, Sri Lanka to northern half of Australia from Shark Bay to Moreton Bay, New Guinea, Philippines to southern Japan.	Old World
<i>Xenichthys xanti</i>	Eastern Pacific: southern Gulf of California to Peru.	New World
<i>Xenistius californiensis</i>	Eastern Pacific: Monterey Bay in California, USA to Peru; common in southern California but rare north of Point.	New World

any two of the three superfamilies. However, he found that the families Haemulidae and Inermiidae (bonnetmouths) share a unique projection on the lateral side of the metapterygoid and several other osteological features and certain muscle complexes, placing them in a proposed monophyletic superfamily Haemuloidea. He described the family Haemulidae as having an often upturned small to moderate mouth; well-developed ethmo-maxillary and palato-maxillary ligaments; presence of cardiform teeth in jaws with the outer row often enlarged; vertebrae 10 + 16 or 11 + 16; pleural ribs eight to nine; absence of parapophyses on the first two vertebra but beginning on third or fourth; well-developed teeth often bearing hornlike extensions on large upper and lower pharyngeals; a well-developed supraoccipital crest on the neurocranium; frontals without crests; and absence of median trough in the interorbital area. He contended that bonnetmouths are haemulid derivatives due to the presence of haemuloid suspensorium, enlarged chin pores, and procurrent spur, and assigned them familial status (Inermiidae). The major difference between haemulids and inermiids is accounted for by the specialization in the

parapophyses, upper jaw structure, and in their lifestyle. Inermiids are midwater planktivore while haemulids are mostly benthic carnivores.

Johnson (1980) further subdivided the haemulids into two subfamilies — Haemulinae and Plectorhinchinae, based on a combination of several external and anatomical characters. Haemulinae is comprised of *Haemulon*, *Haemulopsis*, *Orthopristis*, *Pomadasys*, *Anisotremus*, *Conodon*, *Isacia*, *Genyatremus*, *Boridia*, *Brachydeuterus*, *Xenistius*, *Xenichthys*, *Xenocys* and *Parakuhlia*. *Plectorhinchus*, *Diagramma* and *Parapristipoma* are included in the subfamily Plectorhinchinae. Plectorhinchinae have dorsal rays IX to XIV, 17 to 26; anal rays III, six to eight; vertebrae 11 + 16; pleural ribs nine; epipleural ribs ten to 12; no epibranchial toothplate on the third position; between four to six chin pores; origin of retractor dorsalis on the second vertebrae; absence of parapophyses on the first two vertebrae; absence of fronto-ethmoid trough on the neurocranium; and absence of specialization on the upper jaw. Adult members of Plectorhinchinae often have thick fleshy lips, hence called rubberlips or sweetlips. Haemulinae have dorsal rays XI to XIV, 11 to 18; anal rays III, seven to eight; vertebrae 10 + 16; pleural ribs eight; epipleural ribs seven to eleven; a third epibranchial tooth plate; two enlarged chin pores behind symphysis or a median longitudinal groove or pit, or both; large retractor dorsalis originating on second or second and third vertebrae; the absence of parapophyses on the first two vertebrae; absence of fronto-ethmoid trough on the neurocranium; and absence of specialization on the upper jaw. The last three characters unite the two subfamilies and also distinguish them from inermiids (Nelson, 2006; Johnson, 1980).

The etymology of the term “haemulid” was from the Greek word “haimaleos” which means bloody or blood gums, referring to the red coloration of the interior mouth (Brown, 1956). The family common name “grunt” was derived from the distinctive stridulatory sound they produced by rubbing their pharyngeal or jaw teeth during feeding or deliberately as a fright response or territorial display (Konchina, 1977). Their swim bladder functions as a resonator amplifying the sounds produced by the pharyngeal teeth (Johnson, 1980). Grunting sounds, in combination with “knocks” or “thuds” for some haemulids becomes weak during competitive feeding and louder under duress (netted, handled, or when an electrical stimulation is applied) (Fish and Mowbray, 1970). The sounds produced are also species-specific which also aids in identification during recordings (Fish and Mowbray, 1970). Other common names include sweetlips, rubberlips, hotlips, velvetchins, roncadors, pigfish, and burros (Allen and Robertson, 1994; McKay, 2001).

Haemulids are oblong, compressed perch-like fishes averaging to 70 cm in total length (McKay, 2001). Earliest members reported for this family have been dated back to the Eocene epoch of the Tertiary period and hypothesized to exhibit the primitive branchial skeleton among the perciforms (Johnson, 1980; Wainwright, 1989). In most cases, all members possess a typical convex head profile, two anterior pores, and a median groove on chin, except for the genus *Genyatremus*, which do not possess a median groove (Lindeman and Toxey, 2003). Other characteristic features of the grunts include a single dorsal fin with fin rays XI to XIV, 11 to 19; a moderate to long pectoral fin and below its base, a pelvic fin with a single spine and five soft rays; anal rays III, six to 13; small to moderate ctenoid scales, extending onto the head but not on the front of

snout, lips, and chin. Grunts have a highly variable coloration, ranging from uniformly colored, striped, banded, blotched, and spotted (Roux et al., 1981). Color patterns are distinctive of adult grunts, while early juveniles of *Anisotremus*, *Orthopristis*, and *Haemulon* ranging from two to five centimeters, exhibit similar dark dorsolateral and midlateral stripes and a caudal spot (Lindeman, 1986). The shape, development of lips, and coloration can be used to discriminate juveniles from adult grunts. McKay (2001) and Courtenay (1961) characterized early juvenile stages of *Haemulon* based on the length of the upper eye stripe in combination with other characters. Although differences in adults can be distinguished by pigmentation, *Haemulon* can easily be distinguished from other haemulid genera by the presence of a serrate opercle, lack of vomerine teeth, association of last dorsal spine with the dorsal rays, and possession of scaly dorsal and anal fins (Hong, 1977). Misidentification of *Plectorhinchus* species is a problem due to diverse coloration in different developmental stages. However, McKay (1984) classified members of the genus *Plectorhinchus* based on a combination of juvenile and adult coloration, dorsal fin counts, and gill raker counts (McKay, 1984). He described *Plectorhinchus* species as having 11 to 14 spines in the dorsal fin and with fewer lateral line scales. The status of the genus *Parakuhlia* is confused because they are variously placed in the Kuhliidae, or in the Haemulidae either as a separate genus or synonymous to *Haemulon* (Johnson, 1980; Nelson, 1994, 2006). Roux et al. (1981) suggested further analysis is needed on several species of West African *Parapristipoma* and *Pomadasys* to establish more clearly their taxonomic status. Moreover, the taxonomic status of the genus *Hapalogenys* within the haemulids remains problematic. Springer and Raasch (1995) established the family name Hapalogeniidae for *Hapalogenys*, a genus of

uncertain relationships (Iwatsuki et al., 2000; Lindeman and Toxey, 2003). McKay (2001) described the similarity of *Hapalogenys* to two species of Dinopercidae, but which lacks intrinsic muscles on the posterior end of the swimbladder. Richardson (1844) described *Hapalogenys* as having the following characteristics: a single notched dorsal fin; papillae on the fleshy lower lip; 10 pores on the chin; a procumbent spinelike process (exposed tip of first pterygiophore) at origin of dorsal fin, and seven branchiostegals, and also classified *Hapalogenys* under haemulids. Iwatsuki et al. (2000) and Iwatsuki and Russell (2006) described a total of seven nominal species of *Hapalogenys*, now accepted as valid species names, under the family Haemulidae. Iwatsuki and Nakabo (2005) further described diagnostic characters supporting *Hapalogenys* position within the haemulids, but acknowledged that further evaluation on the genera and species identity of Haemulidae must be conducted. Further, re-description of certain haemulids has resulted in generic re-assignments, such as in the case of *Orthopristis brevipinnis*, which is now placed in the genus *Microlepidotus* (Cooke, 1992).

Haemulids occur worldwide in tropical and warm temperate seas. *Haemulon* and *Anisotremus* are inhabitants of coral reefs or hard-bottom areas and are nocturnal feeders on sand and grass flats (Helfman et al., 1982). Members of *Genyatremus*, *Pomadasy*, and *Conodon* are mud-bottom dwellers, often in turbid brackish water. *Orthopristis* are found on both soft-bottom and hard-bottom habitats. Most grunts are euryhaline fishes, and may enter rivers but seldom enter freshwater (McKay, 2001). Juveniles inhabit more shallow water compared to an adult depth range down to about 115 m. Juveniles may develop a habitat shift during growth and become ecologically and spatially separated for

a significant period of time (Helfman et al., 1982; Williams et al., 2004a; Morinière et al., 2003).

Juvenile grunts aggregate in multispecies schools swarming over rocky reefs during the day as a co-evolved predator-defense response. This response can be exacerbated by conditions such as increased water turbidity due to siltation, presence of dense particulate matter suspension, or presence of human observers (McLean and Herrnkind, 1971). Although the schooling behavior diminishes as grunts become older, nocturnal migrations of schooling grunts occur over long distances when they leave the reef to feed at night on sandy bottoms and seagrass beds (McFarland and Wahl, 1996; Ogden and Ehrlich, 1977; Nagelkerken and van der Velde, 2004). Most haemulids are carnivorous, feeding opportunistically on crustaceans, polychaete worms, clams, and echinoids, while smaller species primarily feed on plankton (Konchina, 1977; Ogden and Ehrlich, 1977; Williams et al., 2004a). Stomach content and stable isotope analysis revealed that haemulids undergo dietary changes during ontogeny feeding on increasingly larger prey with increasing trophic level (Morinière et al., 2003). Postrecruitment mortality of grunts was positively correlated with density of predators, including larger conspecifics (Tupper and Juanes, 1999). Grunts are considered gonochoristic, with males and females occurring in all size classes, and display no differences in coloration between sexes (Williams et al., 2004a). The description of unusual male urogenital apparatus of grunts suggests that sexual dimorphisms are probably associated with reproduction (Rasotto and Sadovy, 1995). Purcell et al. (2006) hypothesized that *Haemulon* species spawns throughout the year on a bi-monthly basis. Since haemulids are nonguarders and produce pelagic eggs, the production of mucin, a complex protein usually associated with

teleost reproduction, may only indicate a pheromonal role (Rasotto and Sadovy, 1995). Recorded duration of pelagic existence for haemulids is about two weeks, as indicated by the limited period between fertilization and settlement (McFarland et al., 1985). The lack of documented spawning events, however, may suggest that reproduction occurs at dusk (Lindeman and Toxey, 2003; McFarland et al., 1985). Haemulids employ the carangiform mode of swimming and exhibit normal activity level. Haemulids are important reef fish ecologically and economically. They play a significant role in the ecosystem as a nutrient (phosphorus and nitrogen) resource, while also stimulating biological activity in the reef benthic community (Meyer and Schultz, 1985; Ogden and Ehrlich, 1977). They can be used as health indicators of the reef to assess the conditions brought about by anthropogenic effects such as overfishing, dynamite fishing, cyanide fishing, aquarium collection, pollution, and curio collection (Hodgson, 1999; Tupper and Juanes, 1999).

Grunts are an important component in commercial fisheries. The recorded global capture production by continent averaged 74,533 tons from 1950 to 2004 (FAO, 2006). Grunts are caught by hook and line, fish traps, bottom trawls, gillnets, and beach seines, and are marketed fresh, filleted, or salted (Lindeman and Toxey, 2003; McKay, 2001; Roux, 1981). Several grunt species are good recreational gamefish, and others are suitable for aquarium display.

Recombination Activation Gene-1 (RAG1)

Molecular data can help resolve taxonomic uncertainties and answer phylogenetic questions when morphological data is limited. This has proven to be effective in

constructing phylogenies consistent with demographic and speciation models done on *Anisotremus* (Bernardi and Lape, 2005) and in identifying population structure and spatial genetic patterns of *Haemulon* species (Williams et al., 2004a; 2004b; Durán-Gonzalez et al., 1990). The nuclear RAG1 marker has many unique properties including: rarity of indels; minimal saturation of transition changes at third positions of codons; little or no GC and codon biases; and highly stationary base composition that are suitable for reconstructing phylogenies of different vertebrate taxa (Groth, 1999). Reduced level of homoplasies in phylogenetic studies can be accounted for by the slow evolution of RAG1 gene, in combination with other properties mentioned above (Groth, 1999). RAG1 is a single copy gene, hence eliminates questions regarding gene duplication and messy allelic variations. RAG1 and RAG2 were presumably acquired by vertebrates through lateral gene transfer from a viral genome when the first vertebrates evolved 450 million years ago (Feng et al., 2005). Both genes evolved adjacent to one another and mediate V(D)J recombination, suggesting their role in generation of antibodies and T-cell receptors (Feng et al., 2005). Studies show that vertebrates use the RAG1 gene only as part of the machinery that creates somatic gene variability for the immune system. Therefore, this would also suggest that the RAG1 gene is not subject to much selective pressures that can produce homoplasy, or lead to adaptive divergence (Feng et al., 2005). However, Holcroft (2004) acknowledged that given the special properties of the RAG1 gene, careful considerations should be made regarding those species that show apparent departure from base stationarity, level of codon bias, and site saturation, which can lead to false classification of taxa under study.

Nuclear genes, such as RAG1, are recognized for their capacity in detecting historical patterns of population structure and reconstructing deep-level phylogenies of vertebrates (Williams et al., 2002; Groth and Barrowclough, 1999; Holcroft, 2004; Springer et al., 2001). However, it has not been used for inferring intrafamilial phylogenies except in one successful case in combination with other genes (Rüber et al., 2004). Mahon (unpublished PhD thesis) was successful in using RAG1 to infer intrafamilial relationships in another percoid family, Sparidae. This prompted the present study to use RAG1 to infer an intrafamilial phylogeny in Haemulidae, which is putatively closely related to Sparidae.

In this study, the nuclear gene RAG1, mitochondrial gene COI, and RAG1-COI combined sequence data were used to: (1) infer the phylogenetic relationship of the haemulids; (2) test the monophyly of the family Haemulidae and the proposed superfamily Haemuloidea; (3) examine if the putative haemulid genera defined by morphological characters are valid; and (4) to test if the RAG1 gene is useful in inferring phylogeny of genera within a percoid family.

MATERIALS AND METHODS

Methodology followed the standard molecular techniques from extraction (DNeasy® Tissue Handbook 03/2004) to sequencing PCR reactions (QIAquick® Spin Handbook 07/2002).

Chemicals

The chemicals listed in Table 2 were used for DNA extraction, electrophoresis, DNA amplification, clean-up, and sequencing reactions.

Table 2
Chemicals used and product information

Chemical	Product information
Agarose GPG/LE (American Bioanalytical)	Ultra pure; CAS 9012-36-6, AB00972-00500; American Bioanalytical, 15 Erie Drive, Natick, MA 01760 (www.americanbio.com)
Boric acid	A74-3; Fisher Scientific, Fair Lawn, New Jersey
Disodium Ethylenediamine Tetraacetate	S311-3; Fisher Scientific, Fair Lawn, New Jersey
Eco130I (Styl) (Buffer Y ⁺ /Tango™ with BSA)	MBI Fermentas, #ERO411 (www.fermentas.com)
Ethyl alcohol USP	190 proof and 200 proof%; DSP-KY-417; Aaper Alcohol and Chemical Co., Shelbyville, Kentucky
Hi-Di™ Formamide (Genetic Analysis Grade) (Cycle sequencing mix, pGEM Control Template, 21 M13 Control Primer)	Applied Biosystems, 850 Lincoln Centre Drive, Foster City, California 94404 (www.appliedbiosystems.com)
Isoamyl alcohol	Sigma Chemical Co., St. Louis, Missouri
PCR and Sequencing Primers	IDT, Inc., 1710 Commercial Park, Coralville, IA 52241 (www.idtdna.com)
Phenol chloroform	Sigma Chemical Co., St. Louis, Missouri
Proteinase K	Sigma Chemical Co., St. Louis, Missouri
Qiagen DNeasy® Tissue Kit	Qiagen Inc., 27220 Turnberry Lane, Suite 200, Valencia, California 91355 (www.qiagen.com)
QIAquick Gel Extraction Kit (250)	Qiagen Inc., 27220 Turnberry Lane, Suite 200, Valencia, California 91355 (www.qiagen.com)
Sodium Dodecyl Sulphate (SDS)	Fisher Scientific, Fair Lawn, New Jersey
Sodium Acetate	Sigma Chemical Co., St. Louis, Missouri
TaKaRa ExTaq™ (10X PCR Buffer, dNTP Mixture for PCR)	Takara Bio Inc. Code No. RR001A; Takara Mirus Bio 510 Charmany Drive Madison, Wisconsin 53719 (www.takara-bio.co.jp)
Trizma® base, minimum 99.9% titration	T1503-500g; Sigma-Aldrich Chemical Co., St. Louis, MO (www.sigma-aldrich.com)

Equipment

A thermal cycler (TC-312; Techne Incorporated, 3 Terri Lane, Suite 10 Burlington, NJ 08016, USA) was used for amplification and sequencing programs.

Electrophoresis made use of Wide Mini-Sub Gel GT and Power PAC 300 (BioRad, Hercules, CA). Visualization of agarose gels were performed using UV light (Spectroline® Model TR-302 transilluminator, 302nm UV) following electrophoresis. Sequencing products were sequenced using ABI 3100 capillary sequencer (Applied Biosystems). Other equipment included: Dry bath incubator (Fisher Scientific), Fisher Stirrer (Fisher Scientific), and Eppendorf Centrifuge 5415 C (Brinkman Instruments Inc., Rexdale, Ontario).

Specimen Samples

A total of 35 species of Haemulidae were used in this study (Table 3). These tissues were collected fresh from fish markets, trawl-caught, or speared. Muscle tissue or gill clippings of the fish were dissected and stored in a microcentrifuge tube filled with 95% ethanol or 40 to 70% ethanol, whichever was available during field collection or allowed for air shipping. Tubes were sealed with parafilm and were labeled accordingly by locality, date, and catalogue or field number, and logged in field notebooks. The ethanol was replaced new 95% ethanol solution when the tissues were received at the laboratory. Tissues were stored at -20°C until processed in the laboratory.

DNA Extraction

Genomic DNA was extracted from approximately 25 mg tissue using the DNeasy® Kit (Qiagen) protocol and phenol-chloroform techniques. Phenol-chloroform DNA extraction was adapted from Hillis et al. (1996). Using sterilization technique, 25 mg of tissue, cut into smaller pieces, was taken from each sample specimen and placed in

Table 3

List of species and the accession number of haemulid specimens

Species	Accession Number	Locality
<i>Anisotremus dovii</i>	ODU-305, 9659	Panama
<i>Anisotremus interruptus</i>	ODU-294, 9691	Panama
<i>Anisotremus pacifici</i>	ODU-315, 9618; ODU-317	Panama
<i>Anisotremus virginicus</i>	ODU-653; (577/F12)	Belize; sequence from A. Mahon
<i>Brachydeuterus auritus</i>	ODU-641A	Angola; Sequence from A. Mahon
<i>Conodon serrifer</i>	ODU-591, 9628	Panama
<i>Diagramma pictum</i>	ODU-267, RP-2026; ODU-280, PG30, 10/05/99 KECarpenter	Philippines
<i>Genyatremus luteus</i>	ODU-679, 1	KECarpenter
<i>Haemulon aurolineatum</i>	ODU-290, NMFS030; ODU-299, NMFS028; ODU-300, NMFS027; ODU-301, NMFS026	Sequence from Dr. G. Orti
<i>Haemulon flaviguttatum</i>	ODU-297, 9694	KECarpenter
<i>Haemulon plumierii</i>	ODU-647; ODU-649; (594/101,102)	Sequence from Dr. G. Orti
<i>Haemulon sciurus</i>	ODU-282, NMFS018; ODU-283, NMFS017; ODU-285; ODU-286; ODU-287	KECarpenter
<i>Haemulon scudderii</i>	ODU-307, 9663	Panama
<i>Haemulon steindachneri</i>	ODU-308, 9667; ODU-309, 9671	Panama
<i>Haemulopsis axillaries</i>	ODU-312, 9633	Panama
<i>Haemulopsis elongatus</i>	ODU-298, 9695	Panama
<i>Haemulopsis leuciscus</i>	ODU-318, 9612; ODU-319, 9611	Panama
<i>Haemulopsis nitidus</i>	ODU-320, 9602	Panama
<i>Inermia vittata</i>	T-329	Sequence from A. Mahon
<i>Orthopristis chalceus</i>	ODU-306, 9660; ODU-314, 9619	Panama
<i>Parapristipoma octolineatum</i>	ODU-274, 2A10	Angola
<i>Parapristipoma trilineatum</i>	KECarpenter	KECarpenter
<i>Plectorhinchus chaetodonoides</i>	ODU-275, 153	Manila
<i>Plectorhinchus lessonii</i>	ODU-276, 127; BUS-27; 240	Philippines
<i>Plectorhinchus macrolepis</i>	GG-045	Togo
<i>Plectorhinchus sordidus</i>	ODU-279, PG19, 10/03/99 KECarpenter	Kuwait
<i>Plectorhinchus vittatus</i>	ODU-292, RP01-006	Philippines
<i>Pomadasys argenteus</i>	ODU-335, T-291 BUS03	Philippines
<i>Pomadasys argyreus</i>	ODU-268, 07/27/02 KECarpenter	KECarpenter
<i>Pomadasys branickii</i>	ODU-304, 9655	Panama
<i>Pomadasys kaakan</i>	ODU-269, 07/27/01 KECarpenter; ODU-278, PG8, 10/03/99 KECarpenter	Kuwait
<i>Pomadasys maculatus</i>	ODU-810, RP01-055 (ODU-239)	Philippines
<i>Plectorhinchus shotaf</i>	ODU-95, T-304 BUS03	Philippines
<i>Xenichthys xanti</i>	ODU-264, 09/06/00; ODU-265, 09/06/00; ODU- 281, 05/06/00	KECarpenter
<i>Xenistius californiensis</i>	ODU-678; SIO 02-1	KECarpenter

a 0.5 mL microcentrifuge tube. The following were then added to the tube and incubated for a minimum of two hours at 58°C or left overnight at 37°C to accomplish cell lyses: 100 µL of STE buffer (prepared from 0.1 M NaCl, 0.05 M Tris-HCl, and 0.001 M EDTA disodium), 7.5 µL SDS (20% w/v in water), and 7.5 µL proteinase K (10 mg/mL proteinase K in STE buffer). After the tissue dissolves, 100 µL of PCI (phenol:chloroform: isoamyl alcohol, 25:24:1) was added into the tube, vortexed, and left at room temperature for five minutes. The tube was then centrifuged for five minutes at

maximum speed (14,000 rpm). It is important to check the color of PCI and to make sure it remains clear, otherwise a new PCI mix would need to be prepared. The aqueous phase (top layer) was then pipetted carefully, placed in a new tube, and 100 μ L CI (chloroform: isoamyl alcohol, 24:1) was added. The sample was vortexed and centrifuged for five minutes at maximum speed. The top layer was carefully removed, placed in a new tube, and 10 μ L of 3 M sodium acetate ($1/10^{\text{th}}$ of original volume) and 250 μ L of 100% ethanol (2.5 times of original volume) were added. This tube was placed in the freezer for a minimum of two hours. The samples were centrifuged immediately for five minutes after being removed from the freezer. The liquid was then removed by pipette, making sure the pellet at the bottom is not disturbed. Ethanol (300 μ L, 70%) was added to the sample and was vortexed for five minutes. The liquid was removed and samples were placed in a heat-block to dry for approximately one hour at 58°C. Thirty microliters of sterile distilled water was then added to the tube, vortexed, and stored in -20°C until DNA extract was used in the laboratory. DNA extraction following purification of total DNA from animal tissues (DNeasy[®] Tissue Handbook 03/2004) also required approximately 25 mg tissue sample with addition of 180 μ L Buffer ATL and 20 μ L proteinase K. The tube was vortexed and incubated at 55°C until completely lysed. It was necessary to vortex the tube occasionally to allow the sample to disperse. The tube was then vortexed (approximately 15 seconds) and 200 μ L Buffer AL was added before incubating again for 10 minutes at 70°C. After incubation, 200 μ L of 95% ethanol was added to the sample and mixed. The samples were then transferred to DNeasy[®] Mini spin column (provided in Qiagen Kit) and centrifuged for one minute. Several steps of centrifugation were carried out by adding 500 μ L Buffer AW1 for one minute at maximum speed and 500 μ L

Buffer AW2 for three minutes at maximum speed to ensure that no residual ethanol was carried over in the eluate. The spin column was then transferred to a new 1.5 mL microcentrifuge tube and the sample was eluted with 200 μ L Buffer AE. Sample was incubated at room temperature for one minute and centrifuged for another minute to get the final DNA yield. Successful DNA extractions both from Qiagen DNA extraction protocol and phenol-chloroform techniques were checked using (agarose) gel electrophoresis.

Gel Electrophoresis

Gel electrophoresis was used to check if DNA extraction was carried out successfully and if the desired band during PCR was amplified. It was also necessary to purify the PCR product using this technique followed by gel extraction protocol to eliminate problems with primer dimers during PCR. A 5X Tris-borate EDTA (TBE) buffer was prepared as a stock solution to further make 0.5X TBE buffer for use in gel electrophoresis. The stock (10X TBE) buffer was made by mixing 54 g Trizma[®] base, 27.5 g boric acid, and 4.65 g sodium EDTA, and adding deionized distilled water to 1 L using a stirrer magnet. The electrophoresis 0.5X TBE buffer was made by measuring 100 mL of 5X TBE buffer and mixing with 900 mL (natural) water. Agarose gels were prepared by measuring 1 g of agarose and adding 100 mL of 0.5X TBE. The agarose solution was microwaved for 1.5 minutes and 10 μ L ethidium bromide was added to the agarose solution before pouring into a gel chamber to cool. The solidified gel was then placed on the electrophoresis chamber, making sure the chamber had enough 0.5X TBE buffer (marked on chamber). A tracking dye was added to each sample (2 μ L dye: 8 μ L

sample) before loading the samples on the gel. Ladders were placed at the beginning and at the end of the wells for reference. The ladder was prepared using Eco130I digest of Lambda DNA (www.fermentas.com). Electrophoresis was carried out using PowerPAC 300 (Bio-RAD) running 115 volts of current for 30 minutes. The gel was observed under a UV light using Spectroline® Model TR-302 transilluminator (302nm UV) following electrophoresis. Gel photos were taken using the same transilluminator covered with a photo imaging with ethidium bromide filtering film (Kodak Scientific Imaging Systems EDAS 120 Camera, 1D software (KDS1D) v.3.0.0).

DNA Amplification

Two sets of PCR were employed to successfully amplify approximately 1386 base pairs of RAG1 gene from DNA extracts. The nuclear RAG1 gene of the haemulids was PCR-amplified using Primers 2533F (5'- TGAGCTGCAGTCAGTACCATAAGAT GT -3') and 4078R (5'- TGAGCCTCCATGAACTTCTGAAGRTAYTT -3'). The PCR primers used were separately prepared by adding 90 µL sterile distilled water to 10 µL of each primer to make 10 µM PCR primer. PCR reagents for a 25 µL PCR product included: 14.375 µL sterile distilled water, 2.5 µL 10X Ex Taq™ Buffer, 2 µL dNTPs, 1 µL of each PCR primer, 0.125 µL Takara Ex Taq™ DNA polymerase (Takara Bio Inc.), and 4 µL of DNA extract. The amplification condition was at 95°C for one minute initial denaturation; 15 cycles of 95°C for one minute, 53°C for 45 seconds, and 72°C for 1.5 minutes; 15 more cycles of 95°C for 45 seconds, 51°C for 45 seconds, and 72°C for 1.5 minutes; followed by a seven minute extension at 72°C and incubation at 10°C. PCR products were immediately run on a gel electrophoresis to further prevent problems

caused by primer dimers remaining after amplification. PCR products were run on an agarose gel and purified by gel extraction following QIAquick gel extraction protocol from Qiagen. To ensure that enough amount of amplified gene of DNA material can be used for sequencing reactions, the standard protocol of running 25 μ L of PCR reaction was carried out first before running another 100 μ L PCR reaction (re-PCR). The following cocktail for a 100 μ L re-PCR included: 71.5 μ L sterile distilled water, 10 μ L 10X Ex Taq™ Buffer, 8 μ L dNTPs, 4 μ L of each primer, 0.5 μ L Ex Taq™ DNA polymerase, and 2 μ L of the purified PCR product. The same PCR program was used to run this reaction. Re-PCR products were then run on gel and purified using gel extraction following Qiagen protocol.

PCR Purification

To eliminate problems with primer dimers during DNA amplification of RAG1 gene, PCR products were run on a gel electrophoresis and recovery of PCR products from agarose gels was carried out following QIAquick gel extraction kit protocol from Qiagen. After electrophoresis, the desired bands of about 1386 base pairs (marked by RAG1 gene) from PCR products were excised from the gel with a clean sharp blade and each band was placed in a 1.5 mL microcentrifuge tube separately. Samples were weighed and Buffer QG was added to each tube in 3:1 (Buffer QG: weight of excised gel) volume. Samples were incubated at 50°C for 10 minutes or until gels have completely dissolved. To bind the DNA, each sample was transferred to a QIAquick spin column and was centrifuged for one minute at maximum speed. Centrifugation (maximum speed) steps were carried out by adding 0.5 mL Buffer QG (for one minute, to remove

traces of agarose), 0.75 mL Buffer PE (one minute), and additional one-minute centrifugations to remove residual ethanol. Each spin column was then transferred to a clean 1.5 mL microcentrifuge tube and eluted with 30 μ L sterile distilled water. Samples were incubated at room temperature for one minute and were centrifuged again for one minute. Purified samples were stored in approximately -20°C until used in the laboratory.

Sequencing

The sequencing reaction, for both forward and reverse sequences, was prepared by using 5.4 μ L sterile distilled water, 2 μ L of 5X BigDye® v3.1 sequencing buffer (included in BigDye® v3.1 Terminator Cycle Sequencing Kit), 2 μ L of sequencing primer (either forward or reverse primer), 8 μ L of purified re-PCR product, and 2.6 μ L BigDye® Terminator (Applied Biosystems). The sequencing reaction was carried with the following program: 25 cycles of 96°C for 30 seconds, 50°C for 15 seconds, and 60°C for four minutes. Unincorporated dyes were removed from sequencing reactions following ethanol and sodium acetate precipitation. The protocol included addition of 50 μ L 100% ethanol, 2 μ L of EDTA, and 2 μ L of 3 M sodium acetate to each tube; 20 minute incubation followed by 20 minute centrifugation (maximum speed). The liquid was removed and 70 μ L ethanol was added to each tube. Samples were left at room temperature for five minutes and were centrifuged for another five minutes. The liquid was removed with a pipette and incubated at 90°C for one minute. Dried pellets were stored in the freezer at approximately -20°C until used or re-suspended directly with 15 μ L Hi-Di formamide (Applied Biosystems), and incubated at 95°C. The samples were

immediately placed on ice after incubation and sequenced using ABI 3100 capillary sequencer (Applied Biosystems) following standard operating methods. Standard precautions such as sterilization techniques and use of positive and negative controls were followed.

Data Analyses

The forward and reverse sequences for each species were run in BLAST (Basic Local Alignment Search Tool) to confirm that sequences matched the RAG1 gene (Altschul et al., 1990). BLAST was also used to determine overlap of the forward and reverse sequences and generate 1386 base pairs of RAG1 gene sequence. Reverse complements of sequences were obtained using BCM Search Launcher (Smith et al., 1996). Fasta files for RAG1 sequences were generated using BioEdit (Hall, 2001). Multiple alignments of sequences were performed using ClustalX (Thompson et al., 1997). Fine-tuning of the aligned sequences was done by comparing the fasta files with peaks from the chromatogram sequence data. Multiple alignment parameters for ClustalX were performed using default settings (Hall, 2004). Samples of haemulid tissues were submitted to the Barcode of Life Project to obtain the COI sequence data, which was used to corroborate the RAG1 dataset. The aligned COI sequences (585 base pairs) were analyzed both independently and combined with the RAG1 sequences.

Data saturation, Maximum Parsimony (MP), and Maximum Likelihood (ML) analyses were performed using PAUP* version 4.0b10 (Swofford, 2002; Hall, 2004). Data saturation is a measurement of the expected number of changes or substitutions depending on the overall genetic distance. Data saturation was performed to examine if

the gene used was evolving too quickly for the phylogenetic level examined (Orrell, 2000). Saturation analysis included only the ingroup and was performed for total, first, second, and third codon positions for RAG1, COI, and RAG1-COI combined analyses. To estimate the phylogeny of haemulids, MP was run to search for the best tree with the shortest number of steps, or fewest series of mutations. MrModeltest2.0 (Nylander, 2004) was used to determine the best-fit model for use in ML analysis, which searches for the topology that maximizes the likelihood of observing the data given a model of evolution. Bootstrap analysis was conducted for both MP and ML. The analyses included sequences of an inermiid (*Inermia vittata*) considered in the same superfamily as the Haemulidae, and two sparid fishes, *Diplopodus bermudensis* and *Pagrus pagrus* that are proposed potential outgroups (Johnson, 1980). RAG1 sequences for the two sparids, *A. virginicus*, *B. auritus*, *P. macrolepis*, and *I. vittata* were supplied by Andrew Mahon (unpublished PhD thesis) while sequences for *H. aurolineatum* and *H. plumierii* were obtained from the existing RAG1 mega dataset (University of Nebraska-Lincoln, c/o Dr. Guillermo Orti). Resulting topology for MP and ML analyses were viewed using Treeview (Page, 1996).

RESULTS

The RAG1 gene has a total of 1386 characters (Appendix 1), of which 903 are constant, 199 are variable but parsimony uninformative, and 284 are parsimony informative. The COI gene has a total of 585 characters (Appendix 2), of which 372 are constant, 12 are variable but parsimony uninformative, and 201 are parsimony informative. The combined RAG1-COI genes (Appendix 3) exhibited 1308 constant characters, 208 parsimony uninformative, and 455 parsimony informative characters.

Plots of total number of substitutions as a function of sequence divergence (Tamura and Nei, 1993) of pooled transitions and transversions for RAG1, COI, and RAG1-COI gene sequence data are given in Figs. 1 to 6 (plots for first and second codon positions are not shown). Plots of pooled transitions and transversions for RAG1 and combined RAG1 and COI gene sequence data show a positive relationship and appear to increase linearly with sequence divergence. Plots for COI, however, follow a scattered nonlinear relationship, with a large number of substitutions occurring both at low and high genetic distance for third codon position. Therefore, in Figs. 3 and 4, the third codon position data is saturated for both transitions and transversions for COI gene sequence data.

The general, time reversible plus proportion invariant with gamma distribution (GTR+I+G) model of evolution was found to be the best substitution model for each of the RAG1, COI, and RAG1 and COI combined data sets using MrModeltest v2.2 (Nylander, 2004). The estimated nucleotide frequencies for RAG1 data were: A=0.26100, C=0.24590, G=0.27480, T=0.21830; for COI A=0.25790, C=0.33100,

G=0.13680, T=0.27430; and for RAG1-COI A=0.25280, C=0.27350, G=0.24000, T=0.23370. The proportion of invariable sites was estimated to be 0.33 for RAG1, 0.63 for COI, and 0.46 for RAG1-COI. The shape of gamma parameter was estimated to be 0.76 for RAG1, 1.03 for COI, and 0.40 for RAG1-COI dataset. The model of nucleotide evolution was incorporated in PAUP* using the ML optimality criterion. The resulting topology for MP and ML analyses were tested using 100 bootstrap replicates.

For the RAG1 dataset, both MP and ML cladograms show high node support for a monophyletic Haemuloidea (Figs. 7 and 8). However, in MP, *Inermia* resulted as an outgroup in relation to haemulids, while ML recovered *Inermia* as an ingroup within the haemulids. Both topologies produced are well supported, therefore, no conclusive evidence was found to elucidate the relationship of the inermiid with the haemulids. Also, no COI sequence was available for *Inermia* to corroborate this result. RAG1, COI, and RAG1-COI sequence data all recovered a well-supported monophyletic Haemulidae for all MP and ML analyses conducted (Figs. 7 to 12). Since MP and ML cladograms produced similar components of clades in the case of RAG1 dataset, COI dataset, and RAG1-COI dataset, only the ML cladograms will be used to highlight subclades produced from RAG1, COI, and RAG1-COI dataset.

RAG1 recovered two well-supported clades separating the two subfamilies Haemulinae (Clade A) and Plectorhinchinae (Clade B) (Fig. 8). The Plectorhinchinae clade consists of *Parapristipoma octolineatum*, *P. trilineatum*, *Diagramma pictum*, *Plectorhinchus chaetodonoides*, *P. vittatus*, *P. macrolepis*, and *P. schotaf*. The Haemulinae clade produced six distinct well-supported subclades (Fig. 8). Clade 1 is

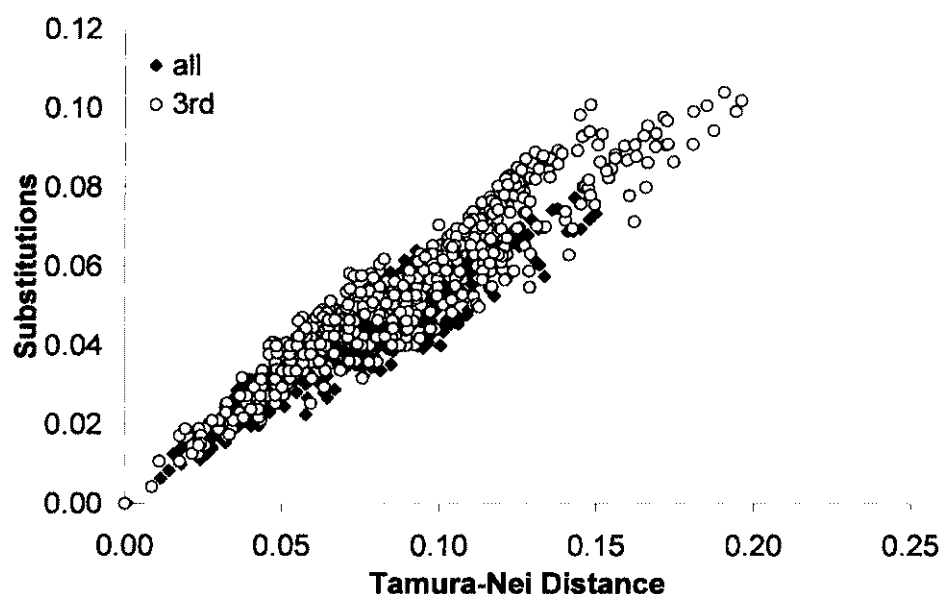


Fig. 1. Total number of substitutions plotted as a function of percent sequence divergence of pooled transitions for RAG1 sequence data. The shaded diamonds represent transitions at all positions, while the non-shaded circles represent transitions at third codon position.

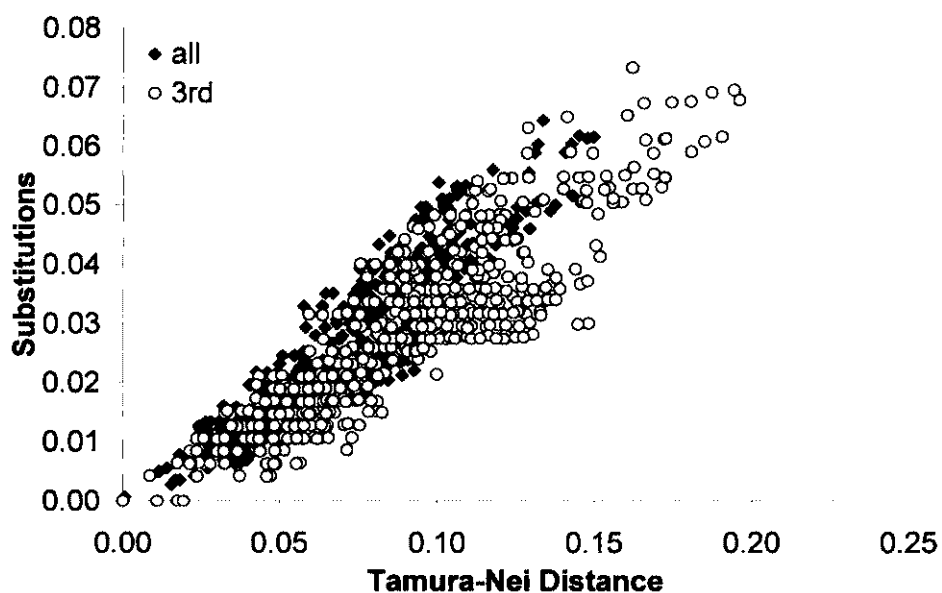


Fig. 2. Total number of substitutions plotted as a function of percent sequence divergence of pooled transversions for RAG1 sequence data. The shaded diamonds represent transversions at all positions, while the non-shaded circles represent transversions at third codon position.

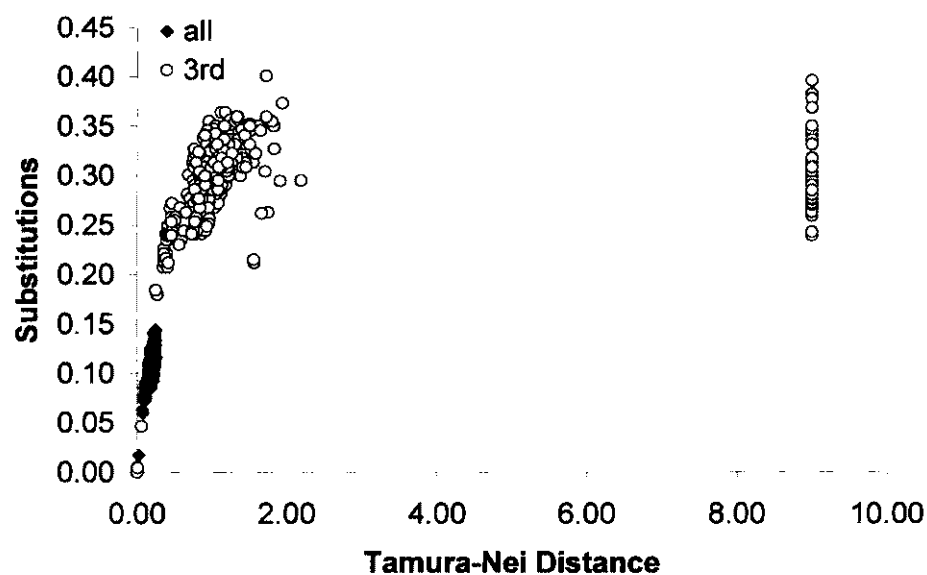


Fig. 3. Total number of substitutions plotted as a function of percent sequence divergence of pooled transitions for COI sequence data. The shaded diamonds represent transitions at all positions, while the non-shaded circles represent transversions at third codon position.

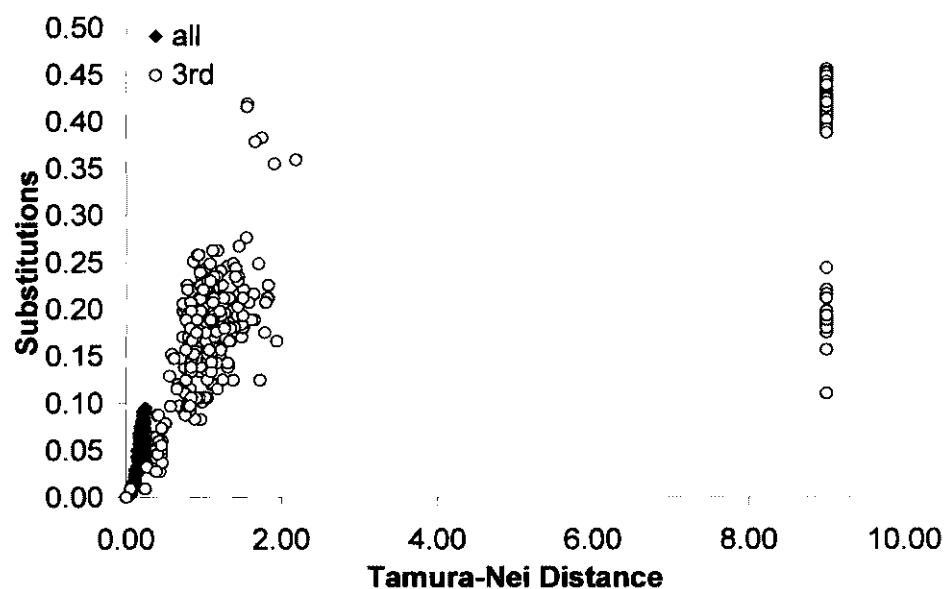


Fig. 4. Total number of substitutions plotted as a function of percent sequence divergence of pooled transversions for COI sequence data. The shaded diamonds represent transversions at all positions, while the non-shaded circles represent transversions at third codon position.

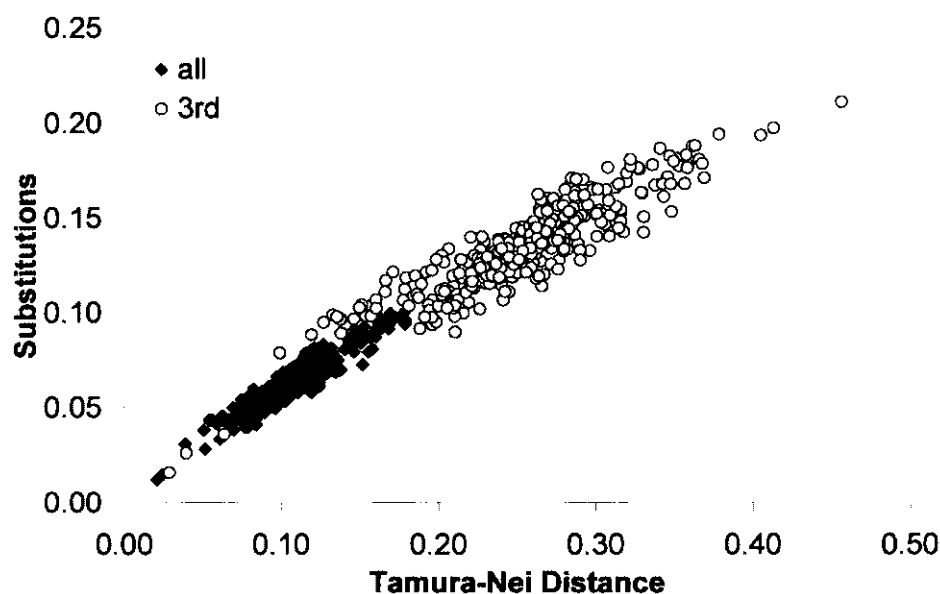


Fig. 5. Total number of substitutions plotted as a function of percent sequence divergence of pooled transitions for RAG1-COI sequence data. The shaded diamonds represent transitions at all positions, while the non-shaded circles represent transversions at third codon position.

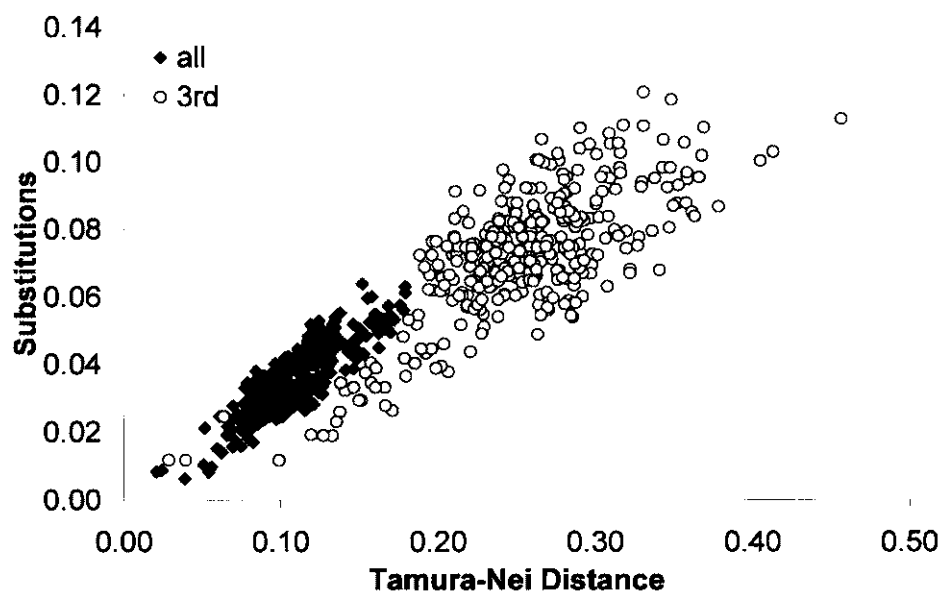


Fig. 6. Total number of substitutions plotted as a function of percent sequence divergence of pooled transversions for RAG1-COI sequence data. The shaded diamonds represent transversions at all positions, while the non-shaded circles represent transversions at third codon position.

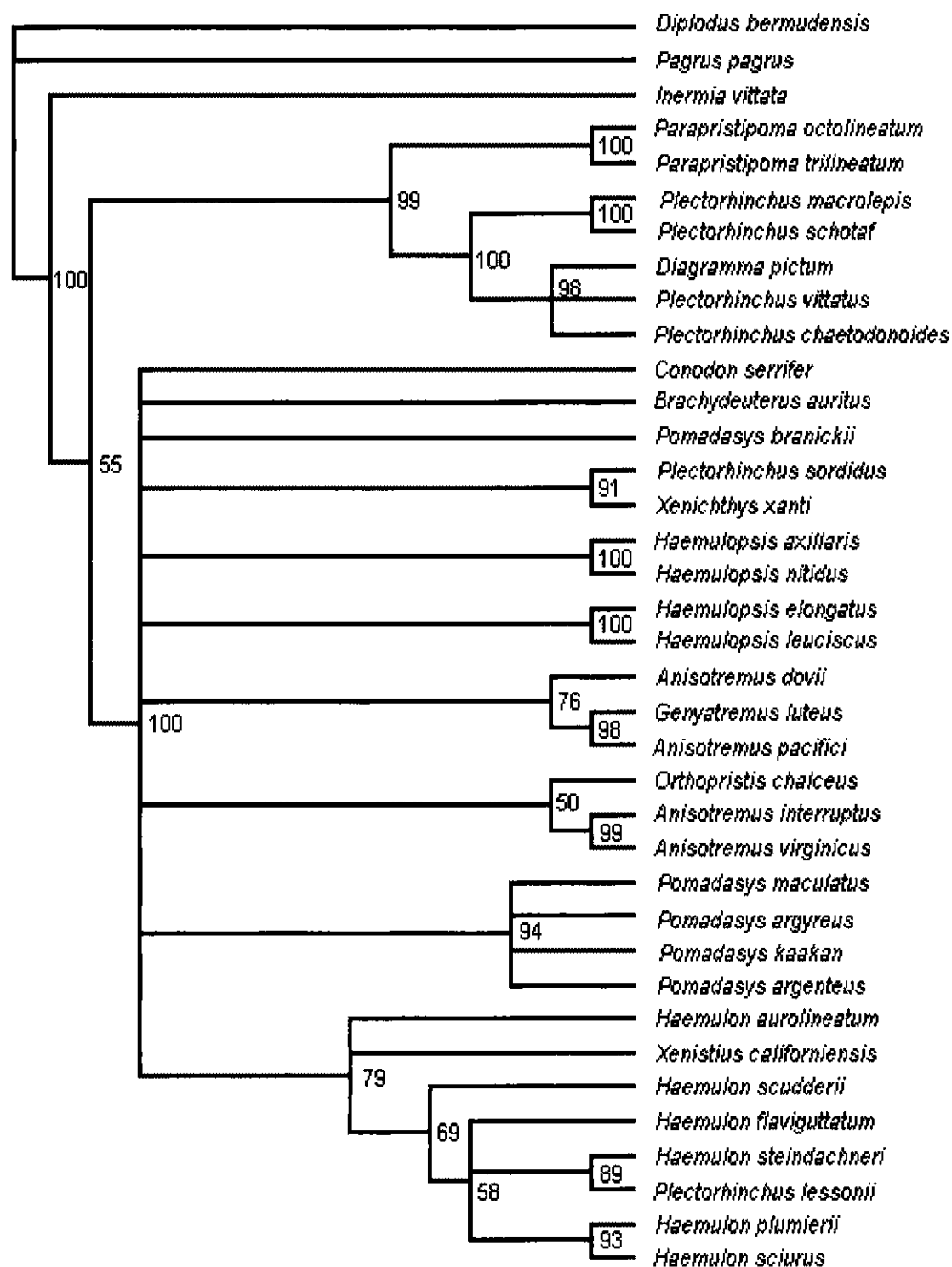


Fig. 7. Maximum Parsimony cladogram with nodes collapsed with less than 50% bootstrap support and bootstrap values of 50 and above indicated at the respective nodes for RAG1 sequence data.

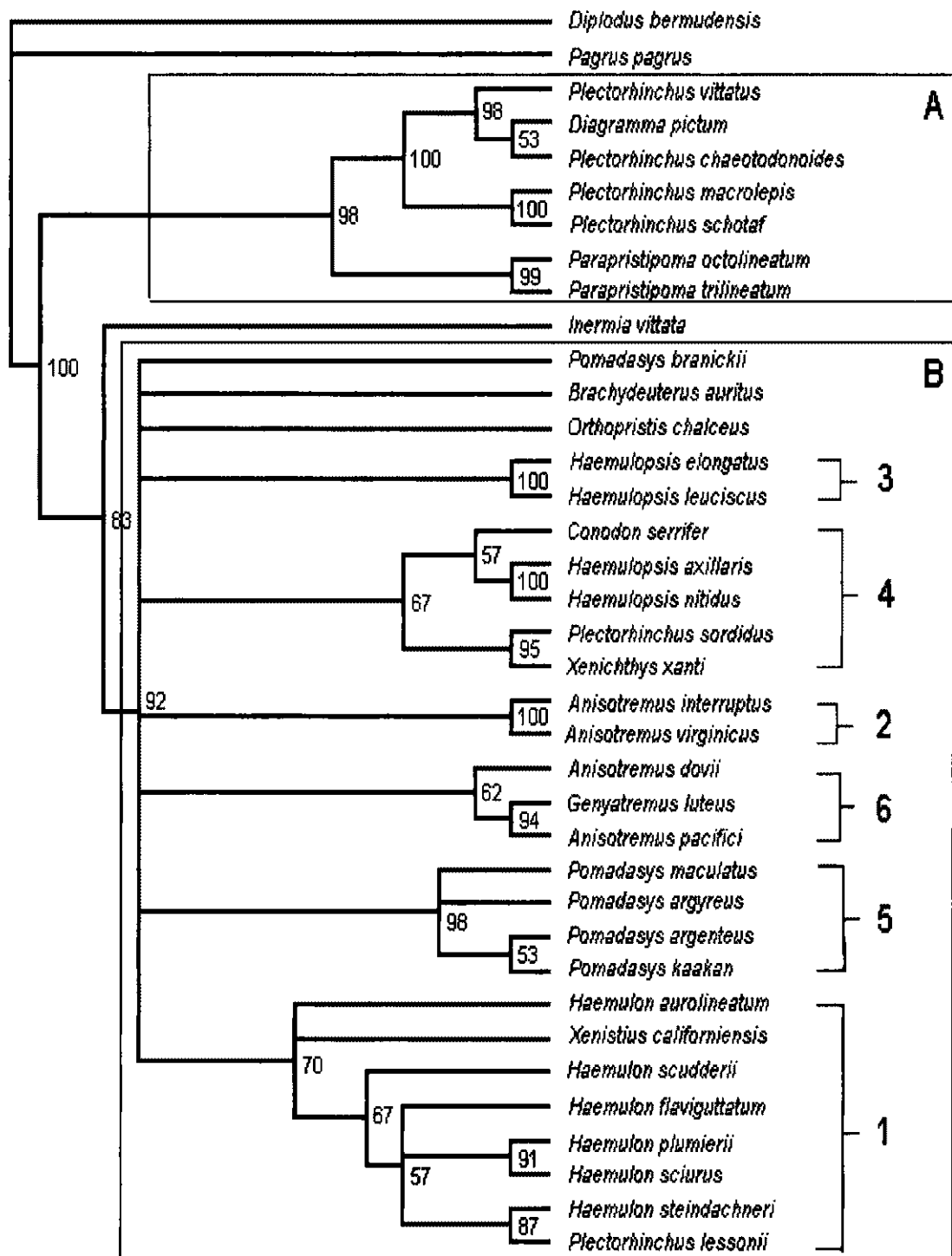


Fig. 8. Maximum Likelihood cladogram with nodes collapsed with less than 50% bootstrap support and bootstrap values of 50 and above indicated at the respective nodes for RAG1 sequence data.

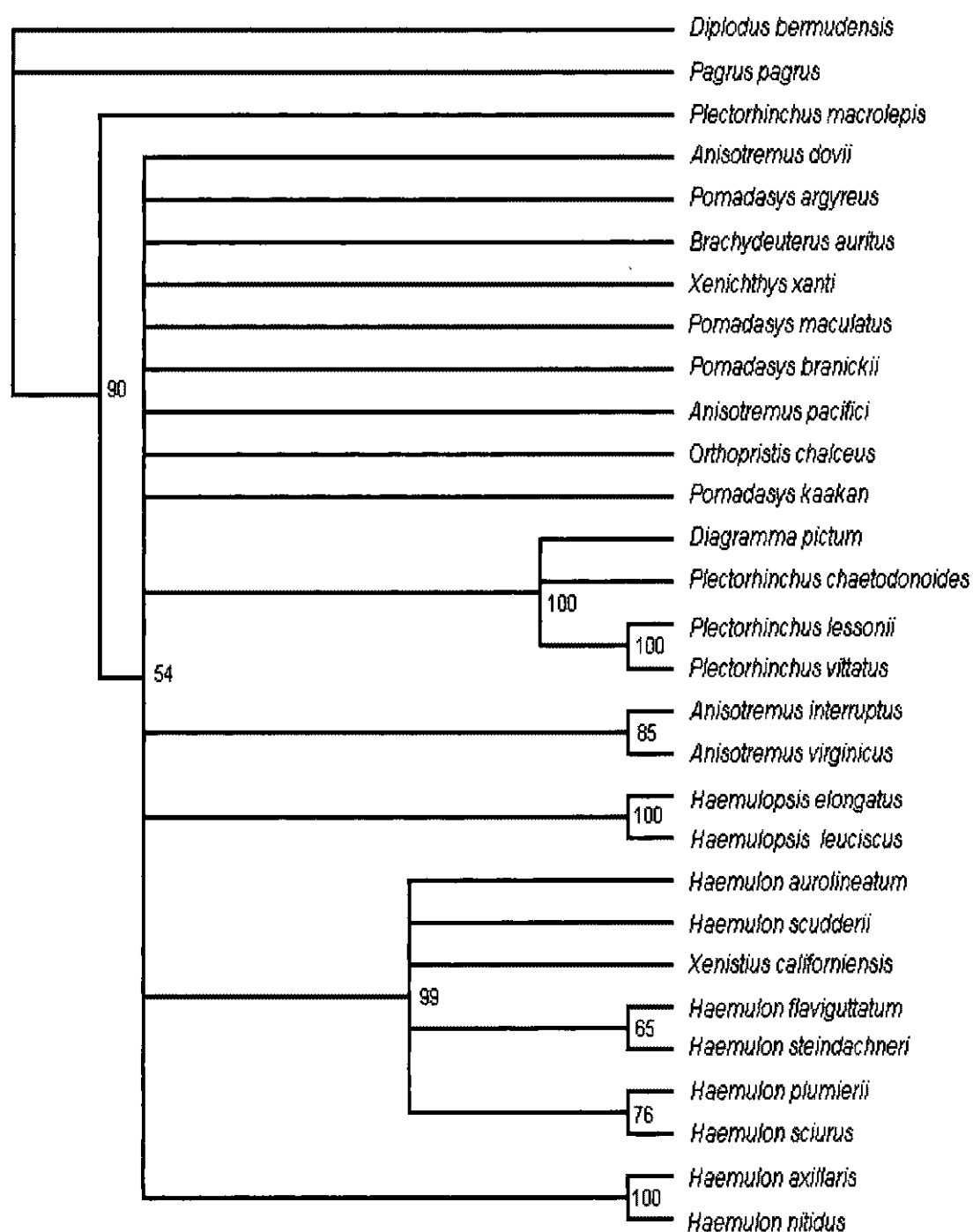


Fig. 9. Maximum Parsimony cladogram with nodes collapsed with less than 50% bootstrap support and bootstrap values of 50 and above indicated at the respective nodes for COI sequence data.

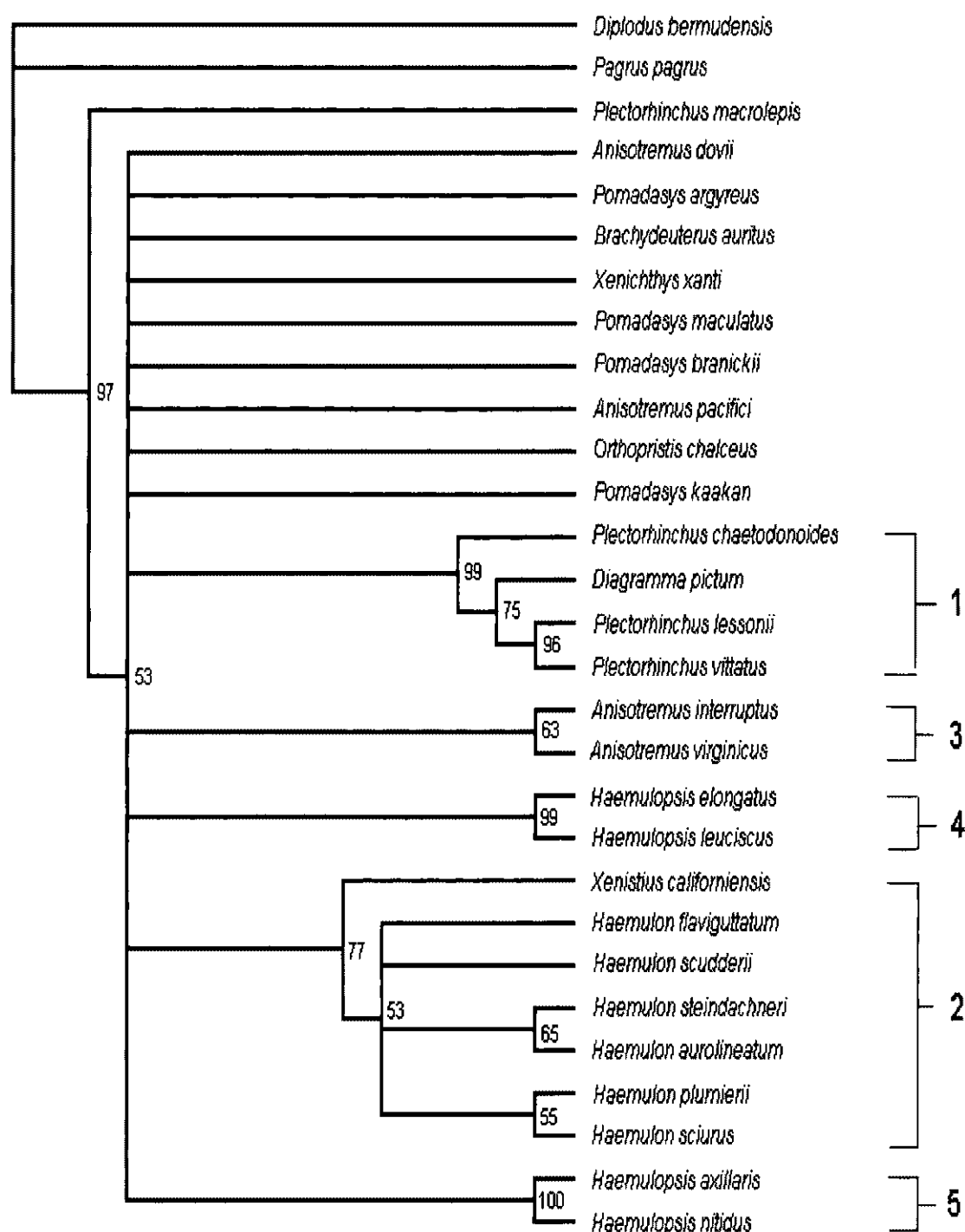


Fig. 10. Maximum Likelihood cladogram with nodes collapsed with less than 50% bootstrap support and bootstrap values of 50 and above indicated at the respective nodes for COI sequence data.

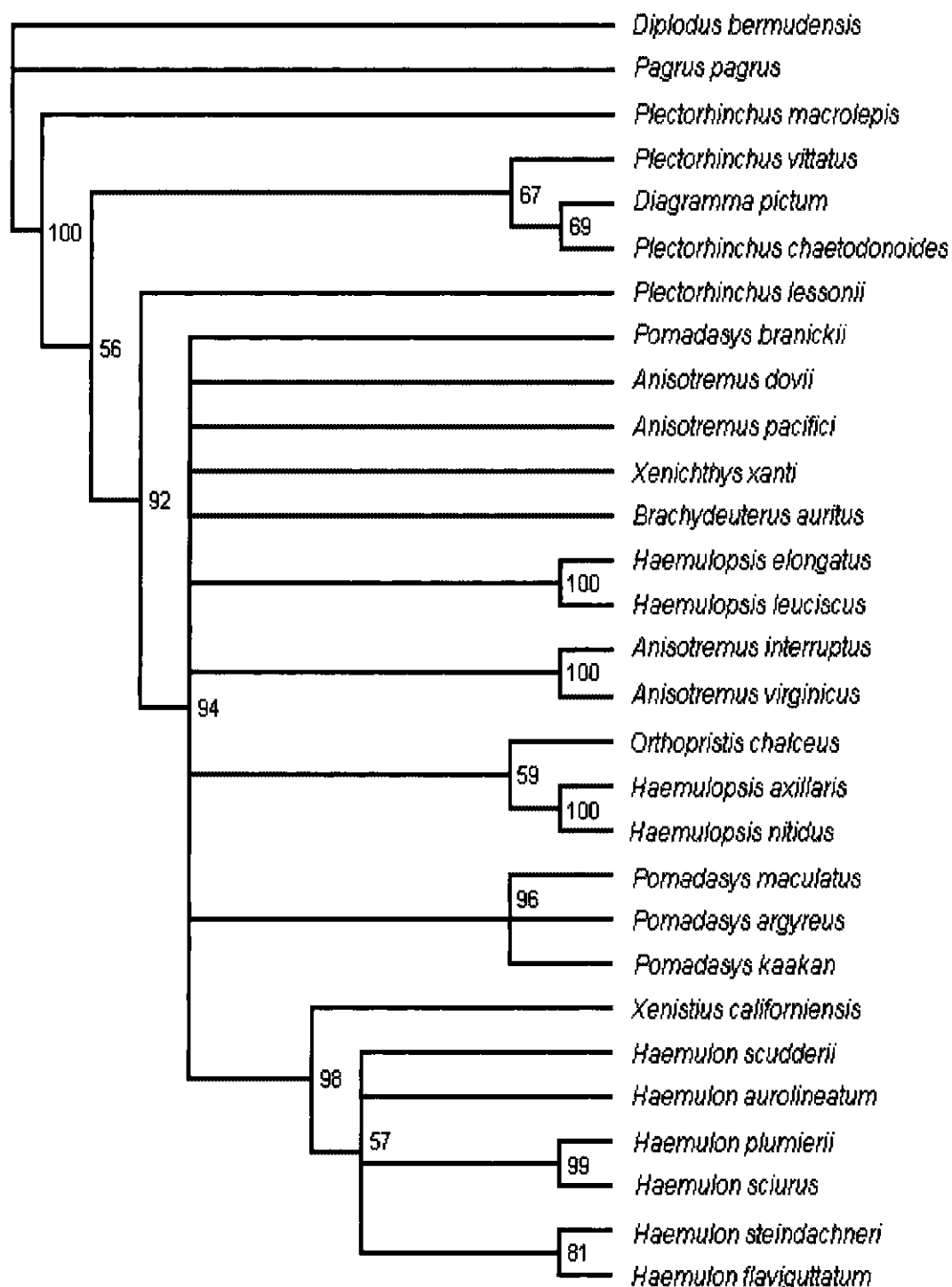


Fig. 11. Maximum Parsimony cladogram with nodes collapsed with less than 50% bootstrap support and bootstrap values of 50 and above indicated at the respective nodes for RAG1-COI sequence data.

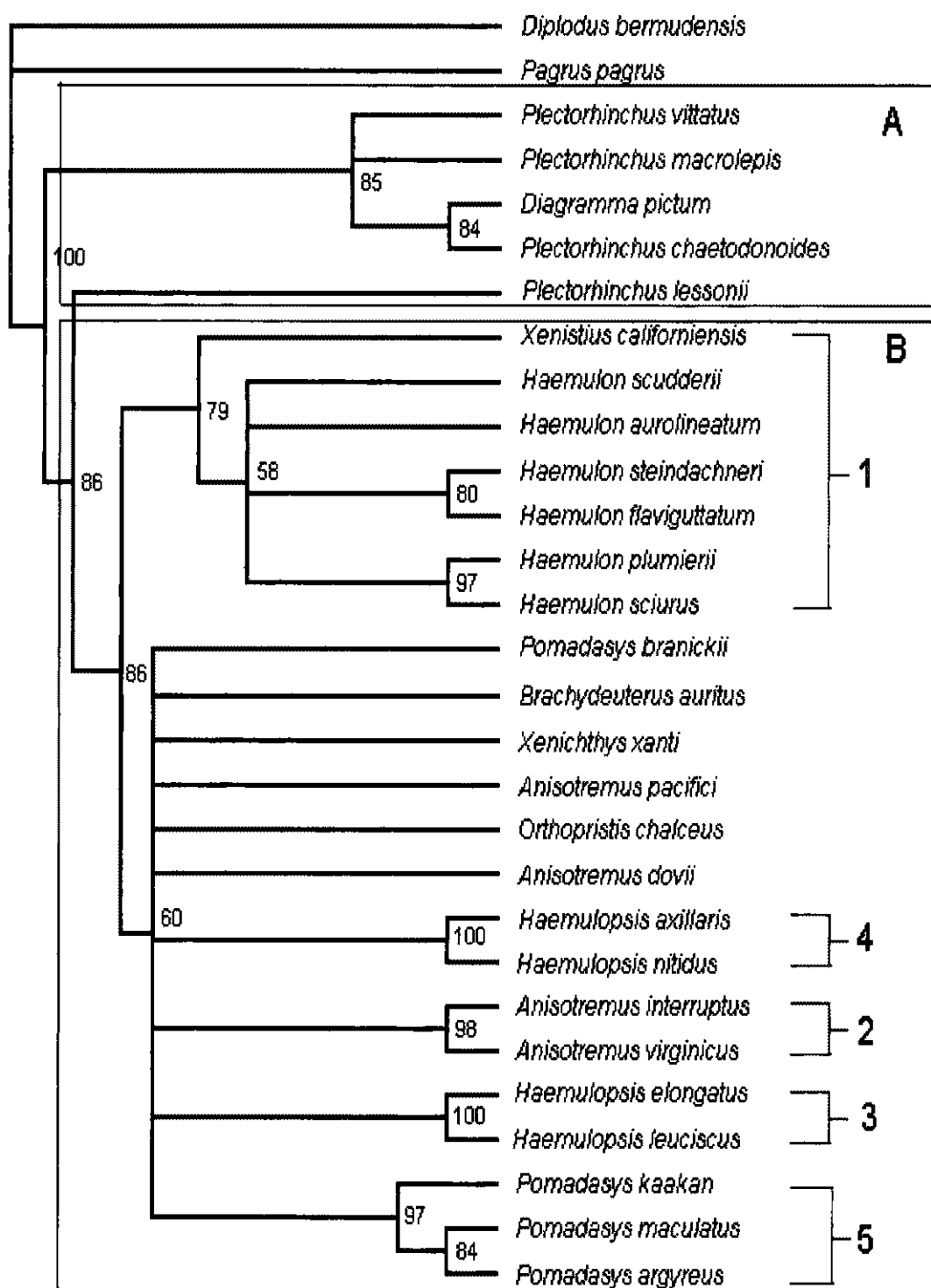


Fig. 12. Maximum Likelihood cladogram with nodes collapsed with less than 50% bootstrap support and bootstrap values of 50 and above indicated at the respective nodes for RAG1-COI sequence data.

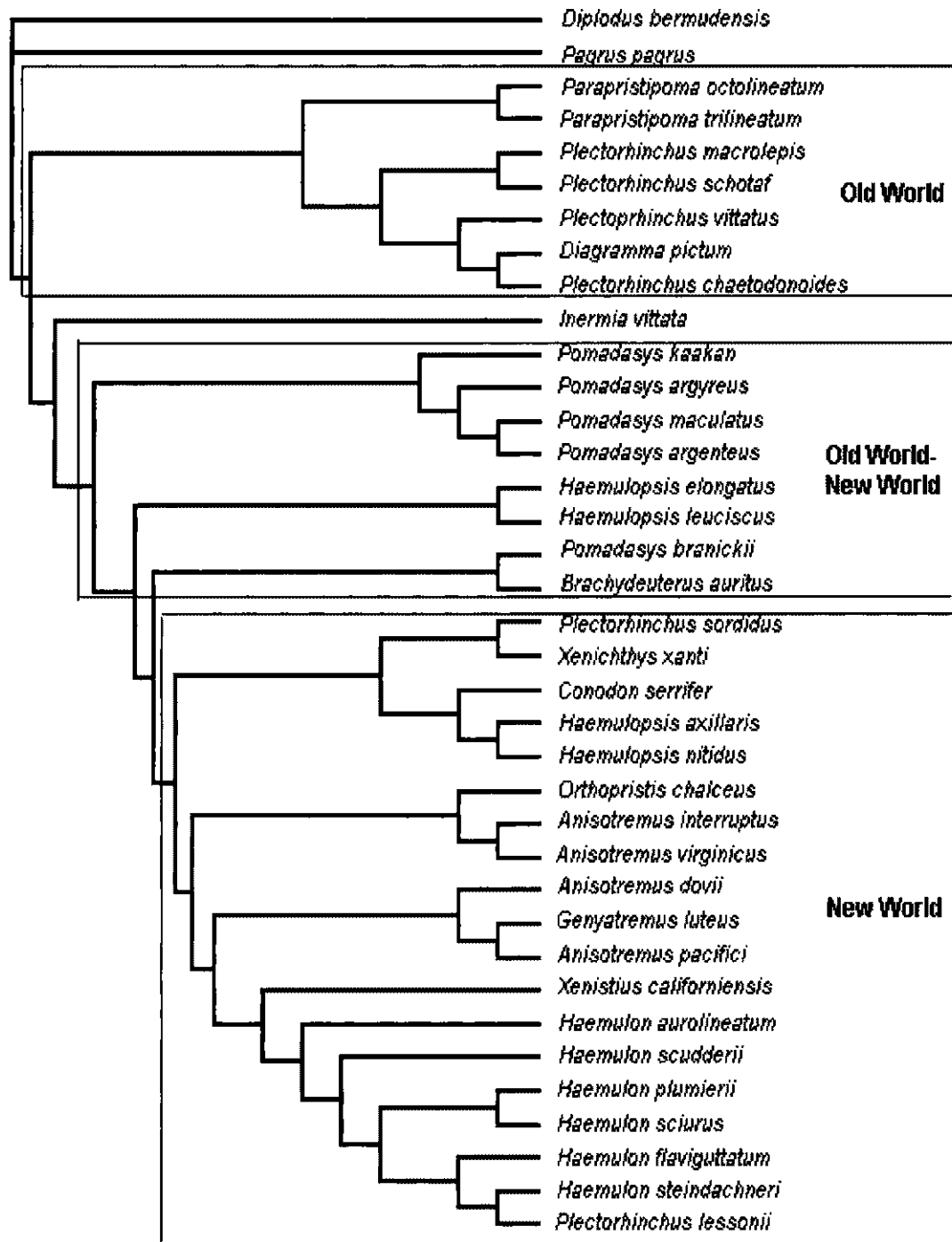


Fig. 13. Proposed radiation of the members of Haemulidae using Maximum Likelihood analysis for RAG1 sequence data. The basal clade is composed of Old World species. The intermediate clade is composed of both Old World and New World species. The more derived clade is composed of New World species.

composed of all the *Haemulon* species (*H. aurolineatum*, *H. scudderii*, *H. flaviguttatum*, and *Genyatremus luteus*. *Orthopristis chalceus*, *Brachydeuterus auritus*, and *P. branickii* resulted in separate branches within the major Haemulinae clade.

RAG1 recovered two well-supported clades separating the two subfamilies Haemulinae (Clade A) and Plectorhinchinae (Clade B) (Fig. 8). The Plectorhinchinae clade consists of *Parapristipoma octolineatum*, *P. trilineatum*, *Diagramma pictum*, *Plectorhinchus chaetodonoides*, *P. vittatus*, *P. macrolepis*, and *P. schotaf*. The Haemulinae clade produced six distinct well-supported subclades (Fig. 8). Clade 1 is composed of all the *Haemulon* species (*H. aurolineatum*, *H. scudderii*, *H. flaviguttatum*, *H. steindachneri*, *H. plumierii*, and *H. sciurus*) and is monophyletic with the inclusion of *Xenistius californiensis*. Clade 2 is a well-supported group composed of *Anisotremus virginicus* and *A. interruptus*. Clade 3 is composed of *Haemulopsis elongatus* and *H. leuciscus*. Clade 4 is composed of *H. axillaris* and *H. nitidus* and clumped together with *Conodon serrifer* and *Xenichthys xanti*. Clade 5 is composed of *Pomadasys maculatus*, *P. argyreus*, *P. argenteus*, and *P. kaakan*. Clade 6 is composed of *A. dovii*, *A. pacifici*, and *Genyatremus luteus*. *Orthopristis chalceus*, *Brachydeuterus auritus*, and *P. branickii* resulted in separate branches within the major Haemulinae clade.

Only five clades are recovered for COI cladograms (Fig. 10). One of the clades (Clade 1, Figure 10) is composed of members of the subfamily Plectorhinchinae, which are *Plectorhinchus chaetodonoides*, *P. lessonii*, *P. vittatus*, and *Diagramma pictum*. Clade 2 is composed of *Xenistius californiensis*, *Haemulon flaviguttatum*, *H. scudderii*, *H. steindachneri*, *H. aurolineatum*, *H. plumierii*, and *H. sciurus*. Clade 3 is composed of *Anisotremus interruptus* and *A. virginicus*. Clade 4 is composed of *Haemulopsis*

elongatus and *H. leuciscus*. Clade 5 is composed of *Haemulopsis axillaris* and *H. nitidus*. The RAG1-COI cladogram (Fig. 12) recovered the subfamilies Haemulinae and Plectorhinchinae, and produced five subclades. Clade 1 is composed of all *Haemulon* species and *X. californiensis*. Clade 2 is composed of *A. interruptus* and *A. virginicus*. Clade 3 is composed of *H. elongatus* and *H. leuciscus*. Clade 4 is composed of *H. axillaris* and *H. nitidus*. Clade 5 is composed of *P. maculatus*, *P. argyreus*, and *P. kaakan*.

A discrepancy between RAG1, COI, and RAG1-COI result is the placement of *Plectorhinchus lessonii* and *P. sordidus* in RAG1 cladograms. RAG1 placed *P. lessonii* and *P. sordidus* outside the clade Plectorhinchinae, while corroborated analyses using COI and RAG1-COI positioned *P. lessonii* close to Plectorhinchinae and outside Haemulinae. No COI sequence was available for *P. sordidus*.

Molecular data using RAG1, COI, and RAG1-COI combined genes consistently generated groupings that contradict the monophyly of three haemulid genera: *Anisotremus*, *Haemulopsis*, and *Pomadasys*. For the genus *Anisotremus*, two well-supported clades are produced and are both monophyletic with the inclusion of another genus: one clade includes *A. virginicus*, *A. interruptus*, and *Orthopristis chalceus*; the other clade includes *A. dovii*, *A. pacifici*, and *Genyatremus luteus*. Two clades are also recovered for the genus *Haemulopsis*, one clade is composed of *H. elongatus* and *H. leuciscus*, while the other clade is composed of *H. axillaris*, *H. nitidus*, *Conodon serrifer*, and *Xenichthys xanti*. Two clades are also produced for the genus *Pomadasys*: one group is composed of *Pomadasys kaakan*, *P. argyreus*, *P. maculatus*, and *P. argenteus* and a separate branch for *P. branickii*.

The fully resolved cladogram produced from ML (Fig. 13) recovered groups that are consistent with Old World and New World distributions. The members of the subfamily Plectorhinchinae are primarily Old World species (Clade A, Figs. 8 and 13). Members of the subfamily Haemulinae (Clade B) are composed of both Old World and New World species (Figs. 8 and 13). The ML together (Fig. 13) with these Old World and New World designations may be indicative of radiation of the haemulids: basal Old World group (composed of members of the subfamily Plectorhinchinae); a derived New World group (all *Haemulon* species, all *Anisotremus* species, *Xenistius californiensis*, *G. luteus*, *O. chalceus*, *Conodon serrifer*, *Xenichthys xanti*, *Haemulopsis axillaris*, and *H. nitidus*); and an intermediate Old World-New World group, which is composed of both Old World and New World haemulids (all *Pomadasys* species, *Brachydeuterus auritus*, *Haemulopsis elongatus*, and *H. leuciscus*).

DISCUSSION

Comparison of the number of substitutions with genetic distance at all and each codon position suggested that there is no saturation that has occurred for RAG1 and combined RAG1 and COI dataset. Mutational saturation occurs if there is a deviation from a linear relationship between the number of nucleotide substitutions and the measure of sequence divergence or if data points for substitutions show a leveling off of change as sequence divergence increases (Holcroft, 2004). Figs. 3 and 4 indicate that saturation of third codon substitutions has occurred. Transitional substitutions at third codon position, however, can still be phylogenetically informative even when saturation is indicated (Källersjö et al., 1999; Folmer et al., 1994). COI gene is a mitochondrial gene that evolves very rapidly and contains the most variable sites particularly in third codon positions, which makes it very useful in population genetic studies. Therefore site saturation, especially at third codon positions, is expected for COI dataset at the phylogenetic level being examined in this study. The independent analysis using the COI gene, however, is a suspect using this dataset.

The MP and ML analyses produced mostly similarly distinct clades (Figs. 7 to 12). One exception was *Inermia vittata* that resulted as a haemulid outgroup in MP analysis and as an ingroup in ML analysis for RAG1 dataset. One possible reason for this might be that variation in the RAG1 gene is insufficient to clearly resolve this phylogenetic relationship. Both topologies are well supported by bootstrap values and, therefore, the phylogenetic relationship of inermiids with the haemulids is inconclusive

with this dataset. There was no available COI sequence data for *Inermia* that could potentially resolve its position in relation to haemulids.

The two subfamilies Plectorhinchinae and Haemulinae were recovered with high bootstrap support (Figs. 7, 8, 11, and 12) corroborating the previously proposed classification based on morphological data (Johnson, 1980). Although independent analyses using RAG1 gene and COI genes produced a separate clade for members of Plectorhinchinae, only the RAG1 gene recovered both a Plectorhinchinae clade and a Haemulinae clade (Figs. 7 and 8). However, one of the inconsistencies in this study was the placement of *Plectorhinchus lessonii* and *P. sordidus*, which were placed outside the Plectorhinchinae clade in the RAG1 dataset (Figs. 7 and 8). *P. sordidus* was placed in a clade composed of members of *Haemulopsis*, *Conodon*, and *Xenichthys*, while *P. lessonii* was clumped together with members of the genus *Haemulon*. Three potential explanations can account for this anomaly: (1) misidentification of the samples or specimens used, (2) both species may not belong in the genus *Plectorhinchus*, or (3) the RAG1 gene may not have enough variation and that a limited number of mutations made these species look genetically more similar to members outside their true clade. In the latter case, an artifactual phylogenetic grouping could be the result of random convergent or parallel changes in the RAG1 gene. A comparison of RAG1 versus COI results indicates that inherent invariability in the RAG1 gene is the probable cause of misplacement of *P. lessonii* and *P. sordidus*, rather than misidentification or incorrect taxonomic assignment. As a nuclear gene, RAG1 evolves more slowly than the COI mitochondrial gene. The analysis using the COI gene (amplified from the same source of fish tissue) and combined analysis using RAG1 and COI genes, placed *P. lessonii* with

other members of the genus *Plectorhinchus*. The added variability of the mitochondrial gene apparently was better capable of resolving this intrageneric phylogenetic relationship. This refutes the possibility of misidentification or incorrect taxonomy for *P. lessonii*. However, no COI sequence was available for *P. sordidus* to further test the position of this species within the haemulids. Placement of this species needs further evaluation.

One interesting result is a well-supported clade comprised of all *Haemulon* species in all the analyses conducted in this study (Figs. 7 to 12), and includes *Xenistius californiensis* most consistently as sister to *Haemulon*. Morphological examinations and inclusion of both species of *Xenistius* in a molecular analysis may further elucidate this relationship.

Molecular data using RAG1, COI, and RAG-COI combined genes consistently generated clades that question the monophyly of three haemulid genera: *Anisotremus*, *Haemulopsis*, and *Pomadasys* (Figs. 7 to 12). Two well-supported clades are produced for the genus *Anisotremus*: one clade includes *A. virginicus* and *A. interruptus* (and *Orthopristis chalceus* in MP cladogram in Fig. 7); the other clade includes *A. dovii*, *A. pacifici*, and *Genyatremus luteus*. The members of the genus *Anisotremus* has been described based only on various superficial characters. Some obvious morphological characters appear to corroborate the clades in the molecular analyses. For example, the suborbital distance (the distance between the eye and the mouth) of *A. dovii*, *A. pacifici*, and *G. luteus* is significantly shorter than the suborbital distance of *A. virginicus*, *A. interruptus*, and *O. chalceus*. Other morphological characters include the general body and fin configurations that are different between the two clades and similar within clades.

A. virginicus, *A. interruptus*, and *O. chalceus* display a deeper body configuration, with depths typically 30 to 40% of their body length, and are more elongated compared to members of the other clade, which are more ovate than oblong. In addition, *A. virginicus*, *A. interruptus*, and *O. chalceus* possess dorsal rays XII, 14 to 17; anal rays III, 8 to 11. *A. dovii* and *A. pacifici* both have dorsal rays IX, 13 to 16; anal rays III, 9 to 10, while *G. luteus*, has dorsal rays XIII, 12; anal rays III, 11. A complete morphological examination is beyond the scope of this thesis. However, the molecular evidence and superficial morphological evidence indicates that further comprehensive examination of the osteological and other morphological characters of the members of these genera may result in a revision of generic assignments within the Haemulidae.

The molecular evidence in this study indicates two strong sister group relationships within the genus *Haemulopsis*, whose four members are all represented in this study. A clade with *H. axillaris* and *H. nitidus* and a separate clade with *H. elongatus* and *H. leuciscus* are consistently recovered with strong nodal support from bootstrap values (Figs. 7 to 12). However, there is no support for a clade that includes all four species of *Haemulopsis* and further molecular and morphological information are needed to test the monophyly of this genus.

Results of MP and ML analyses using RAG1 question the monophyly of the genus *Pomadasys*. These analyses recovered a separate clade for *P. branickii* and another clade consisting of *P. maculatus*, *P. argenteus*, *P. kaakan*, and *P. argyreus* (Figs. 7 and 8). Superficial morphological differences are evident between *P. branickii* and the larger clade such as the size and length of the anal spine. In addition, *P. branickii* possess dorsal rays XIII, 11 to 12 while the other *Pomadasys* species have dorsal rays XII, 13 to

14. A more comprehensive examination of members of this genus using both molecular and morphological characters may help further resolve the generic limits of *Pomadasys*. There is also a potential biogeographic explanation for these differences that will be discussed later.

The phylogenies inferred from MP and ML analyses of RAG1 gave unexpected insights into the possible biogeographic radiation of haemulid clades of Old World (Indo-Pacific and Eastern Atlantic) and New World (both coasts of the Americas) species with respect to tectonic movements. All members of the subfamily Plectorhinchinae are Old World while the most derived members of Haemulinae are New World species (Fig. 13). As stated previously, the presence of two *Plectorhinchus* species in the New World clade is suspected to be an anomaly based on lack of variability in RAG1. An example of an Old World distribution within Plectorhinchinae is the well-supported clade composed of *Parapristipoma trilineatum* and *P. octolineatum*. This is a distinct genus within Plectorhinchinae and both species look very similar but are disjunctly distributed. *Parapristipoma trilineatum* is distributed in the Indo-West Pacific from southern Japan, East China Sea to Taiwan, while *P. octolineatum* occurs in the Eastern Atlantic region ranging from Angola northward and extending into the Western Mediterranean and along the coasts of Portugal and Spain. This genus possibly had a continuous Tethyan distribution prior to the closing of the Tethys Sea when Africa and the Middle East united with Eurasia near the Oligocene-Miocene boundary. Haemulids first evolved during the Cenozoic at a time when the Tethys was circumtropical and later ceased to exist (Hobson, 2006). Haemulids are coastal species and presumably require a more-or-less continuous coastline to become reproductively cohesive. These coastal fishes evolved in continuous

distribution during the period of a continuous Tethys Sea. When the Tethys Sea closed and the Atlantic Ocean widened, haemulids would have continued to evolve in allopatry. The Plectorhinchinae (*Plectorhinchus* and *Parapristipoma*) have members both in the Indo-Pacific and eastern Atlantic and presumably evolved prior to the closing of the Tethys but after the Atlantic Ocean widened to an extent that prevented migration across that ocean.

The derived species of Haemulinae are restricted to the New World and presumably evolved after the closing of the Tethys Sea and the widening of Atlantic Ocean (Fig. 13). All species belonging to this clade either occur in the Eastern Pacific or Western Central Atlantic and presumably evolved prior to the relatively recent closing of the Panamanian Isthmus (Bernardi and Lape, 2005). However, other haemulines, including *Haemulopsis elongatus*, *H. leuciscus*, *Brachydeuterus auritus*, *Pomadasys kaakan*, *P. argyreus*, *P. maculatus*, and *P. argenteus* are a paraphyletic group between Old World and New World groups and occur in both the Old and New Worlds. Both *H. elongatus* and *H. leuciscus* occurs in the Eastern Pacific while *B. auritus* occurs in the Eastern Atlantic (from Morocco to Angola). Moreover, members of the genus *Pomadasys* occur in two clades that correspond to New World and Old World distributions. Molecular data suggests a separate clade for *P. branickii* (New World) and another clade consisting of *P. maculatus*, *P. argenteus*, *P. kaakan*, and *P. argyreus* (all Old World). As stated above, there are morphological characters that indicate that *Pomadasys* needs revision with respect to its members. There are two other possibilities for these two separate *Pomadasys* clades. One of them is the inability of RAG1 gene to resolve this relationship and may not have enough variation to resolve the position of *P.*

branickii within the genus *Pomadasys*. The other possibility relates to the separation of the Old World and New World tectonic plates and the widening of the Atlantic Ocean that may have resulted in two separately evolving clades. Further sampling of *Pomadasys* species around the Atlantic is required to test this hypothesis. Alternatively the intermediate paraphyletic group composed of both Old World and New World species could potentially have been evolving as the Atlantic Ocean widened and prior to the closing of the Tethys Sea.

CONCLUSION

The RAG1 gene was found to be useful in inferring phylogeny at the intrafamilial level within the Haemulidae. The RAG1 gene provided evidence for a monophyletic Haemulidae and recovered well-supported clades for the subfamilies Plectorhinchinae and Haemulinae. The placement of *Inermia* in relation to haemulids, however, remains inconclusive using this dataset. Further molecular work and inclusion of another inermiid species may further elucidate this relationship.

The phylogenies inferred from MP and ML analyses of RAG1 gave unexpected insights into the possible biogeographic radiation of haemulid clades of Old World (Indo-Pacific and Eastern Atlantic) and New World (both coasts of the Americas) species, which can be accounted for by the closing of the Tethys Sea and the widening of the Atlantic Ocean. In addition, this study also highlighted potential problems regarding the validity of putative genera within the haemulids, including *Anisotremus*, *Haemulopsis*, and *Pomadasys*. Molecular data using RAG1, COI, and combined RAG1 and COI genes consistently generated clades that question the monophyly of three haemulid genera, and indicate that further comprehensive examination of the osteological and other morphological characters of the members of these genera may result in a revision of generic assignments within the Haemulidae. Further molecular phylogenetic studies that include species of *Hapalogenys*, *Microlepidotus*, *Parakuhlia*, *Boridia*, and *Isacia* may further elucidate the phylogeny of members of Haemulidae.

REFERENCES

- Allen, G.R., Robertson, D.R., 1994. Fishes of the Tropical Eastern Pacific. University of Hawaii Press, 2840 Kolowalu Street, Honolulu, Hawaii.
- Altschul, S.F., Gish, W., Miller, W., Myers, E.W., Lipman, D.J., 1990. Basic local alignment search tool. *J. Mol. Biol.* 215, 403-410.
- Bernardi, G., Lape, J., 2005. Tempo and mode of speciation in the Baja California disjunct fish species *Anisotremus davidsonii*. *Mol. Ecol.* 14, 4085-4096.
- Brown, R.W., 1956. Composition of scientific words: A manual of methods and a lexicon of materials for the practice of logotechnics. Smithsonian Institution Press, Washington, D.C.
- Cooke, R., 1992. Prehistoric nearshore and littoral fishing in the Eastern Tropical Pacific: An Ichthyological evaluation. *J. World Prehist.* 6, 1-49.
- Courtenay, W.R. Jr., 1961. Western Atlantic fishes of the genus *Haemulon* (Pomadasyidae): Systematic status and juvenile pigmentation. *Bull. Mar. Sci. Gulf & Carib.* 11, 66-149.
- Durán-González, A., García-Rucias, C.E., Laguarda-Figuerras, A., 1990. The karyotype and “G” bands of *Haemulon aurolineatum* Cuvier, 1829 (Pisces: Haemulidae). *Anales del Instituto de Ciencias del Mar y Limnología. Evol.* 12, 115–123.
- FAO., 2006. Capture production 1950-2004. FAO Fishery Information, Data and Statistics Unit: FISHSTAT Plus - Universal software for fishery statistical time series [online or CD-ROM]. Food and Agriculture Organization of the United Nations.

- Feng, B., Bulchand, S., Yaksi, E., Friedrich, R.W., Jesuthasan, S., 2005. The recombination activation gene 1 (RAG1) is expressed in a subset of zebrafish olfactory neurons but is not essential for axon targeting or amino acid detection. *BMC Neurosci.* 6, 46.
- Fish, M.P., Mowbray, W.H., 1970. Sounds of Western North Atlantic fishes. A reference file of biological underwater sounds. The John Hopkins Press, Baltimore.
- Folmer, O., Black, M., Hoeh, W., Lutz, R., Vrijenhoek, R., 1994. DNA primers for amplification of mitochondrial cytochrome *c* oxidase subunit I from diverse metazoan invertebrates. *Mol. Mar. Biol. Biotechnol.* 3, 294-299.
- Groth, J.G., Barrowclough, G.F., 1999. Basal divergences in birds and the phylogenetic utility of the nuclear RAG-1 gene. *Mol. Phylogenet. Evol.* 12, 115-123.
- Hall, B.G., 2004. Phylogenetic trees made easy: A how-to-manual, Second Edition. Sinauer Associates, Inc.
- Hall, T., 2001. BioEdit v7.0.5. North Carolina State University, Department of Microbiology.
- Helfman, G.S., Meyer, J.L., McFarland, W.M., 1982. The ontogeny of twilight migration patterns in grunts (Pisces: Haemulidae). *Anim. Behav.* 30, 317-326.
- Hillis, D.M., Moritz, C., Mable, B.K., 1996. Molecular Systematics. Sinauer Associates, Inc. Sunderland, MA. 655 pp.
- Hobson, E.S., 2006. Evolution. In: Allen, L.G., Pondella, D.J., Horn, M.H. (Eds.), *The Ecology of Marine Fishes: California and Adjacent Waters*, p. 55-80. University of California Press, Berkeley, CA.

- Hodgson, G., 1999. A global assessment of human effects on coral reefs. *Mar. Pollut. Bull.* 38, 345-355.
- Holcroft, N.I., 2004. A molecular test of alternative hypotheses of tetraodontiform (Acanthomorpha: Tetraodontiformes) sister group relationships using data from the RAG1 gene. *Mol. Phylogenet. Evol.* 32, 749-760.
- Hong, S.L., 1977. Review of Eastern Pacific *Haemulon* with notes on juvenile pigmentation. *Copeia* 3, 493-501.
- Iwatsuki, Y., Nakabo, T., 2005. Redescription of *Hapalogenys nigripinnis* (Schlegel in Temminck and Schlegel, 1843), a senior synonym of *H. nitens* Richardson, 1844, and a new species from Japan. *Copeia* 4, 854-867.
- Iwatsuki, Y., Russell, B.C., 2006. Revision of the genus *Hapalogenys* (Teleostei: Perciformes) with two new species from the Indo-West Pacific. *Mem. Mus. Vic.* 63, 29-46.
- Iwatsuki, Y., Satapoomin, U., Amaoka, K., 2000. New species: *Hapalogenys merguiensis* (Teleostei: Perciformes) from Andaman Sea. *Copeia* 1, 129-139.
- Johnson, G.D., 1980. The limits and relationships of the Lutjanidae and associated families. *Bull. Scripps. Inst. Oceanogr. Univ. Calif.* 24, 1-114.
- Källersjö, M., Albert, V.A., Farris, J.S., 1999. Homoplasy increases phylogenetic structure. *Cladistics* 15, 91-93.
- Konchina, Y.V., 1977. The systematics and distribution of the grunts family (Pomadasyidae). *J. Ichthyol.* 16, 883-900.

- Lindeman, K.C., 1986. Development of larvae of the French grunt, *Haemulon flavolineatum*, and comparative development of twelve western Atlantic species of *Haemulon* (Percoidei, Haemulidae). Bull. Mar. Sci. 39, 673-716.
- Lindeman, K.C., Toxey, C., 2003. Haemulidae – Grunts. pp. 1375-2127. In: Carpenter, K.E. (Ed.), The living marine resources of the Western Central Atlantic. Volume 3: Bony fishes part 2 (Opistognathidae to Molidae), sea turtles and marine mammals. FAO species identification guide for fishery purposes and American Society of Ichthyologists and Herpetologists Special Publication No. 5. FAO, Rome, Italy. pp. 1522-1550.
- McFarland, W.N., Brothers, E.B., Ogden, J.C., Shulman, M.J., Bermingham, E.L., Kotchian-Prentiss, N.M., 1985. Recruitment patterns in young French grunts, *Haemulon flavolineatum* (Family Haemulidae), at St Croix, Virgin Islands. Fish. Bull. US 83, 413-426.
- McFarland, W.N., Wahl, C.M., 1996. Visual constraints on migration behavior of juvenile French grunts. Environ. Biol. Fishes 46, 109-122.
- McKay, R.J., 1984. Haemulidae. In: Fischer, W., Bianchi, G. (Eds.), FAO species identification sheets for fishery purposes. Western Indian Ocean (Fishing Area 51). Vol. 2. Bony fishes (Congiopodidae to Lophotidae). FAO, Rome. pag. var.
- McKay, R.J., 2001. Haemulidae (= Pomadasysidae). Grunts (also sweetlips, rubberlips, hotlips, and velvetchins). pp. 2961-2976. In: Carpenter, K.E., Niem, V. (Eds.), FAO species identification guide for fishery purposes. The living marine resources of the Western Central Pacific. Volume 5. Bony fishes part 3 (Menidae to Pomacentridae). FAO, Rome, Italy.

- McLean, R., Herrnkind, W.F., 1971. Compact schooling during a mass movement by grunts. *Copeia*, 328-330.
- Meyer, J.L., Shultz, E.T., 1985. Migrating haemulid fishes as a source of nutrients and organic matter on coral reefs. *Limnol. Oceanogr.* 30, 146-156.
- Morinière, E.C., Pollux, B.J.A., Nagelkerken, I., Veld, G., 2003. Diet shifts of Caribbean grunts (Haemulidae) and snappers (Lutjanidae) and the relation with nursery-to-coral reef migrations. *Estuar. Coast. Shelf Sci.* 57, 1079-1089.
- Nagelkerken, I., van der Velde, G., 2004. Relative importance of interlinked mangroves and seagrass beds as feeding habitats for juvenile reef fish on a Caribbean island. *Mar. Ecol. Prog. Ser.* 274, 153-159.
- Nelson, J.S., 1994. *Fishes of the World*, 3rd Edition. John Wiley & Sons, Inc., Hoboken, New Jersey. 600 pp.
- Nelson, J.S., 2006. *Fishes of the World*, 4th Edition. John Wiley & Sons, Inc., Hoboken, New Jersey. 624 pp.
- Nylander, J.A.A., 2004. MrModeltest v2. Program distributed by the author. Evolutionary Biology Centre, Uppsala University.
- Ogden, J.C., Ehrlich, P.R., 1977. The behavior of heterotypic resting schools of juvenile grunts (Pomadasyidae). *Mar. Biol.* 42, 273-280.
- Orrell, T.M., 2000. A molecular phylogeny of the Sparidae (Perciformes: Percoidae). PhD Thesis. Faculty of the School of Marine Science, The College of William and Mary.
- Page, R.D.M., 1996. TREEVIEW: An application to display phylogenetic trees on personal computers. *CABIOS* 12, 357-358.

- Purcell, J.F.H., Cowen, R.K., Hughes, C.R., Williams, D.A., 2006. Weak genetic structure indicates strong dispersal limits: a tale of two coral reef fish. *Proc. R. Soc. Biol. Sci. Ser. B* 273, 1483-1490.
- Rasotto, M.B., Sadovy, Y., 1995. Peculiarities of the male urogenital apparatus of two grunt species (Teleostei: Haemulidae). *J. Fish Biol.* 46, 936-948.
- Richardson, J., 1844. Description of a genus of Chinese fish. *Ann. Mag. Nat. Hist. (N. S.)* 13, 461-462.
- Roux, C., 1981. Pomadasyidae. In: Fischer, W., Bianchi, G., Scott, W.B. (Eds.), *FAO species identification sheets for fishery purposes. Eastern Central Atlantic; fishing areas 34, 47 (in part). Department of Fisheries and Oceans Canada and FAO. Vol. 3. Bony fishes (Malacanthidae to Scombridae)* pag. var.
- Rüber, L., Britz, R., Kullander, S.O., Zardoya, R., 2004. Evolutionary and biogeographic patterns of the Badidae (Teleostei: Perciformes) inferred from mitochondrial and nuclear DNA sequence data. *Mol. Phylogenet. Evol.* 32, 1010-1022.
- Smith, R.F., Wiese, B.A., Wojzynski, M.K., Davison, D.B., Worley, K.C., 1996. BCM Search Launcher - An integrated interface to molecular biology data base search and analysis services available on the World Wide Web. *Genome Res.* 6, 454-62.
- Springer, M.S., DeBry, R.W., Douady, C., Amrine, H.M., Madsen, O., de Jong, W.W., Stanhope, M.J., 2001. Mitochondrial versus nuclear gene sequences in deep-level mammalian phylogeny reconstruction. *Mol. Biol. Evol.* 18, 132-143.
- Springer, V.G., Raasch, M.S., 1995. Fishes, angling, and finfish fisheries on stamps of the world. American Topical Association. Handbook 129. Tucson, Arizona. 110 pp.

- Swofford, D.L., 2002. PAUP*: Phylogenetic analysis using parsimony (*and other methods), version 4.0B10. Sinauer Associates, Sunderland, MA.
- Tamura, T., Nei, M., 1993. Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA data. *Mol. Biol. Evol.* 10, 512-526.
- Thompson, J.D., Gibson, T.J., Plewniak, F., Jeanmougin, F., Higgins, D.G., 1997. The ClustalX windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res.* 24, 4876-4882.
- Tupper, M., Juanes, F., 1999. Effects of a marine reserve on recruitment of grunts (Pisces: Haemulidae) at Barbados, West Indies. *Environ. Biol. Fishes* 55, 53-63.
- Wainwright, P.C., 1989. Functional morphology of the pharyngeal jaw apparatus in perciform fishes: An experimental analysis of the Haemulidae. *J. Morphol.* 200, 231-245.
- Williams, A., Begg, G., Garrett, R., Larson, H., Griffiths, S., 2004a. Coastal Fishes. In: National Oceans Office. Description of key species groups in the Northern Planning Area. National Oceans Office, Hobart, Australia.
- Williams, D.A., Purcell, J., Cowen, R.K., Hughes, C.R., 2004b. Microsatellite multiplexes for high-throughput genotyping of French grunts (*Haemulon flavolineatum*, Pisces: Haemulidae) and their utility in other grunt species. *Mol. Ecol. Notes* 4, 46-48.
- Williams, S.T., Jara, J., Gomez, E., Knowlton, N., 2002. The marine Indo-West Pacific break: Contrasting the resolving power of mitochondrial and nuclear genes. *Integr. Comp. Biol.*, 42, 941-952.

APPENDIX 1

RAG1 SEQUENCES OF HAEMULIDS, INCLUDING OUTGROUPS

<i>Diagramma pictum</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Plectorhinchus vittatus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Plectorhinchus chaetodonoides</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Plectorhinchus macrolepis</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Plectorhinchus schotaf</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Parapristipoma octolineatum</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Parapristipoma trilineatum</i>	GATTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Haemulopsis elongatus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Haemulopsis leuciscus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Haemulopsis axillaris</i>	ACTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulopsis nitidus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Conodon serrifer</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Plectorhinchus sordidus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Xenichthys xanti</i>	ACTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Pomadasys branickii</i>	ACTTCTCGCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Brachydeuterus auritus</i>	GCTGACCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTATAAGAAATG
<i>Anisotremus interruptus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Anisotremus virginicus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Orthopristis chalcus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Genyatremus luteus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Anisotremus pacifici</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Anisotremus dovii</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Pomadasys maculatus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Pomadasys argenteus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Pomadasys kaakan</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Pomadasys argyreus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Haemulon plumierii</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulon sciurus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulon steindachneri</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Plectorhinchus lessonii</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulon scudderii</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulon flaviguttatum</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulon aurolineatum</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Xenistius californiensis</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Inermia vittata</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAGGAATG
<i>Diplodus bermudensis</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Pagrus pagrus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Diagramma pictum</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCGCTGGATGGGCT
<i>Plectorhinchus vittatus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCGCTGGATGGGCT
<i>Plectorhinchus chaetodonoides</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCGCTGGATGGGCT
<i>Plectorhinchus macrolepis</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCGCTGGATGGGCT
<i>Plectorhinchus schotaf</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCGCTGGATGGGCT
<i>Parapristipoma octolineatum</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCGCTGGATGGGCT
<i>Parapristipoma trilineatum</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCGCTGGATGGGCT
<i>Haemulopsis elongatus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulopsis leuciscus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulopsis axillaris</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulopsis nitidus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Conodon serrifer</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Plectorhinchus sordidus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Xenichthys xanti</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Pomadasys branickii</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Brachydeuterus auritus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Anisotremus interruptus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Anisotremus virginicus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Orthopristis chalcus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Genyatremus luteus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Anisotremus pacifici</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Anisotremus dovii</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Pomadasys maculatus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Pomadasys argenteus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT

<i>Pomadasys kaakan</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Pomadasys argyreus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon plumierii</i>	TGTCGACATCTTGCAACGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon sciurus</i>	TGTCGACATCTTGCAACGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon steindachneri</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Plectorhinchus lessonii</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon scudderii</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon flaviguttatum</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon aurolineatum</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Xenistius californiensis</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Inermia vittata</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Diplodus bermudensis</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Pagrus pagrus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Diagramma pictum</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Plectorhinchus vittatus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Plectorhinchus chaetodonoides</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Plectorhinchus macrolepis</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Plectorhinchus schotaf</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Parapristipoma octolineatum</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Parapristipoma trilineatum</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Haemulopsis elongatus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Haemulopsis leuciscus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Haemulopsis axillaris</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Haemulopsis nitidus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Conodon serrifer</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Plectorhinchus sordidus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Xenichthys xanti</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Pomadasys branickii</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Brachydeuterus auritus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Anisotremus interruptus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Anisotremus virginicus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Orthopristis chalcus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Genyatremus luteus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Anisotremus pacifici</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Anisotremus dovii</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Pomadasys maculatus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Pomadasys argenteus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Pomadasys kaakan</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Pomadasys argyreus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Haemulon plumierii</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Haemulon sciurus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Haemulon steindachneri</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Plectorhinchus lessonii</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Haemulon scudderii</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Haemulon flaviguttatum</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Haemulon aurolineatum</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Xenistius californiensis</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Inermia vittata</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Diplodus bermudensis</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Pagrus pagrus</i>	TCCTCAGTGGATGACTCCCCAGCTGACACCATCACTCGGCGGTTTCGCTA
<i>Diagramma pictum</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Plectorhinchus vittatus</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Plectorhinchus chaetodonoides</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Plectorhinchus macrolepis</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Plectorhinchus schotaf</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Parapristipoma octolineatum</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Parapristipoma trilineatum</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Haemulopsis elongatus</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Haemulopsis leuciscus</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Haemulopsis axillaris</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Haemulopsis nitidus</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Conodon serrifer</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Plectorhinchus sordidus</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Xenichthys xanti</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Pomadasys branickii</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Brachydeuterus auritus</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Anisotremus interruptus</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Anisotremus virginicus</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG
<i>Orthopristis chalcus</i>	TGATGTGGCGCTGGTGTGAGCATTAAAGGATCTGGAGGAGGACATCATGG

<i>Haemulon aurolineatum</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTCGAAGCCTCGGGGT
<i>Xenistius californiensis</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTCGAGGCCTCGGGGT
<i>Inermia vittata</i>	ATACGATGAGAAGATGGTGCGCGAGGTAGAGGGGCTCGAGGCCTCGGGGT
<i>Diplodus bermudensis</i>	AGGCGATGAGAGAATGATACGTGAGATGGCGGCCCTCGTGGCGTCAGGGT
<i>Pagrus pagrus</i>	ATACGACGAGAAGATGGTGCGCGAGATGGAGGGGCTCGAGGCCTCGGGGT
<i>Diagramma pictum</i>	CCACCTACATCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCCTCCGAA
<i>Plectorhinchus vittatus</i>	CCACCTACATCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCCTCCGAA
<i>Plectorhinchus chaetodonoides</i>	CCACCTACATCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCCTCCGAA
<i>Plectorhinchus macrolepis</i>	CCACCTACATCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCCTCTGAG
<i>Plectorhinchus schotaf</i>	CCACCTACATCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCCTCCGAG
<i>Parapristipoma octolineatum</i>	CTTCCTATGTCTGCACCTGTGTGACTCCAGCCGGGCGAGAGGCCTCTCAA
<i>Parapristipoma trilineatum</i>	CCACCTACGTCTGCACCTGTGTGACTCCAGCCGGGCGAGAGGCCTCTCAA
<i>Haemulopsis elongatus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCCTCTCAA
<i>Haemulopsis leuciscus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCCTCTCAA
<i>Haemulopsis axillaris</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCCTCTCAA
<i>Haemulopsis nitidus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Conodon serrifer</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Plectorhinchus sordidus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Xenichthys xanti</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Pomadasys branickii</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Brachydeuterus auritus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Anisotremus interruptus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Anisotremus virginicus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Orthopristis chalcus</i>	CGACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Genyatremus luteus</i>	CGACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Anisotremus pacifici</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Anisotremus dovii</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Pomadasys maculatus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Pomadasys argenteus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Pomadasys kaakan</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Pomadasys argyreus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Haemulon plumierii</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Haemulon sciurus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Haemulon steindachneri</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Plectorhinchus lessonii</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Haemulon scudderii</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Haemulon flaviguttatum</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Haemulon aurolineatum</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Xenistius californiensis</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Inermia vittata</i>	CCACCTATGTCTGCACTCTGTGTGACTCCAGCCGGGCGAGAGGCATCTCAA
<i>Diplodus bermudensis</i>	CCACCTATGCTTGCATTCTATGCGACTCCAGTCGAGCGGACTCGTCTCAA
<i>Pagrus pagrus</i>	CCACCTACGTCTGCACTCTGTGCGACTCCAGTCGCGGCGAGGCCTCTCAA
<i>Diagramma pictum</i>	AACATGGTGCTCCACTCTGTGCACACGACGCCATGAAGAGAACCTGGACCC
<i>Plectorhinchus vittatus</i>	AACATGGTGCTCCACTCTGTGCACACGACGCCATGAAGAGAACCTGGACCG
<i>Plectorhinchus chaetodonoides</i>	AACATGGTGCTCCACTCTGTGCACACGACGCCATGAAGAGAACCTGGACCG
<i>Plectorhinchus macrolepis</i>	AACATGGTGCTCCACTCTGTGCACACGACGCCATGAAGAGAACCTGGACCG
<i>Plectorhinchus schotaf</i>	AACATGGTGCTCCACTCTGTGCACACGACGCCATGAAGAGAACCTGGACCG
<i>Parapristipoma octolineatum</i>	AACATGGTGCTCCACTCCGTGCACACGCTGTGCATGAAGAGAACCTGGACCG
<i>Parapristipoma trilineatum</i>	AACATGGTGCTCCACTCCGTGCACACGCTGTGCATGAAGAGAACCTGGACCG
<i>Haemulopsis elongatus</i>	AACATGGTGCTACACTCCATCAGCAGCCATGAAGAGAATCTGGACCG
<i>Haemulopsis leuciscus</i>	AACATGGTGCTACACTCCATCAGCAGCCATGAAGAGAATCTGGACCG
<i>Haemulopsis axillaris</i>	AACATGGTGCTTCACTCTGTGCACAGTAGCCACGAAGAGAACCTGGACCG
<i>Haemulopsis nitidus</i>	AACATGGTGCTTCACTCTGTGCACAGTAGCCACGAAGAGAACCTGGACCG
<i>Conodon serrifer</i>	AACATGGTGCTGCACTCAGTCACACGACGCCACGAAGAGAACCTGGACCG
<i>Plectorhinchus sordidus</i>	AACATGGTGCTGCACTCTGTGCACACGACGCCACGAAGAGAACCTGGACCG
<i>Xenichthys xanti</i>	AACATGGTGCTGCACTCTGTGCACACGACGCCACGAAGAGAACCTGGACCG
<i>Pomadasys branickii</i>	AACATGGTGCTGCACTCTGTGCACACGACGCCACGAAGAGAACCTGGACCG
<i>Brachydeuterus auritus</i>	AACATGGTGCTGCACTCTGTGCACACGACGCCACGAAGAGAACCTGGACCG
<i>Anisotremus interruptus</i>	AACATGGTGCTGCACTCTGTGCACACGACGCCACGAAGAGAACCTGGACCG
<i>Anisotremus virginicus</i>	AACATGGTGCTGCACTCTGTGCACACGACGCCACGAAGAGAACCTGGACCG
<i>Orthopristis chalcus</i>	AACATGGTGCTGCACTCTGTGCACACGTAACACGAGGAGAACCTGGACCG
<i>Genyatremus luteus</i>	AACATGGTGCTCCACTCTGTGCACACGACGCCACGAAGAGAACCTGGACCG
<i>Anisotremus pacifici</i>	AACATGGTGCTGCACTCTGTGCACACGACGCCACGAAGAGAACCTGGACCG
<i>Anisotremus dovii</i>	AACATGGTGCTGCACTCTGTGCACACGACGCCACGAAGAGAACCTGGACCG
<i>Pomadasys maculatus</i>	AACATGGTGCTGCACTCCATCTCAGCAGCCACGAAGAGAACCTGGACCG
<i>Pomadasys argenteus</i>	AACATGGTGCTGCACTCCATCAGCAGCCACGAAGAGAACCTGGACCG
<i>Pomadasys kaakan</i>	AACATGGTGCTGCACTCCATCAGCAGCCACGAAGAGAACCTGGACCG
<i>Pomadasys argyreus</i>	AACATGGTGCTGCACTCTATCAGCAGCCACGAAGAGAACCTGGACCG
<i>Haemulon plumierii</i>	AACATGGTGCTGCACTCCATCAGCAGCCACCATGAAGAGAACCTGGACCG

<i>Pomadasyys maculatus</i>	GAGACAGAGTCAAAGGAGTCTCTGCCAAGCCCTTCTTGGAGACCCATCCC
<i>Pomadasyys argenteus</i>	GAGACAGAGTCAAAGGAGTCTCTGCCAAGCCCTTTCATGGAGACCCATCCC
<i>Pomadasyys kaakan</i>	GAGACAGAGTCAAAGGAGTCTCTGCCAAGCCCTTTCATGGAGACCCATCCC
<i>Pomadasyys argyreus</i>	GAGACAGAGTCAAAGGAGTCTCTGCCAAGCCCTTTCATGGAGACCCATCCC
<i>Haemulon plumierii</i>	GAGACAGAGTCAAAGGAGTTTCTGCCAAGCCCTTTCATGGAGACTCATCCC
<i>Haemulon sciurus</i>	GAGACAGAGTCAAAGGAGTTTCTGCCAAGCCCTTTCATGGAGACTCATCCC
<i>Haemulon steindachneri</i>	GAGACAGAGTCAAAGGAGTTTCTGCCAAGCCCTTTCATGGAGACTCATCCC
<i>Plectorhinchus chaetodonoides</i>	GAGACAGAGTCAAAGGAGTTTCTGCCAAGCCCTTTCATGGAGACTCATCCC
<i>Haemulon scudderii</i>	GAGACAGAGTCAAAGGAGTCTCTGCCAAGCCCTTTCATGGAGACCCATCCC
<i>Haemulon flaviguttatum</i>	GAGACAGAGTCAAAGGAGTTTCTGCCAAGCCCTTTCATGGAGACTCATCCC
<i>Haemulon aurolineatum</i>	GAGACAGAGTCAAAGGAGTCTCTGCCAAGCCCTTTCATGGAGACCCATCCC
<i>Xenistius californiensis</i>	GAGACAGAGTCAAAGGAGTCTCTGCCAAGCCCTTTCATGGAGACCCATCCC
<i>Inermia vittata</i>	GGGACAGAGTCAAAGGGATTCTCTGCCAAGCCCTTTCATGGAGACCCATCCC
<i>Diplodus bermudensis</i>	GAGACAGAGTCAAAGGAGTGTGGAACAAGCCCTTTCATGGAGACCCATCAC
<i>Pagrus pagrus</i>	GAGACAGAGTCAAAGGGGTCTCGGCCAAGCCCTTTCATCGAGACCCATCCC
<i>Diagramma pictum</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Plectorhinchus vittatus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Plectorhinchus chaetodonoides</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Plectorhinchus macrolepis</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Plectorhinchus schotaf</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Parapristipoma octolineatum</i>	ACACTGGATGCCTTGCACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Parapristipoma trilineatum</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Haemulopsis elongatus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Haemulopsis leuciscus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Haemulopsis axillaris</i>	ACGCTGGATGCATTACACTGTGACATCGGCAATGCCACTGAGTTCTATAA
<i>Haemulopsis nitidus</i>	TCGCTGGATGCATTACACTGTGACATCGGCAATGCCACTGAGTTCTATAA
<i>Conodon serrifer</i>	ACGCTGGATGCATTACACTGTGACATTGGCAATGCCACCGAGTTCTACAA
<i>Plectorhinchus sordidus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Xenichthys xanti</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Pomadasyys branickii</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Brachydeuterus auritus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Anisotremus interruptus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Anisotremus virginicus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Orthopristis chalcus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Genyatremus luteus</i>	ACGCTAGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Anisotremus pacifici</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCTACTGAGTTCTACAA
<i>Anisotremus dovii</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Pomadasyys maculatus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Pomadasyys argenteus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Pomadasyys kaakan</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Pomadasyys argyreus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Haemulon plumierii</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Haemulon sciurus</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Haemulon steindachneri</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Plectorhinchus lessonii</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Haemulon scudderii</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Haemulon flaviguttatum</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Haemulon aurolineatum</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Xenistius californiensis</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Inermia vittata</i>	ACGCTGGATGCATTACACTGTGACATAGGCAATGCCACTGAGTTCTACAA
<i>Diplodus bermudensis</i>	ACGCTGGACGCGCTGCATTGTGACATCGGCAACGCCACCGAGTTCTACAA
<i>Pagrus pagrus</i>	ACGCTGGACGCGCTGCATTGTGACATCGGCAACGCCACCGAGTTCTACAA
<i>Diagramma pictum</i>	AATCTTCCAGGATGAGATCGGGGAGGTGCACCAAAGAGTCAACCTTAGCC
<i>Plectorhinchus vittatus</i>	AATCTTCCAGGATGAGATCGGGGAGGTGCACCAAAGAGTCAACCTTAGCC
<i>Plectorhinchus chaetodonoides</i>	AATCTTCCAGGATGAGATCGGGGAGGTGCACCAAAGAGTCAACCTTAGCC
<i>Plectorhinchus macrolepis</i>	AATCTTCCAGGATGAGATTGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Plectorhinchus schotaf</i>	AATCTTCCAGGATGAGATTGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Parapristipoma octolineatum</i>	AATTTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTCAACCTTAGCC
<i>Parapristipoma trilineatum</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTCAACCTTAGCC
<i>Haemulopsis elongatus</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Haemulopsis leuciscus</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Haemulopsis axillaris</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Haemulopsis nitidus</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Conodon serrifer</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Plectorhinchus sordidus</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Xenichthys xanti</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Pomadasyys branickii</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Brachydeuterus auritus</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTGAACCTTAGCC
<i>Anisotremus interruptus</i>	AATCTTCCAGGATGAGATCGGGGAGGTGTACCAAAGAGTGAACCTTAGCC

<i>Anisotremus virginicus</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAGAACCCAAACCCGAGCC
<i>Orthopristis chalcus</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Genyatremus luteus</i>	AATTTTCCAGGATGAGATCGGGGAATTGTACCAAAACCCAAACCCGAGCC
<i>Anisotremus pacifici</i>	AATTTTCCAGGATGAGATCGGGGAATTGTACCAAAACCCAAACCCGAGCC
<i>Anisotremus dovii</i>	AATTTTCCAGGATGAGATCGGGGAATTGTACCAAAACCCAAACCCGAGCC
<i>Pomadasys maculatus</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGTC
<i>Pomadasys argenteus</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Pomadasys kaakan</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Pomadasys argyreus</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Haemulon plumierii</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Haemulon sciurus</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Haemulon steindachneri</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Plectorhinchus lessonii</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Haemulon scudderii</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Haemulon flaviguttatum</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Haemulon aurolineatum</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Xenistius californiensis</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Inermia vittata</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Diplodus bermudensis</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Pagrus pagrus</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAACCCAAACCCGAGCC
<i>Diagramma pictum</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Plectorhinchus vittatus</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Plectorhinchus chaetodonoides</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Plectorhinchus macrolepis</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Plectorhinchus schotaf</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Parapristipoma octolineatum</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Parapristipoma trilineatum</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulopsis elongatus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulopsis leuciscus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulopsis axillaris</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulopsis nitidus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Conodon serrifer</i>	GTGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Plectorhinchus sordidus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Xenichthys xanti</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pomadasys branickii</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Brachydeuterus auritus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Anisotremus interruptus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Anisotremus virginicus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Orthopristis chalcus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Genyatremus luteus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Anisotremus pacifici</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Anisotremus dovii</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pomadasys maculatus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pomadasys argenteus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pomadasys kaakan</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pomadasys argyreus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon plumierii</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon sciurus</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon steindachneri</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Plectorhinchus lessonii</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon scudderii</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon flaviguttatum</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon aurolineatum</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Xenistius californiensis</i>	GAGAGCAACGACGCGAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Inermia vittata</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Diplodus bermudensis</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pagrus pagrus</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Diagramma pictum</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Plectorhinchus vittatus</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Plectorhinchus chaetodonoides</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Plectorhinchus macrolepis</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Plectorhinchus schotaf</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Parapristipoma octolineatum</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Parapristipoma trilineatum</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Haemulopsis elongatus</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Haemulopsis leuciscus</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Haemulopsis axillaris</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Haemulopsis nitidus</i>	AAGATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG
<i>Conodon serrifer</i>	AACATGAAGCTAAAGCCAGTAATGAGGATGAATGGGAACCTATGCCCGCCG

Plectorhinchus chaetodonoides GTGCCGCTACAGCTTTAACTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Plectorhinchus macrolepis GTGCCGCTACAGCTTTAACTCCCAGCGCTTTGCTGACCTCCTCTCCTCTA
Plectorhinchus schotaf GTGCCGCTACAGCTTTAACTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Parapristipoma octolineatum GTGCCGCTACAGCTTTAACTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Parapristipoma trilineatum GTGCCGCTACAGCTTTAACTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Haemulopsis elongatus GTGCCGCTACAGCTTTAACTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Haemulopsis leuciscus GTGCCGCTACAGCTTTAACTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Haemulopsis axillaris GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Haemulopsis nitidus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Conodon serrifer GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Plectorhinchus sordidus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Xenichthys xanti GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Pomadasys branickii GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Brachydeuterus auritus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Anisotremus interruptus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Anisotremus virginicus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Orthopristis chalcus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Genyatremus luteus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Anisotremus pacifici GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Anisotremus dovii GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Pomadasys maculatus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Pomadasys argenteus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Pomadasys kaakan GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Pomadasys argyreus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Haemulon plumierii GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Haemulon sciurus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Haemulon steindachneri GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Plectorhinchus lessonii GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Haemulon scudderii GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Haemulon flaviguttatum GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Haemulon aurolineatum GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Xenistius californiensis GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Inermia vittata GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Diplodus bermudensis GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA
Pagrus pagrus GTGCCGCTACAGCTTTAATTTCCCAGCGCTTTGCCGACCTCCTCTCCTCTA

Diagramma pictum CCTTCAAAGTATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
Plectorhinchus vittatus CCTTCAAATATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
Plectorhinchus chaetodonoides CCTTCAAATATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
Plectorhinchus macrolepis CCTTCAAATATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
Plectorhinchus schotaf CCTTCAAATATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
Parapristipoma octolineatum CCTTCAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Parapristipoma trilineatum CCTTCAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Haemulopsis elongatus CCTTCAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Haemulopsis leuciscus CCTTCAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Haemulopsis axillaris CATTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Haemulopsis nitidus CATTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Conodon serrifer CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Plectorhinchus sordidus CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTATCTACACAAGACC
Xenichthys xanti CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTATCTGCACAAGACC
Pomadasys branickii CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Brachydeuterus auritus CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Anisotremus interruptus CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Anisotremus virginicus CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Orthopristis chalcus CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Genyatremus luteus CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Anisotremus pacifici CCTTTAAATATAGGTACAACGGGAAGATAAACCAATTACCTGCACAAGACC
Anisotremus dovii CCTTTAAATATAGGTACAATGGGAAGATGACCAATTACCTGCACAAGACC
Pomadasys maculatus CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAAACC
Pomadasys argenteus CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Pomadasys kaakan CCTTCAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Pomadasys argyreus CCTTTAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC
Haemulon plumierii CCTTTAAATATAGGTACAATGGGAAGATAAACCAATTACCTGCACAAGACC
Haemulon sciurus CCTTTAAATATAGGTACAATGGGAAGATAAACCAATTACCTGCACAAGACC
Haemulon steindachneri CCTTTAAATATAGGTACAATGGGAAGATAAACCAATTACCTGCACAAGACC
Plectorhinchus lessonii CCTTTAAATATAGGTACAATGGGAAGATAAACCAATTACCTGCACAAGACC
Haemulon scudderii CCTTTAAATATAGGTACAATGGGAAGATAAACCAATTACCTGCACAAGACC
Haemulon flaviguttatum CCTTTAAATATAGGTACAATGGGAAGATAAACCAATTACCTGCACAAGACC
Haemulon aurolineatum CCTTTAAATATAGGTACAATGGGAAGATAAACCAATTACCTGCACAAGACC
Xenistius californiensis CCTTTAAATATAGGTACAATGGGAAGATAAACCAATTACCTGCACAAGACC
Inermia vittata CCTTCAAATATAGGTACAATGGAAAGATAAACCAATTACCTGCACAAGACC

<i>Diplodus bermudensis</i>	CCTTCAAATATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
<i>Pagrus pagrus</i>	CCTTCAAGTACAGGTACAACGGAAAGATAACCAACTACCTGCGCAAAACG
<i>Diagramma pictum</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Plectorhinchus vittatus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Plectorhinchus chaetodonoides</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Plectorhinchus macrolepis</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Plectorhinchus schotaf</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Parapristipoma octolineatum</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Parapristipoma trilineatum</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulopsis elongatus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulopsis leuciscus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulopsis axillaris</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulopsis nitidus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Conodon serrifer</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Plectorhinchus sordidus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Xenichthys xanti</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pomadasys branickii</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Brachydeuterus auritus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Anisotremus interruptus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Anisotremus virginicus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Orthopristis chalcus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Genyatremus luteus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Anisotremus pacifici</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Anisotremus dovii</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pomadasys maculatus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pomadasys argenteus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pomadasys kaakan</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pomadasys argyreus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon plumierii</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon sciurus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon steindachneri</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Plectorhinchus lessonii</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon scudderii</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon flaviguttatum</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon aurolineatum</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Xenistius californiensis</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Inermia vittata</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Diplodus bermudensis</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pagrus pagrus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Diagramma pictum</i>	GGCCAGTGAGGGGAACGAGTCGGCAAAACAACTGTT
<i>Plectorhinchus vittatus</i>	GGCCAGTGAGGGGAATGAGTCGGCAAAACAACTGTT
<i>Plectorhinchus chaetodonoides</i>	GGCCAGTGAGGGGAACGAGTCGGCAAAACAACTGTT
<i>Plectorhinchus macrolepis</i>	GGCCAGTGAGGGGAACGAGTCGGCAAAACAACTGTT
<i>Plectorhinchus schotaf</i>	GGCCAGTGAGGGGAACGAGTCGGCAAAACAACTGTT
<i>Parapristipoma octolineatum</i>	GGCCAGTGAGGGGAACGAGTCGGCAAAACAACTGTT
<i>Parapristipoma trilineatum</i>	GGCCAGTGAGGGGAACGAGTCGGCAAAACAACTGTT
<i>Haemulopsis elongatus</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Haemulopsis leuciscus</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Haemulopsis axillaris</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Haemulopsis nitidus</i>	GACCATTGATGGGAAGGAATCAGCAAAACGAAGTATT
<i>Conodon serrifer</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Plectorhinchus sordidus</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Xenichthys xanti</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Pomadasys branickii</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Brachydeuterus auritus</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Anisotremus interruptus</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Anisotremus virginicus</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Orthopristis chalcus</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Genyatremus luteus</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Anisotremus pacifici</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Anisotremus dovii</i>	GGCCATTGATGGGCAAGGATCAGCAAAACGAAGTATT
<i>Pomadasys maculatus</i>	GGCCATTGATGGGCAAGGATCAGCAAAACGAAGTATT
<i>Pomadasys argenteus</i>	GTCCATTGATGGGCAAGGATCAGCAAAACGAAGTATT
<i>Pomadasys kaakan</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Pomadasys argyreus</i>	GGCCATTGATGGGCAAGGATCAGCAAAACGAAGTATT
<i>Haemulon plumierii</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Haemulon sciurus</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Haemulon steindachneri</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACAACTGTT
<i>Plectorhinchus lessonii</i>	GGCCAGTGAGGGGAACGAGTCAGCAAAACGAAGTATT

<i>Haemulon scudderii</i>	GGCCAGCGAGGGGAACGAGTCAGCAAACAAACTGTT
<i>Haemulon flaviguttatum</i>	GGCCAGCGAGGGGAACGAGTCAGCAAACAAACTGTT
<i>Haemulon aurolineatum</i>	GGCCAGTGAGGGGAACGAGTCAGCAAACAAACTATT
<i>Xenistius californiensis</i>	GGCCAGCGAGGGGAACGAGTCAGCAAACAAACTGTT
<i>Inermia vittata</i>	GGCCAGCGAGGGGAACGAGTCGGCAAACAAGCTCTT
<i>Diplodus bermudensis</i>	GGCCAGCGAGGGGAACGAGTCGGCAAACAAGCTGTT
<i>Pagrus pagrus</i>	GGCCAGCGAGGGGAATGAGTCGCCGAACAAACTATT

APPENDIX 2

COI SEQUENCES OF HAEMULIDS, INCLUDING OUTGROUPS

<i>Plectorhinchus lessonii</i>	TATCCGAGCAGAAATTAAGCCAACCCGGCGCTCTCCTGGGAGACGACCAGA
<i>Plectorhinchus vittatus</i>	TATCCGAGCAGAAATTAAGCCAACCCGGCGCTCTCCTGGGAGACGACCAGA
<i>Diagramma pictum</i>	CATCCGGGCAGAGTTAAGCCAACCCGGCGCGCTCCTAGGAGACGACCAGA
<i>Plectorhinchus chaetodonoides</i>	CATCCGGGCAGAAATTAAGCCAACCCGGCGCTCTCCTAGGAGACGACCAGA
<i>Anisotremus dovii</i>	CATCCGAGCAGAGCTCAGCCAACCCAGGCGCCCTCCTTGGGGATGACCAAA
<i>Pomadasys argyreus</i>	TATCCGAGCAGAACTTAGCCAGCCAGGCGCCCTCCTGGGAGATGACCAGA
<i>Brachydeuterus auritus</i>	TATCCGAGCAGAACTCAGCCAACCCAGGCGCCCTCCTGGGGATGACCAAA
<i>Xenichthys xanti</i>	TATCCGAGCAGAGCTCAGCCAACCCAGGCGCTCTCCTCGGAGACGACCAGA
<i>Pomadasys maculatus</i>	CATCCGAGCAGAACTCAGCCAACCCGGGTGCACCTCCTCGGGAGCAGACCAGA
<i>Anisotremus interruptus</i>	CATCCGAGCAGAACTCAGCCAACCCGGCGCCCTCCTCGGAGACGACCAGA
<i>Anisotremus virginicus</i>	TATTCGAGCAGAGCTCAGTCAGCCGGGCGCCCTCCTCGGAGACGACCAGA
<i>Pomadasys branickii</i>	CATCCGGGCAGAGCTTAGCCAACCCGGGCGCTCTCCTAGGGGACGACCAGA
<i>Haemulopsis elongatus</i>	CATCCGAGCGGAACTCAGCCAACCCGGGCGCTCTCCTCGGGGACGACCAGA
<i>Haemulopsis leuciscus</i>	CATCCGAGCGGAACTCAGCCAACCCGGGCGCTCTCCTCGGGGACGACCAGA
<i>Haemulon flaviguttatum</i>	TATCCGGGCAGAACTAAGCCAACCCAGGCGCACTCCTTGGGGACGACCAGA
<i>Haemulon steindachneri</i>	TATCCGGGCAGAACTAAGCCAACCCGGGTGCACCTCCTCGGGGACGATCAAA
<i>Haemulon aurolineatum</i>	TATCCGGGCAGAACTGAGCCAACCCAGGCGCACTCCTGGGAGACGACCAGA
<i>Haemulon plumierii</i>	TATCCGGGCAGAACTAAGCCAAGCCAGGCGCACTCCTGGGGACGACCAGA
<i>Haemulon sciurus</i>	CATCCGGGCAGAACTGAGCCAACCCAGGCGCACTCCTGGGAGACGACCAGA
<i>Haemulon scudderii</i>	AATCCGAGCAGAACTAAGCCAACCCAGGCGCACTCCTAGGGGACGACCAGA
<i>Xenistius californiensis</i>	CATCCGGGCAGAACTTAGCCAACCCAGGCGCTCTCCTTGGAGACGACCAGA
<i>Haemulopsis axillaris</i>	TATCCGAGCAGAACTTAGCCAGCCGGGCGCCCTCCTCGGGGACGATCAGA
<i>Haemulopsis nitidus</i>	TATCCGAGCAGAACTTAGCCAGCCGGGCGCCCTCCTCGGGGACGATCAGA
<i>Anisotremus pacifici</i>	GATCCGAGCTGAACTCAGTCAACCCGGGCGCTCTCCTTGGGGACGATCAGA
<i>Orthopristis chalceus</i>	AATTCGAGCAGAACTCAGCCAGCCTGGAGCTCTCCTCGGAGACGACCAGA
<i>Pomadasys kaakan</i>	TATCCGAGCCGAACTCAGCCAACCCGGGCGCTCTCCTCGGGGACGACCAGA
<i>Diplodus bermudensis</i>	CATTTCGAGCCGAACTAAGCCAGCCTGGGCGCCCTCCTTGGCGACGACCAGA
<i>Pagrus pagrus</i>	TATTCGAGCTGAGCTTAGCCAGCCGGGCGCTCTCCTAGGCGACGACCAGA
<i>Plectorhinchus macrolepis</i>	TATCCGGGCTGAACTAAGCCAACCTGGGCGCTCTTTAGGGGACGACCAGA
<i>Plectorhinchus lessonii</i>	TTTATAATGTTATTGTTACGGCACATGCATTTCGTAATAATCTTTTATG
<i>Plectorhinchus vittatus</i>	TTTATAATGTTATTGTTACGGCACATGCATTTCGTAATAATCTTTTATG
<i>Diagramma pictum</i>	TTTACAATGTTATCGTTACGGCGCATGCGTTTCGTAATAATCTTCTTTATA
<i>Plectorhinchus chaetodonoides</i>	TTTACAATGTTATTGTTACGGCGCACGCGTTTCGTAATAATCTTCTTTATG
<i>Anisotremus dovii</i>	TTTACAATGTCATCGTTACCGCCACGCATTTCGTAATAATTTTCTTTATA
<i>Pomadasys argyreus</i>	TTTATAATGTAATCGTCACTGCCATGCTTTCGTAATAATTTTCTTTATA
<i>Brachydeuterus auritus</i>	TCTATAACGTAATCGTTACTGCACATGCATTTCGTAATAATTTTCTTTATG
<i>Xenichthys xanti</i>	TTTACAACGTAATCGTTACTGCACATGCATTTCGTAATAATCTTCTTTATA
<i>Pomadasys maculatus</i>	TTTATAACGTAATCGTTACTGCACATGCCTTCGTAATAATTTTCTTTATA
<i>Anisotremus interruptus</i>	TTTATAATGTAATGTTACTGCACACGCGTTTCGTAATAATTTTCTTTATA
<i>Anisotremus virginicus</i>	TTTATAATGTAATGTTACTGCACATGCGTTTCGTAATAATTTTCTTTATA
<i>Pomadasys branickii</i>	TTTATAATGTTATTGTTACTGCACATGCGTTTCGTAATAATTTTCTTTATA
<i>Haemulopsis elongatus</i>	TTTACAATGTTATTGTTACCGCCACGCGTTTCGTGATAATTTTCTTTATA
<i>Haemulopsis leuciscus</i>	TTTACAATGTTATTGTTACCGCCACGCGTTTCGTGATAATTTTCTTTATA
<i>Haemulon flaviguttatum</i>	TTTATAACGTAATGTTACTGCGCATGCGTTTCGTAATAATTTTCTTTATA
<i>Haemulon steindachneri</i>	TCTATAACGTAATGTTACTGCGCATGCGTTTCGTGATAATTTTCTTTATA
<i>Haemulon aurolineatum</i>	TCTATAACGTAATGTTACTGCGCATGCGTTTCGTGATAATTTTCTTTATA
<i>Haemulon plumierii</i>	TCTATAACGTAATGTTACTGCGCATGCGTTTCGTAATAATTTTCTTTATA
<i>Haemulon sciurus</i>	TCTATAACGTAATGTTACTGCGCATGCGTTTCGTAATAATTTTCTTTATA
<i>Haemulon scudderii</i>	TTTATAACGTAATGTTACTGCGCATGCGTTTCGTAATAATTTTCTTTATA
<i>Xenistius californiensis</i>	TCTATAATGTCATTGTTACTGCGCATGCGTTTCGTAATAATTTTCTTTATA
<i>Haemulopsis axillaris</i>	TTTACAATGTAATCGTAACCGCACACGCATTTCGTAATAATCTTCTTTATA
<i>Haemulopsis nitidus</i>	TTTACAATGTAATCGTAACCGCACACGCATTTCGTAATAATCTTCTTTATA
<i>Anisotremus pacifici</i>	TTTACAATGTGATCGTTACCGCCACGCATTTCGTAATAATCTTCTTTATA
<i>Orthopristis chalceus</i>	TTTACAATGTGATCGTTACTGCACATGCATTTCGTAATAATCTTCTTTATA
<i>Pomadasys kaakan</i>	TTTATAATGTAATCGTCACTGCACACGCCCTTTCGTAATAATTTTCTTTATA
<i>Diplodus bermudensis</i>	TTTATAATGTAATGTTTACAGCACATGCGTTTCGTAATAATTTTCTTTATA
<i>Pagrus pagrus</i>	TTTATAATGTTATTGTTACAGCACACGCGTTTCGTAATAATTTTCTTTATA
<i>Plectorhinchus macrolepis</i>	TCTACAATGTAATCGTTACAGCCACGCTTTCGTAATAATTTTCTTTATA
<i>Plectorhinchus lessonii</i>	GTTATACCTATCCTAATTTGGAGGATTTCGGAACCTGACTGGTCCCATTAAAT
<i>Plectorhinchus vittatus</i>	GTTATACCTATCCTAATTTGGAGGATTTCGGAACCTGACTGGTCCCATTAAAT

<i>Haemulon aurolineatum</i>	TTCGACCCGCGCGGAGGAGGTGACCCCATTCCTTTA
<i>Haemulon plumierii</i>	TTTGACCCGCGCGGAGGGGGTGACCCATTCCTTTA
<i>Haemulon sciurus</i>	TTTGACCCGCGCGGAGGGGGCGACCCATTCCTTTA
<i>Haemulon scudderii</i>	TTCGACCCGCGCGGAGGAGGTGATCCCATTCCTTTA
<i>Xenistius californiensis</i>	TTCGACCCGCGCGGAGGAGGTGACCCCATTCCTCTA
<i>Haemulopsis axillaris</i>	TTTGACCCAGCAGGAGGGGGTGACCCCAATTCCTGTA
<i>Haemulopsis nitidus</i>	TTTGACCCAGCAGGAGGGGGTGACCCGATTCCTGTA
<i>Anisotremus pacifici</i>	TTTGATCCCGCTGGAGGAGGAGACCCATTCCTATA
<i>Orthopristis chalceus</i>	TTTGACCCGCTGGAGGAGGTGACCCCATTCCTTTA
<i>Pomadasy kaakan</i>	TTCGACCCGCTGGAGGGGGTGACCCCAATCCTGTA
<i>Diplodus bermudensis</i>	TTCGACCCTGCAGGGGGAGGAGACCCGATTCCTTTA
<i>Pagrus pagrus</i>	TTCGACCCGCGAGGAGGAGGGGACCCCAATTCCTCTA
<i>Plectorhinchus macrolepis</i>	TTTGACCCTGCAGGAGGAGGGGACCCCAATTCCTGTA

APPENDIX 3

RAG1-COI CONCATENATED SEQUENCES OF HAEMULIDS, INCLUDING OUTGROUPS

<i>Diagramma pictum</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Plectorhinchus vittatus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Plectorhinchus chaetodonoides</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Plectorhinchus macrolepis</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Haemulopsis elongatus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Haemulopsis leuciscus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Anisotremus interruptus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Anisotremus virginicus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Pomadasys branickii</i>	ACTTCTCGCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Anisotremus dovii</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulopsis axillaris</i>	ACTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulopsis nitidus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Orthopristis chalcus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Anisotremus pacifici</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Xenichthys xanti</i>	ACTTCCCCCGGCTTTCGCGGGTTGAATGGCCGCAACTCTCAGGAAGG
<i>Brachydeuterus auritus</i>	GCTGACCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTATAAGAAATG
<i>Pomadasys maculatus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAATGGCAGCCAGCTCTCATGAATG
<i>Pomadasys argyreus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Pomadasys kaakan</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCATGAATG
<i>Haemulon plumierii</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulon sciurus</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulon steindachneri</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulon flaviguttatum</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulon scudderii</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Haemulon aurolineatum</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCCCTCACNAATG
<i>Xenistius californiensis</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Plectorhinchus lessonii</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCACGAATG
<i>Diplodus bermudensis</i>	GCTTCTCCCTGGCTTTCACAAATTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Pagrus pagrus</i>	GCTTCAACCCTGGCTTTCACCGGTTTGAGTGGCAGCCAGCTCTCAAGAATG
<i>Diagramma pictum</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGACTCGCTGGATGGGCT
<i>Plectorhinchus vittatus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCGCTGGATGGGCT
<i>Plectorhinchus chaetodonoides</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCGCTGGATGGGCT
<i>Plectorhinchus macrolepis</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCGCTGGATGGGCT
<i>Haemulopsis elongatus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulopsis leuciscus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Anisotremus interruptus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Anisotremus virginicus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Pomadasys branickii</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Anisotremus dovii</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulopsis axillaris</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulopsis nitidus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Orthopristis chalcus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Anisotremus pacifici</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Xenichthys xanti</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Brachydeuterus auritus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Pomadasys maculatus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Pomadasys argyreus</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Pomadasys kaakan</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon plumierii</i>	TGTCGACATCTTGCAACGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon sciurus</i>	TGTCGACATCTTGCAACGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon steindachneri</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon flaviguttatum</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon scudderii</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Haemulon aurolineatum</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Xenistius californiensis</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Plectorhinchus lessonii</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Diplodus bermudensis</i>	TGTCGACATCTTGCAATGTTGGCATTATTAATGGGCTCTCTGGATGGGCT
<i>Pagrus pagrus</i>	TGTCGACATCTTGCAACGTTGGCATTATTAATGGGCTCTCTGGATGGGCT

<i>Haemulopsis nitidus</i>	AGGGACTGAAAGAGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Orthopristis chalceus</i>	AGGGACTGAAAAAGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Anisotremus pacifici</i>	AGGGACTGAGAGAGAGTGGGCTGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Xenichthys xanti</i>	AGGGACTGAGAGAGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Brachydeuterus auritus</i>	AGGGACTGAGAGAGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Pomadasys maculatus</i>	AGGGACTGAGAGAGAGGGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Pomadasys argyreus</i>	AGGGACTGAGAGAGAGGGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Pomadasys kaakan</i>	AGGGACTGAGAGAGAGGGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Haemulon plumierii</i>	AGGGGCTGAGAGGGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Haemulon sciurus</i>	AGGGGCTGAGAGGGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Haemulon steindachneri</i>	AGGGGCTGAGAGGGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Haemulon flaviguttatum</i>	AGGGGCTGAGAGGGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Haemulon scudderii</i>	AGGGGCTGAGAGGGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Haemulon aurolineatum</i>	AGGGGCTGCGAGGGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Xenistius californiensis</i>	AGGGGCTGAGAGGGAGTGGGATGGAAGACAGTGCTTGTACCTCAGGCTTC
<i>Plectorhinchus lessonii</i>	ACGGGCTGAGAGAGAATGGGATGGAAGACAGCGCTTGCACCTCAGGCTTC
<i>Diplodus bermudensis</i>	ACGGGCTGAGAGAGAATGGGATGGAAGACAGCGCTTGCACCTCAGGCTTC
<i>Pagrus pagrus</i>	ACGGGCTGAGAGAGAATGGGATGGAAGACAGCGCTTGCACCTCAGGCTTC
<i>Diagramma pictum</i>	AGTGTCTATGATCAAGGAATCCTGTGATGGCATGGGTGATGTCAGCGAGAA
<i>Plectorhinchus vittatus</i>	AGTGTCTATGATCAAGGAATCCTGTGATGGCATGGGTGATGTCAGCGAGAA
<i>Plectorhinchus chaetodonoides</i>	AGTGTCTATGATCAAGGAATCCTGTGATGGCATGGGTGATGTCAGCGAGAA
<i>Plectorhinchus macrolepis</i>	AGTGTCTATGATCAAGGAATCGTGTGATGGCATGGGTGATGTCAGCGAGAA
<i>Haemulopsis elongatus</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Haemulopsis leuciscus</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Anisotremus interruptus</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Anisotremus virginicus</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Pomadasys branickii</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Anisotremus dovii</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGTGATGTCAGCGAGAA
<i>Haemulopsis axillaris</i>	AGTGTCTATGATCAAGGAATCGTGTGATGGCATGGGTGATGTCAGTGAGAA
<i>Haemulopsis nitidus</i>	AGTGTCTATGATCAAGGAATCGTGTGATGGCATGGGTGATGTCAGTGAGAA
<i>Orthopristis chalceus</i>	AGTGTCTATGATAAAGGAATCTTGGCATGGCATGGGTGACGTGACCCAGAA
<i>Anisotremus pacifici</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Xenichthys xanti</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Brachydeuterus auritus</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Pomadasys maculatus</i>	AGTGTCTATGATCAAGGAGTCTTGTGACGGCATGGGTGATGTCAGCGAGAA
<i>Pomadasys argyreus</i>	AGTGTCTATGATCAAGGAATCTTGTGACGGCATGGGCGATGTCAGCGAGAA
<i>Pomadasys kaakan</i>	AATGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Haemulon plumierii</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGTGATGTCAGCGAGAA
<i>Haemulon sciurus</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Haemulon steindachneri</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Haemulon flaviguttatum</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGTGATGTCAGCGAGAA
<i>Haemulon scudderii</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Haemulon aurolineatum</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Xenistius californiensis</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Plectorhinchus lessonii</i>	AGTGTCTATGATCAAGGAATCTTGTGATGGCATGGGCGATGTCAGCGAGAA
<i>Diplodus bermudensis</i>	AGAGTCTATGATCAAGGAATCCTGTGACGGCATGGGCGATGTCAGCGAGAA
<i>Pagrus pagrus</i>	AGAGTCTATGATCAAGGAGTCTTGGCAGGCATGGGCGATGTCAGCGAGAA
<i>Diagramma pictum</i>	GCATGGCGGAGGACCAGCTGTTCCCTGAGAAGGCTGTGCGTTTCTCTTTCA
<i>Plectorhinchus vittatus</i>	GCATGGCGGAGGACCAGCTGTTCCCTGAGAAGGCTGTGCGTTTCTCTTTCA
<i>Plectorhinchus chaetodonoides</i>	GCATGGCGGAGGACCAGCTGTTCCCTGAGAAGGCTGTGCGTTTCTCTTTCA
<i>Plectorhinchus macrolepis</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTGCGTTTCTCTTTCA
<i>Haemulopsis elongatus</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Haemulopsis leuciscus</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Anisotremus interruptus</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Anisotremus virginicus</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Pomadasys branickii</i>	GCACGGTGGAGGACCAGTGTTCCTGATAAGGCTGTACGTTTCTCTTTCA
<i>Anisotremus dovii</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Haemulopsis axillaris</i>	GCATGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Haemulopsis nitidus</i>	GCATGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Orthopristis chalceus</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Anisotremus pacifici</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Xenichthys xanti</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Brachydeuterus auritus</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Pomadasys maculatus</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Pomadasys argyreus</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Pomadasys kaakan</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Haemulon plumierii</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Haemulon sciurus</i>	GCACGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA
<i>Haemulon steindachneri</i>	GCATGGTGGAGGACCAGTGTTCCTGAGAAGGCTGTACGTTTCTCTTTCA

<i>Xenichthys xanti</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Brachydeuterus auritus</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Pomadasys maculatus</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Pomadasys argyreus</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Pomadasys kaakan</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Haemulon plumierii</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Haemulon sciurus</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Haemulon steindachneri</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Haemulon flaviguttatum</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Haemulon scudderii</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Haemulon aurolineatum</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Xenistius californiensis</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Plectorhinchus lessonii</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Diplodus bermudensis</i>	TCTATCGGCGGGCTGCTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Pagrus pagrus</i>	TCCATCGGTGGACTACGTCGCTCCTTCCGCTTTCACTTCAGAGGCACGGG
<i>Diagramma pictum</i>	ATACGACGAGAAGATGGTGCGCGAGATGGAAGGGCTCGAGGCCCTCGGGGT
<i>Plectorhinchus vittatus</i>	ATACGACGAGAAGATGGTGCGCGAGATGGAAGGGCTCGAGGCCCTCGGGGT
<i>Plectorhinchus chaetodonoides</i>	ATACGACGAGAAGATGGTGCGCGAGATGGAAGGGCTCGAGGCCCTCGGGGT
<i>Plectorhinchus macrolepis</i>	ATACGACGAGAAGATGGTGCGCGAGATGGAAGGGCTCGAGGCCCTCGGGGT
<i>Haemulopsis elongatus</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Haemulopsis leuciscus</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Anisotremus interruptus</i>	ATATGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCAGGGT
<i>Anisotremus virginicus</i>	ATATGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCAGGGT
<i>Pomadasys branickii</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Anisotremus dovii</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCAGGGT
<i>Haemulopsis axillaris</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Haemulopsis nitidus</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Orthopristis chalceus</i>	ATATGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Anisotremus pacifici</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCAGGGT
<i>Xenichthys xanti</i>	ATATGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Brachydeuterus auritus</i>	ATATGATGAAAAGATGGTGCGAGAAATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Pomadasys maculatus</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTCGAGGCCCTCGGGGT
<i>Pomadasys argyreus</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTCGAGGCCCTCGGGGT
<i>Pomadasys kaakan</i>	ATACGATGAGAAGATGGTGCGAGACATGGAGGGGCTCGAAGCCCTCGGGGT
<i>Haemulon plumierii</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTCGAAGCCCTCGGGGT
<i>Haemulon sciurus</i>	ATACGATGAAAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Haemulon steindachneri</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Haemulon flaviguttatum</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Haemulon scudderii</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTCGAAGCCCTCGGGGT
<i>Haemulon aurolineatum</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTCGAAGCCCTCGGGGT
<i>Xenistius californiensis</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTCGAGGCCCTCGGGGT
<i>Plectorhinchus lessonii</i>	ATACGATGAGAAGATGGTGCGAGAGATGGAGGGGCTTGAGGCCCTCGGGGT
<i>Diplodus bermudensis</i>	AGGCGATGAGAGAATGATACGTGAGATGGCGGCCCTCGTGGCGTCAGGGT
<i>Pagrus pagrus</i>	ATACGACGAGAAGATGGTGCGCGAGATGGAGGGGCTCGAGGCCCTCGGGGT
<i>Diagramma pictum</i>	CCACCTACATCTGCACTCTGTGTGACTCCAGCCGGGCAGAGGCCCTCCGAA
<i>Plectorhinchus vittatus</i>	CCACCTACATCTGCACTCTGTGTGACTCCAGCCGGGCAGAGGCCCTCCGAA
<i>Plectorhinchus chaetodonoides</i>	CCACCTACATCTGCACTCTGTGTGACTCCAGCCGGGCAGAGGCCCTCCGAA
<i>Plectorhinchus macrolepis</i>	CCACCTACATCTGCACTCTATGTGACTCCAGCCGTGACAGAGGCCCTGTAG
<i>Haemulopsis elongatus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCCCTCTCAA
<i>Haemulopsis leuciscus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCCCTCTCAA
<i>Anisotremus interruptus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCGGGGCAGAGGCATCTCAA
<i>Anisotremus virginicus</i>	CCACCTATGTTTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTCAA
<i>Pomadasys branickii</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTCAA
<i>Anisotremus dovii</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTCAA
<i>Haemulopsis axillaris</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTCAA
<i>Haemulopsis nitidus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTCAA
<i>Orthopristis chalceus</i>	CGACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTCAA
<i>Anisotremus pacifici</i>	CGACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTCAA
<i>Xenichthys xanti</i>	CCACCTATGTCTGCACTCCTTGTGACTCCACCCGGGCAGATGCATCTCAA
<i>Brachydeuterus auritus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTCAA
<i>Pomadasys maculatus</i>	CCACCTATGTTTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTGAA
<i>Pomadasys argyreus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTGAA
<i>Pomadasys kaakan</i>	CCACCTATGTTTGCACTCTGTGTGACTCCACAGGGGCAGAGGCATCTGAA
<i>Haemulon plumierii</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTCAA
<i>Haemulon sciurus</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACTCGGGGCAGAGGCATCTCAA
<i>Haemulon steindachneri</i>	CCACCTATGTCTGCACTCTGAGTGACTCCACCCGGGCAGAGGCATCTCAA
<i>Haemulon flaviguttatum</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCAGAGGCATCTCAG
<i>Haemulon scudderii</i>	CCACCTATGTCTGCACTCTGTGTGATTCACCCGGGCAGAGGCATCTCAA
<i>Haemulon aurolineatum</i>	CCACCTATGTCTGCACTCTGTGTGACTCCACTCGGGGCAGAGGCATCTCAA

Xenistius californiensis CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCGAGGGCATCTCAA
Plectorhinchus lessonii CCACCTATGTCTGCACTCTGTGTGACTCCACCCGGGCGAGGGCATCTCAA
Diplodus bermudensis CCACCTATGCTTGCATTCTATGCGACTCCAGTCGAGCGGACTCGTCTCAA
Pagrus pagrus CCACCTACGTCTGCACTCTCTGCGACTCCAGTCGGGCGGAGGGCTCTCAA

Diagramma pictum AACATGGTGCTCCACTCTGTACACGCGAGCCATGAAGAGAACCTGGACCC
Plectorhinchus vittatus AACATGGTGCTCCACTCTGTACACGCGAGCCATGAAGAGAACCTGGACCG
Plectorhinchus chaetodonoides AACATGGTGCTCCACTCTGTACACGCGAGCCATGAAGAGAACCTGGACCG
Plectorhinchus macrolepis AACATGGTGCTTCACTCCATCACACGCGAGCCATGAAGAGAATCTGGACCG
Haemulopsis elongatus AACATGGTGCTTCACTCCATCACACGCGAGCCATGAAGAGAATCTGGACCG
Haemulopsis leuciscus AACATGGTGCTTCACTCCATCACACGCGAGCCATGAAGAGAATCTGGACCG
Anisotremus interruptus AACATGGTGCTGCACTCTGTCCCACGCAACCCACGAGAGAACCTGGACCG
Anisotremus virginicus AACATGGTGCTGCACTCTGTACACGCGAGCCACGAAGAGAACCTGGACCG
Pomadasys branickii AACATGGTGCTGCACTCTGTACACGCAACCCACGAAGAGAACCTGGACCG
Anisotremus dovii AACATGGTGCTGCACTCTGTACACGCGAGCCACGAAGAGAACCTGGACCG
Haemulopsis axillaris AACATGGTGCTTCACTCTGTACACGCTAGCCACGAAGAGAACCTGGACCG
Haemulopsis nitidus AACATGGTGCTTCACTCTGTACACGCTAGCCACGAAGAGAACCTGAACCTG
Orthopristis chalceus AACATGGTGCTGCACTCTGTACACGTAACCCACGAGGAGAACCTGGACCG
Anisotremus pacifici AACATGGTGCTGCACTCTGTACACGCGAGCCACGAAGAGAACCTGGACCG
Xenichthys xanti AACATGGTGCTGCACTCTGTACACGCGAGCCACGAAGAGAACCTGGACCG
Brachydeuterus auritus AACATGGTGCTGCACTCTGTACACGCAACCCACGAAGAGAACCTGGACCG
Pomadasys maculatus AACATGGTGCTGCACTCCATCTCACGCGAGCCACGAAGAGGACCTGGACCG
Pomadasys argyreus AACATGGTGCTGCACTCTATCACACGCGAGCCACGAAGAGAACCTGGACCG
Pomadasys kaakan AACATGGTGCTGCACTCCATCACACGCGAGCCACGAAGAGAACCTGGACCG
Haemulon plumierii AACATGGTGCTGCACTCCATCACACGCAACCCATGAAGAGAACCTGGACCG
Haemulon sciurus AACATGGTGCTGCACTCCATCACACGCAACCCACGAGGAGAACCTGGACCG
Haemulon steindachneri AACATGGTGCTGCACTCCATCACACGCGAGCCACGAAGAGAACCTGGACCG
Haemulon flaviguttatum AACATGGTGCTGCACTCCATCACACGCGAGCCACGAAGAGAACCTGGACCG
Haemulon scudderii AACATGGTGCTGCACTCTATCACACGCAACCCACGAAGAGAACCTGGACCG
Haemulon aurolineatum AACATGGTGCTGCACTCTGTACACGCAACCCACGAAGAGAACCTGGACCG
Xenistius californiensis AACATGGTGCTGCACTCTATCACACGCGGCCACGAAGAGAACCTGGACCG
Plectorhinchus lessonii AACATGGTGCTGCACTCCATCACACGCGAGCCACGAAGAGAACCTGGACCG
Diplodus bermudensis AACATGGTGCTTCCACTCCGTACCCGCAATCAGGACGAGAACAGGGATCG
Pagrus pagrus AACATGGTGCTGCACTCCGTACCCGCAATCAGGACGAGAACCTGGAACG

Diagramma pictum TTACGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGCAGATGAGCTGC
Plectorhinchus vittatus TTACGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGATGAGCTGC
Plectorhinchus chaetodonoides TTACGAAATATGGAGGACCAACCCCTTTTCTGAGTCTGTAGATGAGCTGC
Plectorhinchus macrolepis TTACGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGATGAGCTGC
Haemulopsis elongatus TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCCGTAGAGGAGCTTA
Haemulopsis leuciscus TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCCGTAGAGGAGCTTA
Anisotremus interruptus TTATGAAATATGGAGAACCAACCCCTTTTCTGAATCTGTAGAGGAGCTAC
Anisotremus virginicus TTATGAAATATGGAGAACCAACCCCTTTTCTGAATCTGTAGAGGAGCTAC
Pomadasys branickii TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTGC
Anisotremus dovii TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTAC
Haemulopsis axillaris TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTGC
Haemulopsis nitidus TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTGC
Orthopristis chalceus TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTGC
Anisotremus pacifici TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGCAGAGGAGCTAC
Xenichthys xanti TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTGC
Brachydeuterus auritus TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTGC
Pomadasys maculatus TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTGC
Pomadasys argyreus TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTGC
Pomadasys kaakan TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTGC
Haemulon plumierii TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTAC
Haemulon sciurus TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCCGTAGAGGAGCTAC
Haemulon steindachneri TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGATGAGCTAC
Haemulon flaviguttatum TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTAC
Haemulon scudderii TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTAC
Haemulon aurolineatum TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTAC
Xenistius californiensis TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGAGGAGCTAC
Plectorhinchus lessonii TTATGAAATATGGAGAACCAACCCCTTTTCTGAGTCTGTAGATGAGCTGC
Diplodus bermudensis TTACGAAATCTGGAGCCCCACCCCTTTTCTCGAGTCTGTAGATGAGCTGC
Pagrus pagrus TTACGAAATCTGGAGGACCAACCCCTTTTCTGAGTCCGCAGATGAGCTGC

Diagramma pictum GAGACAGAGTCAAAGGGGTCTTTGACAAGCCCTTCCTGGAGACCCATCCC
Plectorhinchus vittatus GAGACAGAGTCAAAGGGGTCTTACCAAGCCCTTCCTGGAGACCCATCCC
Plectorhinchus chaetodonoides GAGACAGAGTCAAAGGGGTCTTACCAAGCCCTTCCTGGAGACCCATCCC
Plectorhinchus macrolepis GAGACAGAGTCAAAGGTGTCTCTGCCAAGCCCTTCCTGGAGACCCATCCC
Haemulopsis elongatus GAGACAGAGTCAAAGGAGTCTCTGCCAAGCCCTTCATGGAGACCCATCCC
Haemulopsis leuciscus GAGACAGAGTCAAAGGAGTCTCTGCCAAGCCCTTCATGGAGACCCATCCC

<i>Pomadasys argyreus</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Pomadasys kaakan</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Haemulon plumierii</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Haemulon sciurus</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Haemulon steindachneri</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Haemulon flaviguttatum</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Haemulon scudderii</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Haemulon aurolineatum</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Xenistius californiensis</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Plectorhinchus lessonii</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Diplodus bermudensis</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Pagrus pagrus</i>	AATCTTCCAGGATGAGATCGGGGAAGTGTACCAAAAACCCAAACCCGAGCC
<i>Diagramma pictum</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Plectorhinchus vittatus</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Plectorhinchus chaetodonoides</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Plectorhinchus macrolepis</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulopsis elongatus</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulopsis leuciscus</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Anisotremus interruptus</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Anisotremus virginicus</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pomadasys branickii</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Anisotremus dovii</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulopsis axillaris</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulopsis nitidus</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Orthopristis chalceus</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Anisotremus pacifici</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Xenichthys xanti</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Brachydeuterus auritus</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pomadasys maculatus</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pomadasys argyreus</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pomadasys kaakan</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon plumierii</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon sciurus</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon steindachneri</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon flaviguttatum</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon scudderii</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Haemulon aurolineatum</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Xenistius californiensis</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Plectorhinchus lessonii</i>	GAGAGCAACGACGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Diplodus bermudensis</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Pagrus pagrus</i>	GGGAGGAACGGCGCAGCTGGAGGGCAGCCCTAGATAAACAGCTGAGGAAG
<i>Diagramma pictum</i>	AACATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Plectorhinchus vittatus</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Plectorhinchus chaetodonoides</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Plectorhinchus macrolepis</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Haemulopsis elongatus</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Haemulopsis leuciscus</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Anisotremus interruptus</i>	ACGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Anisotremus virginicus</i>	ACGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Pomadasys branickii</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Anisotremus dovii</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Haemulopsis axillaris</i>	AATATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Haemulopsis nitidus</i>	AATATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Orthopristis chalceus</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Anisotremus pacifici</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Xenichthys xanti</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Brachydeuterus auritus</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Pomadasys maculatus</i>	AACATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Pomadasys argyreus</i>	AACATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Pomadasys kaakan</i>	AATATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Haemulon plumierii</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Haemulon sciurus</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Haemulon steindachneri</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Haemulon flaviguttatum</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Haemulon scudderii</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Haemulon aurolineatum</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Xenistius californiensis</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Plectorhinchus lessonii</i>	AAGATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG
<i>Diplodus bermudensis</i>	AAAATGAAGCTTAAAGCCAGTAATGAGGATGAATGGGAACATATGCCCGCCG

<i>Anisotremus dovii</i>	AAGCCTGTGTGGCGCGCCACCTGTCCAGCCAAGGAGTGCCCTGACCAGCT
<i>Haemulopsis axillaris</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCGGCCAAGGAGTGCCCTGACCAGCT
<i>Haemulopsis nitidus</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCGGCCAAGGAGTGCCCTGACCAGCT
<i>Orthopristis chalcus</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCGGCCAAGGAGTGCCCTGACCAGCT
<i>Anisotremus pacifici</i>	AAGCCTGTGTGGCGTGCCACCTGTCCAGCCAAGGAGTGCCCTGACCAGCT
<i>Xenichthys xanti</i>	AAGCCTGTGTGGCGTGCCCTCCTGTCCAGCCAAGGAGTGCCCTGACCAGCT
<i>Brachydeuterus auritus</i>	AAGCCTGTGTGGCGCGCCACCTGTCCGGCCAAGGAGTGTCCTGACCAGCT
<i>Pomadasys maculatus</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCGGCCAAGGAGTGCCCTGACCAGCT
<i>Pomadasys argyreus</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCGGCCAAGGAGTGCCCTGACCAGCT
<i>Pomadasys kaakan</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCGGCCAAGGAGTGCCCTGACCAGCT
<i>Haemulon plumierii</i>	AAGCCTGTGTGTGGCGCGCCCTCCTGTCCAGCCAAGGAGTGCCCCGACCAGCT
<i>Haemulon sciurus</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCAGCCAAGGAGTGCCCCGACCAGCT
<i>Haemulon steindachneri</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCAGCCAAGGAGTGCCCCGACCAGCT
<i>Haemulon flavivittatum</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCAGCCAAGGAGTGCCCCGACCAGCT
<i>Haemulon scudderii</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCAGCCAAGGAGTGCCCTGACCAGCT
<i>Haemulon aurolineatum</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCAGCCAAGGAGTGCCCCGACCAGCT
<i>Xenistius californiensis</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCAGCCAAGGAGTGCCCCGACCAGCT
<i>Plectorhinchus lessonae</i>	AAGCCTGTGTGTGGCGCGCCACCTGTCCAGCCAAGGAGTGCCCCGACCAGCT
<i>Diplodus bermudensis</i>	AAGCCCGTGTGGCGAGCCACCTGTCCCCAGCTAAAGAGTGCCCCGACCAGCT
<i>Pagrus pagrus</i>	AAGCCCGTGTGGCGAGCCAGCTGCCCCGCCAAGGAGTGCCCCGACCAGCT

<i>Diagramma pictum</i>	CCTTCAAGTATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
<i>Plectorhinchus vittatus</i>	CCTTCAAATATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
<i>Plectorhinchus chaetodonoides</i>	CCTTCAAATATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
<i>Plectorhinchus macrolepis</i>	CCTTCAAATATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
<i>Haemulopsis elongatus</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Haemulopsis leuciscus</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Anisotremus interruptus</i>	CCTTTAAATATAGGTACAATGGAAAGATAACCAATTACCTGCACAAGACC
<i>Anisotremus virginicus</i>	CCTTTAAATATAGGTACAATGGAAAGATAACCAATTACCTGCACAAGACC
<i>Pomadasys branickii</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Anisotremus dovii</i>	CCTTTAAATATAGGTACAATGGGAAGATGACCAATTACCTGCACAAGACC
<i>Haemulopsis axillaris</i>	CATTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Haemulopsis nitidus</i>	CATTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Orthopristis chalceus</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Anisotremus pacifici</i>	CCTTTAAATATAGGTACAACGGGAAGATAACCAATTACCTGCACAAGACC
<i>Xenichthys xanti</i>	CCTTTAAATATAGATACAATGGGAAGATAACCAATTATCTGCACAAGACC
<i>Brachydeuterus auritus</i>	CCTTTAAATATAGGTACAATGGAAAAATAACCAATTACCTGCACAAGACC
<i>Pomadasys maculatus</i>	CCTTTAAATATAGGTACAATGGAAAGATAACCAATTACCTGCACAAAACC
<i>Pomadasys argyreus</i>	CCTTTAAATATAGGTACAATGGAAAGATAACCAATTACCTGCACAAGACC
<i>Pomadasys kaakan</i>	CCTTCAAATATAGGTACAATGGAAAGATAACCAATTACCTGCACAAGACC
<i>Haemulon plumierii</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC

<i>Haemulon sciurus</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Haemulon steindachneri</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Haemulon flaviguttatum</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Haemulon scudderii</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Haemulon aurolineatum</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Xenistius californiensis</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Plectorhinchus lessonii</i>	CCTTTAAATATAGGTACAATGGGAAGATAACCAATTACCTGCACAAGACC
<i>Diplodus bermudensis</i>	CCTTCAAATATAGGTACAACGGAAAGATAACCAATTACCTGCACAAGACC
<i>Pagrus pagrus</i>	CCTTCAAGTACAGGTACAACGGAAAGATAACCAACTACCTGCGCAAAACG
<i>Diagramma pictum</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Plectorhinchus vittatus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Plectorhinchus chaetodonoides</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Plectorhinchus macrolepis</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulopsis elongatus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulopsis leuciscus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Anisotremus interruptus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGCTCCATAGGAGCCTG
<i>Anisotremus virginicus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pomadasys branickii</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Anisotremus dovii</i>	CTGGCCCAAGTACCTGAAATCATCAAGAAAGATGGATCCATAGGAGCCTG
<i>Haemulopsis axillaris</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulopsis nitidus</i>	CTGGCCCAAGTACCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Orthopristis chalcus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Anisotremus pacifici</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Xenichthys xanti</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Brachydeuterus auritus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pomadasys maculatus</i>	CTGGCCCAAGTACCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pomadasys argyreus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pomadasys kaakan</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon plumierii</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon sciurus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon steindachneri</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon flaviguttatum</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon scudderii</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Haemulon aurolineatum</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Xenistius californiensis</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Plectorhinchus lessonii</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Diplodus bermudensis</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Pagrus pagrus</i>	CTGGCCCATGTGCCTGAAATCATAGAGAGAGATGGATCCATAGGAGCCTG
<i>Diagramma pictum</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGGGCAGAGT
<i>Plectorhinchus vittatus</i>	GGCCAGTGAGGGGAATGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Plectorhinchus chaetodonoides</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Plectorhinchus macrolepis</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Haemulopsis elongatus</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Haemulopsis leuciscus</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Anisotremus interruptus</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Anisotremus virginicus</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Pomadasys branickii</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Anisotremus dovii</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Haemulopsis axillaris</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Haemulopsis nitidus</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Orthopristis chalcus</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Anisotremus pacifici</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Xenichthys xanti</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Brachydeuterus auritus</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Pomadasys maculatus</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Pomadasys argyreus</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Pomadasys kaakan</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Haemulon plumierii</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Haemulon sciurus</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Haemulon steindachneri</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Haemulon flaviguttatum</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Haemulon scudderii</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Haemulon aurolineatum</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Xenistius californiensis</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Plectorhinchus lessonii</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Diplodus bermudensis</i>	GGCCAGTGAGGGGAACGAGTCGGCAAACAAACTGTTTATCCGAGCAGAAT
<i>Pagrus pagrus</i>	GGCCAGTGAGGGGAATGAGTCGGCAAACAAACTATTTATCCGAGCTGAGC
<i>Diagramma pictum</i>	TAAGCCAACCCGGCGGCTCCTAGGAGACGACCAGATTACAATGTTATC

<i>Orthopristis chalceus</i>	AATTGGCGGATTGGAACTGACTTGTCCCCCTGATGATTGGGGCCCCCTG
<i>Anisotremus pacifici</i>	CATCGGCGGCTTTGGCAACTGGCTTGTGCCCTGATGATCGGGGCCCCCTG
<i>Xenichthys xanti</i>	AATTGGAGGGTTTGGCAACTGACTTGTCCCCCTCATGATTGGTGCCCCCG
<i>Brachydeuterus auritus</i>	CATCGGGGGTTTCGGCAACTGACTTGTTCCTTATGATTGGGGCCCCCTG
<i>Pomadasys maculatus</i>	AATTGGTGGTTTCGGAACTGACTCGTGCCCCCTAATGATTGGAGCGCCTG
<i>Pomadasys argyreus</i>	AATTGGGGGTTTTGGTAACTGACTGGTCCCCCTAATAATTGGGGCCCCCG
<i>Pomadasys kaakan</i>	AATCGGCGGCTTTGGAACTGACTCGTGCCCCCTAATGATCGGGGCCCCCTG
<i>Haemulon plumierii</i>	CATTGGAGGATTTCGGAACTGACTCGTTCCTTATGATTGGGGCCCCCG
<i>Haemulon sciurus</i>	CATTGGAGGGTTTGGAACTGACTCGTTCCTTATGATTGGGGCACCCTG
<i>Haemulon steindachneri</i>	CATCGGAGGATTTCGGAACTGACTCGTTCCTTATGATCGGGGCGCCCG
<i>Haemulon flaviguttatum</i>	CATTGGAGGATTTCGGAACTGACTTGTTCCTTATGATCGGAGCGCCCG
<i>Haemulon scudderii</i>	CATTGGAGGATTTCGGAACTGACTCGTTCCTTATGATTGGGGCCCCCG
<i>Haemulon aurolineatum</i>	CATTGGAGGATTTCGGAACTGACTTGTTCCTTATGATCGGGGCGCCTG
<i>Xenistius californiensis</i>	CATTGGAGGGTTTGGAACTGACTTATCCCCCTATGATCGGGGCGCCCG
<i>Plectorhinchus lessonii</i>	AATTGGAGGATTTCGGAACTGACTGGTCCCATTATAATCGGGGACCCTG
<i>Diplodus bermudensis</i>	GATTGGAGGCTTTGGAACTGACTAATCCCCTTATGATCGGTGGCCCCCTG
<i>Pagrus pagrus</i>	GATTGGAGGCTTTGGAACTGATTAAATCCACTTATGATTGGTGCCCCCTG
<i>Diagramma pictum</i>	ACATGGCATTCCCCGAATGAACAATATGAGTTTCTGACTTCTCCCTCCG
<i>Plectorhinchus vittatus</i>	ACATGGCATTCCCTCGAATGAACAATATGAGCTTCTGACTTCTCCACCA
<i>Plectorhinchus chaetodonoides</i>	ACATGGCATTCCCCGAATAAACAATATGAGCTTCTGACTTCTCCACCA
<i>Plectorhinchus macrolepis</i>	ACATAGCATTCCCTCGAATAAACAACATGAGCTTCTGACTTCTACCCCC
<i>Haemulopsis elongatus</i>	ACATGGCATTCCCGGATAAACAACATGAGTTCTGACTTCTCCCTCC
<i>Haemulopsis leuciscus</i>	ACATGGCATTCCCGGATAAACAACATGAGTTCTGACTTCTCCCTCC
<i>Anisotremus interruptus</i>	ACATGGCATTCCCCGAATAAACAACATGAGCTTCTGGCTCCTCCACCT
<i>Anisotremus virginicus</i>	ATATGGCATTCCCCGAATAAACAATATGAGCTTCTGGCTCCTCCACCT
<i>Pomadasys branickii</i>	ACATGGCATTCCCCCGGATAAACAACATGAGCTTCTGGCTCCTCCCTCC
<i>Anisotremus dovii</i>	ACATAGCATTCCCTCGAATGAACAACATGAGTTCTGGCTCCTTCCCTCC
<i>Haemulopsis axillaris</i>	ACATAGCATTCCCTCGGATAAATAATATGAGCTTCTGACTCCTCCCTCC
<i>Haemulopsis nitidus</i>	ACATAGCATTCCCTCGGATAAATAATATGAGCTTCTGACTCCTCCCTCC
<i>Orthopristis chalceus</i>	ACATAGCATTCCCGGATAAACAACATGAGCTTCTGACTCCTCCCTCC
<i>Anisotremus pacifici</i>	ACATGGCTTTCCTCGAATGAACAACATGAGCTTTGGCTGCTCCCCCT
<i>Xenichthys xanti</i>	ACATGGCATTCCCCGAATAAATAACATGAGCTTTGGCTGCTCCGCT
<i>Brachydeuterus auritus</i>	ACATAGCATTCCCTCGAATAAATAATATGAGCTTTTGACTTCTACCTCT
<i>Pomadasys maculatus</i>	ATATGGCATTCCCTCGGATGAACAACATGAGTTTCTGACTCCTCCCTCC
<i>Pomadasys argyreus</i>	ACATGGCCTTCCCTCGAATAAATAATATAAGCTTCTGACTCCTCCACCC
<i>Pomadasys kaakan</i>	ACATAGCATTCCCTCGGATGAACAACATGAGCTTCTGACTCCTCCCTCC
<i>Haemulon plumierii</i>	ACATGGCATTCCCCGAATGAATAACATGAGCTTTTGACTCCTCCCTCT
<i>Haemulon sciurus</i>	ATATGGCATTCCCCGAATGAATAACATGAGCTTTTGACTCCTCCCTCC
<i>Haemulon steindachneri</i>	ATATGGCATTCCCGGATAAACAACATGAGCTTTTGACTCCTCCCTCT
<i>Haemulon flaviguttatum</i>	ACATGGCATTCCCCGAATGAACAATATGAGCTTTTGACTCCTCCCTCT
<i>Haemulon scudderii</i>	ACATGGCATTCCCCGGATGAATAACATGAGCTTTTGACTCCTCCCTCT
<i>Haemulon aurolineatum</i>	ATATGGCATTCCCGGATAAACAACATGAGCTTTTGACTCCTCCCTCA
<i>Xenistius californiensis</i>	ATATGGCATTCCCCGAATGAACAACATGAGCTTCTGACTCCTCCCTCT
<i>Plectorhinchus lessonii</i>	ACATGGCATTCCCTCGAATGAACAATATGAGCTTCTGACTTCTCCACCA
<i>Diplodus bermudensis</i>	ACATAGCATTCCCCGAATAAATAACATGAGCTTCTGACTTCTGCCCTCC
<i>Pagrus pagrus</i>	ACATAGCATTCCCCGAATGAACAACATGAGTTCTGACTTCTCCCTCC
<i>Diagramma pictum</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGCGTAGAAGCCGGGCGAGGAC
<i>Plectorhinchus vittatus</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGCGTAGAAGTGGAGCAGGGAC
<i>Plectorhinchus chaetodonoides</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGCGTAGAAGCCGGAGCAGGAAC
<i>Plectorhinchus macrolepis</i>	TCATTCTTCTCCTTGCCTCCTCAGGAGTTGAGGCGGGAGCTGGGAC
<i>Haemulopsis elongatus</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGGGTGGAGGCCGGTGCCGGTAC
<i>Haemulopsis leuciscus</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGGGTGGAGGCCGGTGCCGGTAC
<i>Anisotremus interruptus</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGCGTAGAAGCCGGAGCCGGTAC
<i>Anisotremus virginicus</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGCGTAGAAGTGGGGCCGGCAC
<i>Pomadasys branickii</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGGGTAGAAGCCGGGCTGGTAC
<i>Anisotremus dovii</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGGGTAGAAGCAGGGCTGGTAC
<i>Haemulopsis axillaris</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGGGTAGAAGCCGGCGCCGGCAC
<i>Haemulopsis nitidus</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGGGTAGAAGCCGGCGCCGGCAC
<i>Orthopristis chalceus</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGTGTGAAGCCGGAGCTGGTAC
<i>Anisotremus pacifici</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGGGTAGAAGCAGGGCTGGGAC
<i>Xenichthys xanti</i>	TCCTTCCTTCTCCTTGCCTCCTCAGGGGTAGAAGCCGGGCGAGGAAC
<i>Brachydeuterus auritus</i>	TCATTCTTCTTCTTGCCTCATCAGGCGTAGAAGCAGGGGCTGGGAC
<i>Pomadasys maculatus</i>	TCCTTCCTTCTTCTTGCCTCCTCAGGGGTGAGGCTGGGGCCGGAAC
<i>Pomadasys argyreus</i>	TCCTTCCTTCTTCTTGCCTCATCAGGTGTGAAGCTGGGGCAGGGAC
<i>Pomadasys kaakan</i>	TCCTTCCTTCTTCTTGCCTCCTCAGGGGTGAGGCGGGGCTGGAAC
<i>Haemulon plumierii</i>	TCCTTCCTTCTTCTTGCCTCCTCAGGCGTAGAGGCTGGGGCCGGGAC
<i>Haemulon sciurus</i>	TCCTTCCTTCTTCTTGCCTCCTCAGGCGTAGAGGCTGGGGCCGGGAC
<i>Haemulon steindachneri</i>	TCCTTCCTTCTTCTTGCCTCCTCAGGGGTAGAGGCTGGAGCTGGAAC
<i>Haemulon flaviguttatum</i>	TCCTTCCTTCTTCTTGCCTCCTCAGGGGTAGAGGCTGGGGCTGGAAC

Haemulon scudderii
Haemulon aurolineatum
Xenistius californiensis
Plectorhinchus lessonii
Diplodus bermudensis
Pagrus pagrus

Diagramma pictum
Plectorhinchus vittatus
Plectorhinchus chaetodonoides
Plectorhinchus macrolepis
Haemulopsis elongatus
Haemulopsis leuciscus
Anisotremus interruptus
Anisotremus virginicus
Pomadasys branickii
Anisotremus dovii
Haemulopsis axillaris
Haemulopsis nitidus
Orthopristis chalcus
Anisotremus pacifici
Xenichthys xanti
Brachydeuterus auritus
Pomadasys maculatus
Pomadasys argyreus
Pomadasys kaakan
Haemulon plumierii
Haemulon sciurus
Haemulon steindachneri
Haemulon flaviguttatum
Haemulon scudderii
Haemulon aurolineatum
Xenistius californiensis
Plectorhinchus lessonii
Diplodus bermudensis
Pagrus pagrus

Diagramma pictum
Plectorhinchus vittatus
Plectorhinchus chaetodonoides
Plectorhinchus macrolepis
Haemulopsis elongatus
Haemulopsis leuciscus
Anisotremus interruptus
Anisotremus virginicus
Pomadasys branickii
Anisotremus dovii
Haemulopsis axillaris
Haemulopsis nitidus
Orthopristis chalcus
Anisotremus pacifici
Xenichthys xanti
Brachydeuterus auritus
Pomadasys maculatus
Pomadasys argyreus
Pomadasys kaakan
Haemulon plumierii
Haemulon sciurus
Haemulon steindachneri
Haemulon flaviguttatum
Haemulon scudderii
Haemulon aurolineatum
Xenistius californiensis
Plectorhinchus lessonii
Diplodus bermudensis
Pagrus pagrus

Diagramma pictum
Plectorhinchus vittatus
Plectorhinchus chaetodonoides
Plectorhinchus macrolepis

TCTTTCTCTCTCTCTCTCGCCTCTTCAGGAGTAGAGGCCGGGGCTGGGAC
TCTTTCTCTCTCTCTCTTGCCTCTTCAGGCGTAGAAGCTGGGGCTGGGAC
TCTTTCTCTCTCTCTCTCGCCTCTTCAGGAGTAGAGGCCGGGGCGGGAAC
TCCTTCTCTCTCTCTCTTGCCTCTTCAGGCGTAGAAGCTGGAGCAGGGAC
TCATTCTCTCTCTCTGCTAGCCTCGTCCGGAGTTAGGGCTGGGGCGGTAC
TCATTCTCTCTCTACTTGCCTCTTCAGGAGTTGAAGCCGGAGCTGGCAC

TGGTTGAACAGTCTACCCCCCGTTGGCCGGAATTTAGCGCACGCAGGTG
TGGTTGAACAGTCTACCCCCCACTAGCCGGCAATTTAGCGCACGCAGGAG
TGGTTGAACAGTCTACCCCCCATTTGGCCGGTAATCTGGCGCACGCAGGTG
AGGATGAAGTGTATATCCACCCTGGCTGGCAACCTAGCCACGCTGGGG
TGGGTGGACAGTCTACCCGCCCTGGCCGGGAACCTCGCCCATGCAGGAG
TGGGTGGACAGTCTACCCGCCCTGGCCGGGAACCTCGCCCATGCAGGAG
CGGGTGGACAGTCTACCTTCCCTTAGCCGGGAACCTAGCTCAGCCGGAG
CGGATGAACAGTTTACCCTCCCTTAGCCGGAAATCTAGCGCATGCCGGGG
GGGGTGGACAGTTTACCCTCCTTAGCCGGAAACCTAGCCACGCGGGGG
CGGATGGACAGTCTATCCCCCTTAGCTGGGAACCTAGCTCATGCCGGGG
AGGATGAACAGTCTACCTTCCCTTAGGCTGGAACTTAGCCACGCGGGAG
AGGATGGACAGTTTACCCTCCCTTAGCCGGGAACCTTAGCCCATGCCGGAG
AGGGTGGACCGTATACCCGCCCTTAGCCGGGAACCTGGCACACGCCGGAG
CGGATGAACAGTTTACCCTCCTTAGCCGGGGAACTGGCCACGCAGGTG
CGGATGAACAGTTTACCCTCCCTTAGCCGGCAACCTAGCCACGCAGGAG
CGGATGAACAGTCTACCCCTCTTAGCTGGAAACCTAGCCACGCAGGGG
TGGGTGGACAGTATACCCCTCTTAGCCGGGAACCTGGCTCAGCGAGGAG
TGGGTGGACTGTTTACCCTCCCTTAGCCGGGAATCTAGCACATGCTGGGG
GGGGTGAACAGTCTTACCCTCCCTTAGCCGGGAACCTAGCACACGCTGGGG
GGGGTGAACAGTCTTACCCTCCCTTAGCCGGGAATCTGGCACACGCCGGGG
GGGGTGAACAGTCTTACCCTCCCTTAGCTGGGAATCTAGCACATGCCGGGG
TGGGTGAACAGTCTTACCCTCCCTTAGCTGGTAACCTAGCCACGCAGGAG
TGGATGAACAGTCTTACCCTCCCTTAGCTGGTAACCTAGCCACGCAGGAG
CGGATGAACAGTTTATCCGCCACTAGCTGGGAATCTTGCCACGCAGGAG

CATCTGTAGACCTCAGCATCTTCTCCCTACATCTGGCCGGTATCTCATCA
CATCTGTTGACCTAACAATCTTCTCTCTCATCTGGCCGGTATCTCCTCA
CATCTGTTGACCTAACAATCTTTTCCCTTCATCTGGCCGGTATCTCCTCA
CATCTGTTGACCTCACCATCTTCTCCCTACACCTAGCAGGTGTTCTCTCA
CATCTGTGACCTGACCATCTTCTCCCTCCACTTAGCTGGTGTCTCTCTCA
CATCTGTGACCTGACCATCTTCTCCCTCCACTTAGCTGGCGTCTCCTCA
CATCTGTGACCTAACAATTTTCTCCCTTCACCTAGCAGGTGTTCTCTCA
CATCCGTTGACCTAACAATTTTCTCCCTCCACTTAGCAGGTGTTCTCTCA
CATCCGTTGACCTAACAATTTTCTCCCTTCACTTGGCAGGTGTCTCTCA
CATCCGTTGACCTGACAATTTTCTCCCTCCACTTAGCAGGGGTCTCTCT
CATCTGTTGACCTGACAATCTTTTCACTTCACCTAGCAGGGGTATCATCT
CATCTGTTGACCTGACAATCTTTTCACTTCACCTAGCAGGGGTATCATCT
CATCCGTTGACCTAACAATTTTCTCTCTTCACTAGCAGGTGTCTCTCTCA
CTTCCGTTGACTTAACCATCTTCTCCCTTCATCTGACAGGTGTATCTCTCA
CATCAGTTGACCTAACAATCTTTTCTCTCCACTTAGCAGGGGTCTCTCTCA
CCTCGGTAGACCTGACCATCTTTTCTCTGCATCTAGCAGGTGTTCTCTCA
CATCAGTTGACCTAACCATTTTCTCCCTTCACTTGGCCGGTGTCTCTCTCA
CATCAGTCGACCTGACAATTTTCTCACTTCACCTCGCAGGTGTTCTCTCA
CTTCCGTTGACCTAACAATCTTCTCCCTTCACTTGGCAGGTGTCTCTCTCA
CATCAGTTGACCTTACAATTTTCTCCCTTCACTTAGCAGGTGTCTCATCA
CATCAGTTGACCTTACAATCTTTTCCCTTCACTTAGCAGGTGTCTCGTCA
CATCGGTTGATCTTACAATCTTCTCCCTTCACTTAGCAGGTGTCTCATCA
CGTCCGTTGACCTTACAATCTTCTCCCTCCACTTAGCAGGTGTCTCATCG
CATCAGTTGATCTTACAATTTTCTCCCTCCACTTAGCAGGGGTCTCATCC
CATCAGTTGACCTTACAATCTTCTCCCTCCACTTAGCAGGTGTCTCATCA
CATCAGTTGACCTCACAATTTTCTCTCTCCACTTAGCAGGTGTCTCATCA
CATCTGTTGACCTAACAATCTTCTCTCTTCATCTGGCCGGTATCTCTCTCA
CATCAGTTGACTTAACATCTTTTCTCTTCACTTGGCCGGAAATTCATCT
CATCAGTAGACCTAACCATCTTTTCTCTCCACTAGCTGGAATCTCATCA

ATTCTCGGGGCGATTAATTTTATTACAACAATCATTAACATGAAACCCCC
ATTCTCGGGGCAATTAATTTTATTACAACAATCATTAACATGAAACCCCC
ATTCTTGAGGCAATCAATTTTATTACAACAATTTAATCAATGAAACCCCC
ATTTTAGGGGCTATTAACTTTATTACAACCATCTTAACATGAAACCTCC

<i>Plectorhinchus lessonii</i>	GGGAGGGGATCCAATTCTCTA
<i>Diplodus bermudensis</i>	GGGAGGAGACCCGATTCTTTA
<i>Pagrus pagrus</i>	AGGAGGGGACCCAATTCTCTA