Old Dominion University [ODU Digital Commons](https://digitalcommons.odu.edu/)

[Biological Sciences Faculty Publications](https://digitalcommons.odu.edu/biology_fac_pubs) **Biological Sciences** Biological Sciences

2013

Fire, Hurricane and Carbon Dioxide: Effects on Net Primary Production of a Subtropical Woodland

Bruce A. Hungate

Frank P. Day Old Dominion University

Paul Dijkstra

Benjamin D. Duval

C. Ross Hinkle

See next page for additional authors

Follow this and additional works at: [https://digitalcommons.odu.edu/biology_fac_pubs](https://digitalcommons.odu.edu/biology_fac_pubs?utm_source=digitalcommons.odu.edu%2Fbiology_fac_pubs%2F249&utm_medium=PDF&utm_campaign=PDFCoverPages)

C Part of the [Plant Sciences Commons](https://network.bepress.com/hgg/discipline/102?utm_source=digitalcommons.odu.edu%2Fbiology_fac_pubs%2F249&utm_medium=PDF&utm_campaign=PDFCoverPages)

Original Publication Citation

Hungate, B. A., Day, F. P., Dijkstra, P., Duval, B. D., Hinkle, C. R., Langley, J. A., . . . Drake, B. G. (2013). Fire, hurricane and carbon dioxide: Effects on net primary production of a subtropical woodland. New Phytologist, 200(3), 767-777. doi:10.1111/nph.12409

This Article is brought to you for free and open access by the Biological Sciences at ODU Digital Commons. It has been accepted for inclusion in Biological Sciences Faculty Publications by an authorized administrator of ODU Digital Commons. For more information, please contact [digitalcommons@odu.edu.](mailto:digitalcommons@odu.edu)

Authors

Bruce A. Hungate, Frank P. Day, Paul Dijkstra, Benjamin D. Duval, C. Ross Hinkle, and J. Adam Langley

This article is available at ODU Digital Commons: https://digitalcommons.odu.edu/biology_fac_pubs/249

Fire, hurricane and carbon dioxide: effects on net primary production of a subtropical woodland

Bruce A. Hungate 1 , Frank P. Day 2 , Paul Dijkstra 1 , Benjamin D. Duval 1,3 , C. Ross Hinkle 4 , J. Adam Langley 5 , J. Patrick Megonigal 6 , Peter Stiling 7 , Dale W. Johnson 8 and Bert G. Drake 6

¹Department of Biological Sciences and Ecosystem Science & Society Center, Northern Arizona University, Flagstaff, AZ, 86011, USA; ²Department of Biological Sciences, Old Dominion University, Norfolk, VA 23529, USA; ³USDA-ARS, Dairy Forage Research Center, 1925 Linden Dr, Madison, WI, 53706, USA; ⁴University of Central Florida, Orlando, FL 32816, USA; ⁵Department of Biology, Villanova University, Villanova, PA 19085, USA; ⁶Smithsonian Environmental Research Center, Edgewater, MD 21037, USA; ⁷University of South Florida, Tampa, FL 33620, USA; ⁸Department of Natural Resources, University of Nevada-Reno, Reno, NV 89557, USA

Author for correspondence: Bruce A. Hungate Tel: +1 928 699 3998 Email: Bruce.Hungate@nau.edu

Received: 16 April 2013 Accepted: 16 June 2013

New Phytologist (2013) 200: 767–777 doi: 10.1111/nph.12409

Key words: disturbance, elevated $CO₂$, fire, global environmental change, hurricane, net primary productivity (NPP), nitrogen cycling, oak woodland.

Summary

 Disturbance affects most terrestrial ecosystems and has the potential to shape their responses to chronic environmental change.

• Scrub-oak vegetation regenerating from fire disturbance in subtropical Florida was exposed to experimentally elevated carbon dioxide (CO₂) concentration (+350 μ II⁻¹) using open-top chambers for 11 yr, punctuated by hurricane disturbance in year 8. Here, we report the effects of elevated $CO₂$ on aboveground and belowground net primary productivity (NPP) and nitrogen (N) cycling during this experiment.

• The stimulation of NPP and N uptake by elevated $CO₂$ peaked within 2 yr after disturbance by fire and hurricane, when soil nutrient availability was high. The stimulation subsequently declined and disappeared, coincident with low soil nutrient availability and with a $CO₂$ induced reduction in the N concentration of oak stems.

 \bullet These findings show that strong growth responses to elevated CO₂ can be transient, are consistent with a progressively limited response to elevated $CO₂$ interrupted by disturbance, and illustrate the importance of biogeochemical responses to extreme events in modulating ecosystem responses to global environmental change.

Introduction

Rising atmospheric carbon dioxide $(CO₂)$ and nitrogen (N) deposition increase directly the resource availability to plants, and can increase plant growth and net primary productivity (NPP; Norby et al., 2005). Natural disturbance also alters resource availability to plants, creating space for colonization and reducing competition for light after the catastrophic removal of biomass (Sousa, 1984), and mobilizing nutrients bound in organic matter to more available mineral forms (Martin et al., 2012; Block et al., 2013; Michalzik & Martin, 2013). Global environmental change occurs against a background of episodic disturbance and recovery, both altering resource availability and thus ecosystem processes. Despite the importance of the potential interactions between global environmental change and disturbance (Körner, 2006; Leuzinger et al., 2011), very few experiments have evaluated how responses of ecosystems to elevated CO₂ interact with disturbance (Henry et al., 2006; Niboyet et al., 2011; Brown et al., 2012).

The response of NPP to elevated $CO₂$ varies, at least in part, because the availability of other resources influences the response (Field et al., 1992; de Graaff et al., 2006; Reich et al., 2006a,b;

 2013 The Authors New Phytologist © 2013 New Phytologist Trust

Wang et al., 2007). Elevated $CO₂$ can cause a sustained stimulation of NPP (Norby et al., 2005; Drake et al., 2011; Zak et al., 2011), with interannual variation caused by precipitation (Owensby et al., 1999; Smith et al., 2000; Niklaus & Körner, 2004; Seiler et al., 2009) and salinity (Rasse et al., 2005). The stimulation of NPP by elevated $CO₂$ can be reduced or absent when nutrient availability is chronically low (Schäppi & Körner, 1996; Dukes et al., 2005; Reich et al., 2006a,b; Reich & Hobbie, 2012), or when nutrient availability declines over time (Norby et al., 2010) as a result of progressive nutrient limitation, a theoretically inevitable influence on the response of NPP to elevated $CO₂$ (or to any growth enhancement), as increased growth and nutrient accumulation in organic matter reduce the nutrient supply to plants (Field, 1999; Luo et al., 2004). N is often the focal element in discussions of progressive nutrient limitation, but the concept applies to any nutrient element limiting to plant growth where the decomposition of plant litter is a major source of the element for plants (e.g. phosphorus, P). Increases in productivity under elevated CO₂ by higher N-use efficiency (NUE; Calfapietra et al., 2007; Finzi et al., 2007; Norby et al., 2010) can mitigate progressive nutrient limitation via flexibility in plant element stoichiometry. Increased nutrient uptake with elevated $CO₂$ indicates

the mobility and redistribution of nutrients from soils to plants (Johnson et al., 2006; Drake et al., 2011), increased nutrient inputs (Zanetti & Hartwig, 1997) or reduced nutrient losses (Hagedorn et al., 2005), all of which can alleviate progressive nutrient limitation (Luo et al., 2004). In addition, episodic disturbance could reset progressive nutrient limitation by interrupting the plantmediated accrual of nutrients in organic matter and converting nutrients to available forms. For example, fire disturbance alleviated the suppressive effect of elevated $CO₂$ on NPP in an annual grassland, possibly by increasing P availability (Henry et al., 2006).

Here, we report the effects of 11 yr of experimental $CO₂$ enrichment on NPP and N cycling in a subtropical scrub-oak woodland. Our goal in this work was to evaluate the response of NPP in this scrub-oak woodland to elevated $CO₂$ after recovery from fire disturbance. A hurricane struck the experiment 8 yr after the fire, providing an additional opportunity to assess the role of disturbance in modulating the responses to elevated $CO₂$.

Materials and Methods

This work was conducted at the Kennedy Space Center, Cape Canaveral, FL, USA (28°38′N, 80°42′W) in a stand of scrub-oak vegetation managed for a fire return cycle approximately every decade (Schmalzer & Hinkle, 1992). In the 20th century, hurricanes made landfall in the state 35 times (Smith, 1999), and thus hurricanes are another frequent agent of disturbance at the site (Li et al., 2007). The climate at the site is subtropical (Mailander, 1990). The 30-yr mean maximum and minimum temperatures are 22.3 and 9.6°C for January and 33.3 and 21.9°C for July. The 100-yr mean precipitation is 1310 mm yr⁻¹, most falling in the wet season from June to October.

The experiment consisted of 16 open-top chambers, built of PVC frames with Mylar windows. The chambers were 2.5 m high with an octagonal surface area of 9.42 m^2 . Blowers distributed air through the chambers, either ambient air or air supplemented with $+350 \mu$ l CO₂ l⁻¹. The soils at the site are classified as Arenic Haplahumods and Spodic Quartzipsamments, are sandy with a low pH $(c.$ pH 4). The vegetation is coastal scruboak palmetto, dominated by three oaks (Quercus myrtifolia Willd., Q. geminata Small and Q. chapmanii Sargenti), which accounted for 96% of the aboveground biomass at the site before the experiment began (Dijkstra et al., 2002). Less abundant species include Serenoa repens Small, Vaccinium myrsinites Lam., Lyonia ferruginea Nutt., Befaria racemosa Vent. and Galactia elliottii Nutt. (Dijkstra et al., 2002; Johnson et al., 2003; Seiler et al., 2009). Many of the species at the site, including the three oaks, resprout from rhizomes after fire disturbance (Webber, 1935; Schmalzer & Hinkle, 1992; Guerin, 1993).

Controlled burns at the site were conducted in August 1995 and January 1996. Biomass that accumulated after the January burn and was clipped at ground level immediately before initiating the $CO₂$ treatments. Thus, recovery from the first major disturbance at the site began with treatment initiation on 14 May 1996. In the fall of 2004, the site was struck by a severe hurricane (Frances) on 5 September 2004, with sustained wind speeds of 169 km h^{-1} (category II). The storm subsided on 9

September 2004. These dates were used as the time-zero points for our analysis of the responses of the ecosystem to elevated $CO₂$ as a function of disturbance.

Biomass, carbon and nitrogen

Total aboveground biomass was estimated from annual surveys in which the diameter of each individual oak stem was measured in all plots (Dijkstra et al., 2002; Seiler et al., 2009). Total aboveground biomass was calculated using allometric relationships developed previously, relating the stem diameter and total mass for each of the three oak species (Seiler et al., 2009). We developed new relationships to describe the partitioning of aboveground biomass into leaf and stem mass. Oak stems collected before the first burn in 1995 and at the final harvest in 2007 were divided into leaves and stems, the dry mass of each fraction was determined, and the leaf mass fraction was calculated as the mass of leaves divided by the total aboveground biomass. Data were binned into increments of $0.1 \log_e$ (diameter), and the relationship between leaf mass fraction and loge (diameter) was determined using linear regression on the binned means (Table 1).

Coarse root biomass was measured directly through a combination of coring and ground penetrating radar in 2002, 2005 and for the final harvest in 2007 (Brown et al., 2007; Stover et al., 2007; Day et al., 2013). For years in which direct measurements were not available, coarse root biomass was estimated from aboveground stem biomass, based on the relationship derived from the simultaneously measured values of both variables:

Coarse root biomass = stem biomass \times 1.9845 + 3132 g m⁻². Eqn 1

The regression was significant (intercept, $P < 0.001$; slope, $P=0.025$), but the coefficient of determination was low $(r^2 = 0.16)$, and the standard errors were non-negligible: 0.8403 for the slope and 797 for the intercept. With carbon (C) turnover through coarse roots estimated at $0.03 \,\mathrm{yr}^{-1}$ (based on the ¹³C tracer described below), it is unlikely that coarse root NPP is extremely dynamic. Even so, these should be considered as rough estimates of coarse root biomass and NPP, and viewed with appropriate caution.

Table 1 Allometric equations^a predicting leaf mass fraction (leaf mass/total aboveground mass) from stem diameter for each of the three oak species

Species	Slope	Intercept	r^2	N(n)
Quercus myrtifolia	-0.4129	-0.2767	0.921	29(655)
Quercus chapmanii	-0.4345	-0.1660	0.900	30 (301)
Quercus geminata	-0.4715	-0.0269	0.890	15(82)

^a Equations are of the form: log_e (leaf mass fraction) = slope \times log_e(stem diameter) + intercept. Coefficient of determination (r^2) is shown, together with the number of samples included in the regression (binned data, N) and in the total sample (number in parentheses, n).

Fine roots were monitored throughout the experiment using cores and minirhizotron images. Fine root samples were collected from cores in 1997, 1998, 1999, 2000, 2001, 2002 and 2007. Frequent sampling was suspended in 2002 to avoid excessive destructive coring in the plots. Surface cores (0–15 cm) were collected during the first 5 yr. More extensive depth profiles were sampled in 2002 (to 1 m) and 2007 (to 2–3 m, depending on the depth of the Bh horizon in each plot). Roots were handpicked, washed, oven dried and weighed. These data are presented elsewhere (Langley et al., 2003; Day et al., 2006; Stover et al., 2007; Brown et al., 2009), but are included here as essential components for the calculation of NPP and N cycling. When multiple measurements of fine root biomass were available for a given year, we used the average in our calculations of NPP and N requirement. The average was selected because not all years had sufficient data to identify the annual maximum. The higher frequency of minirhizotron observations gives this technique more influence in the calculations relative to data from the soil cores.

Leaf and stem tissue samples were collected each year for Q. myrtifolia, in each year except 2005 for Q. geminata, and in each year except 2000, 2001 and 2005 for Q. chapmanii. Element concentrations in Q. chapmanii and Q. myrtifolia were similar, and so the values of the latter were used when measurements of the former were unavailable. For Q. geminata in 2005, values were estimated as the average values from 2004 and 2006 for each individual plot. Samples of fine roots for tissue analysis were available from the core samples taken in 1997–2002 and 2007. Coarse roots for tissue analyses were collected from the 1–3-mdeep cores in 2002 and 2007.

Percentage C, %N, δ^{13} C and δ^{15} N of tissue samples were determined at the Colorado Plateau Analytical Laboratory using an elemental analyzer (Costech CE 2100, Valencia, CA, USA) coupled to an isotope ratio mass spectrometer (ThermoFinnigan Delta-PLUS, Delta-PLUS XL or Delta-V, Bremen, Germany). Plant element mass (g C or g N m $^{-2})$ was calculated as the product of biomass and element concentration for each plant component (leaves, stems, fine roots, coarse roots, litterfall). A ^{15}N tracer was applied directly to the soil surface of all plots in June 1998 at a rate of $0.18 \text{ g}^{15} \text{N m}^{-2}$ as aqueous 0.1 g N 1^{-1} $(^{15}NH_4)_2$ SO₄ (99 atom% ^{15}N) using hand-held sprayers. For all organic matter samples collected, tracer ^{15}N mass (mg ^{15}N m⁻²) was calculated as the atom% excess ¹⁵N concentration (measured atom% 15N minus natural abundance 15N) times the mass of N $(g N m^{-2})$ in the organic matter compartment in question. C, N and 15N in coarse roots were measured directly in 2002 and 2007, and in fine roots in 1998, 1999, 2001, 2002 and 2007; values for the other years were interpolated linearly through time.

Biomass increment and litter production

The annual increments in plant biomass, C and N were calculated for leaves, stems, fine and coarse roots as the difference in average biomass (or C or N) for year $t+1$ minus average biomass in year t.

The annual flux of leaf litter was measured from litterfall collections. Litter traps were placed in each plot and monitored at a

frequency no less than four times per year throughout the experiment. Litter that fell into the traps was sorted by species, dried and weighed (Johnson et al., 2003; Hungate et al., 2006; Stiling et al., 2009). Before 2002, Q. myrtifolia and Q. chapmanii were not distinguished in the sorting; for this period, we estimated litterfall as a proportion of leaf mass for these species. Stem litter production was assumed to be zero, based on observations of littertrap contents in which stem tissue was never observed. This is an underestimate because standing dead biomass was recovered at the end of the experiment. However, it was a small fraction of the total chamber biomass $(4.5 \pm 1.2\%)$, and did not impact appreciably on our estimates of productivity.

Fine and coarse root litter production were calculated as the product of biomass and turnover rate. Turnover rates of fine and coarse roots were measured using a C tracer approach. The fossilderived CO_2 added to the elevated CO_2 -treated plots was depleted in ¹³C relative to atmospheric CO_2 , providing a C isotope tracer: the δ^{13} C of plant C fixed after initiation of the CO₂ treatment was c. 15% lower than that of organic C already present in the ecosystem before the experiment began (Langley et al., 2002). We used the initial rate of incorporation of tracer-derived C into fine and coarse roots to estimate root litter production in $gC m^{-2} yr^{-1}$. Fine roots were collected from ingrowth cores in December 1998 and measured for δ^{13} C (Langley *et al.*, 2002); coarse roots were collected in April 2002 and also measured for δ^{13} C. Coarse root tissue from the ambient CO₂ plots was used as a proxy for the δ^{13} C value of 'old' C, and stem tissue from the elevated $CO₂$ plots was used as an integrative measure of the δ^{13} C value of new C input to the ecosystem incorporating the depleted 13 C tracer. C turnover (*T*) through roots was calculated as:

$$
\frac{T = (\delta^{13}CRe - \delta^{13}CRa)}{\delta^{13}CSe - \delta^{13}CSa)^{\frac{1}{t}}}, \qquad \text{Eqn 2}
$$

where $\delta^{13}C$ is the C isotope signature of roots (R) or stems (S) from the elevated (e) or ambient (a) treatments, and t is the duration of the experiment up to the time of sampling – the time during which the depleted ¹³C tracer could be incorporated into root tissue. Using this approach, C turnover through fine roots was 0.247 yr^{-1} , and through coarse roots was 0.030 yr^{-1} . These rates are substantially lower than those estimated for fine roots from minirhizotron observations (Stover et al., 2010), a discrepancy often observed (Strand et al., 2008). We favored the approach based on isotopic tracer estimates of C flow, because it is more relevant to ecosystem–atmosphere C exchange (Strand et al., 2008).

Net primary productivity, nitrogen cycling and nitrogen-use efficiency

NPP was calculated as the sum of the net biomass increment and litter production. NPP was expressed in units of biomass (g biomass m^{-2} yr⁻¹) and in units of C mass (g C m⁻² yr⁻¹). The N requirement was calculated as the sum of the N increment and N loss in litter production in all measured plant components.

Retranslocation of N during leaf senescence was calculated as the live-equivalent mass of N in litterfall minus the actual N flux of senesced leaves in litterfall (for further details, see Hungate et al., 2006). Retranslocation from fine roots was assumed to be negligible. N uptake was calculated as N requirement minus retranslocation. We estimated NUE as the slope of the relationship between NPP and N uptake, and measured the components of NUE: N productivity was calculated as NPP (g biomass $\mathrm{m}^{-2}\,\mathrm{yr}^{-1}$) divided by total plant N content (g N m^{-2}), and N residence time (yr) was calculated as total plant N content $\rm (g\,N\,m^{-2})$ divided by total plant N uptake (g N m $^{-2}$ yr $^{-1}$).

Soil nutrients

We determined K₂SO₄ extractable N in mineral soil (0–15 cm), the sum of extractable inorganic and organic N, at multiple times during the experiment (July 1997; June, July, September and December 1998; September 1999; and May 2004). Soil subsamples (20–25 g at field moisture content) were extracted in 75 ml $0.5 M K₂SO₄$ and filtered through a paper filter, and extracts were dried at 60°C, ground in a mortar and pestle, and the resulting powder was analyzed for %N by Dumas combustion (http://www.isotope.nau.edu). Extractable N is expressed as $g N m^{-2}$ ground area using the surface bulk density at the site of 0.83 g cm^{-3} (Hungate *et al.*, 2006). Previously published data on soil P (Johnson et al., 2003) are presented and re-analyzed here as a function of time since disturbance, to assess the role of disturbance on soil nutrient availability (see Data analysis, below).

Data analysis

We compared the importance of increased N uptake and increased NUE in the productivity response to elevated $CO₂$. The hypothetical response driven by increased N uptake (ΔNPP_{u}) was calculated as:

$$
\Delta \text{NPP}_u = \Delta U \times E_a, \qquad \text{Eqn 3}
$$

where ΔU is the effect of elevated $CO₂$ on N uptake (elevated minus ambient, $E - A$) and E_a is the NUE observed in the ambient $CO₂$ treatment. Similarly, the hypothetical response driven by increased NUE (ΔNPP_e) was calculated as:

$$
\Delta \text{NPP}_e = \Delta E \times U_a, \qquad \text{Eqn 4}
$$

where ΔE is the effect of elevated CO₂ on NUE (E – A) and U_a is the N uptake observed in the ambient $CO₂$ treatment.

We analyzed NPP, %N concentration in oak stems and soil nutrient availability as a function of time since disturbance. We calculated the time since disturbance as the number of years elapsed between the date of the measurement and the most recent disturbance, whether by fire at the beginning of the experiment or by hurricane in September 2004. We expressed the effect of elevated CO₂ as the absolute difference between elevated and ambient $CO₂$ plots.

We used a combination of analysis of variance (ANOVA) and resampling to draw inferences about the effects of elevated CO₂. We used a repeated-measures ANOVA design to analyze time series. We accepted an α value of 0.1. We also used resampling to estimate the magnitude and confidence limits for the observed effects of $CO₂$, using resampling with replacement, $n=8$ and 1000 resamples of the mean, difference between means, relative effect sizes or slope of the relationship between effect sizes and time since disturbance. We estimated 5% and 95% confidence limits as the 5th and 95th percentiles in the resampled population.

Results

Net primary productivity

Elevated CO₂ increased NPP (repeated-measures ANOVA, Tables 2, 3, Fig. 1), but the effect was pronounced in some years and absent in others. The $CO₂$ effect on total NPP was most apparent in the first (1996), second (1997) and tenth (2005) years of treatment. In these years, the 90% confidence intervals (CIs) for the effect of $CO₂$ on NPP were positive and did not include zero (Table 3). Excluding these years with unusually strong responses (+142% for 1997 and +135% for 2005), the average effect of elevated $CO₂$ on NPP was 5%. Elevated $CO₂$ increased aboveground and belowground NPP (Tables 2, 3,

Table 2 P values from repeated-measures analysis of variance of aboveground, belowground and total net primary productivity (NPP; $g \text{ C m}^{-2} \text{ yr}^{-1}$

NPP, carbon	CO ₂	Year	Year \times CO ₂
Aboveground	0.006	0.018	0.879
Quercus myrtifolia	0.078	0.399	0.922
Q. geminata	0.959	0.014	0.516
Q. chapmanii	0.036	0.034	0.623
Leaves	0.014	< 0.001	0.091
Q. myrtifolia	0.125	< 0.001	0.053
Q. geminata	0.964	< 0.001	0.764
Q. chapmanii	0.046	0.003	0.334
Litter production	0.017	< 0.001	0.079
Q. myrtifolia	0.149	< 0.001	0.073
Q. geminata	0.916	< 0.001	0.437
Q. chapmanii	0.048	< 0.001	0.054
Increment	0.012	< 0.001	0.354
Q. myrtifolia	0.091	< 0.001	0.473
Q. geminata	0.905	< 0.001	0.929
Q. chapmanii	0.147	0.002	0.275
Stem increment	0.001	0.001	0.408
Q. myrtifolia	0.033	< 0.001	0.997
Q. geminata	0.813	0.208	0.387
Q. chapmanii	0.055	0.030	0.500
Belowground	0.022	< 0.001	0.600
Coarse root	0.014	0.079	0.999
Litter production	0.056	< 0.001	< 0.001
Increment	0.010	0.122	0.999
Fine roots	0.375	< 0.001	0.614
Litter production	0.216	< 0.001	0.076
Increment	0.625	< 0.001	0.677
Total	0.005	< 0.001	0.461

Bold indicates significant effects ($P < 0.05$).

New
Phytologist Phytologist Research 771

Table 3 Effect of elevated $CO₂$ on net primary productivity (NPP): total (NPP), aboveground (ANPP) and belowground (BNPP) (elevated minus ambient, $E - A$, g C m $^{-2}$ yr $^{-1}$, with 5% and 95% confidence limits^a)

a Bold indicates confidence intervals that do not overlap zero.

Fig. 1 (a) Total net primary productivity (NPP), (b) ratio of below- to aboveground NPP, (c) NPP aboveground and (d) NPP belowground during 11 yr of $CO₂$ enrichment of a subtropical oak woodland. Symbols are means \pm 2 SE (n = 8). Open circles, ambient; closed circles, elevated.

Fig. 1). As with total NPP, positive effects of elevated $CO₂$ were more pronounced (i.e. 90% confidence limits not overlapping zero) early in the experiment and after the hurricane disturbance (Table 3).

Plants allocated more NPP below ground than above ground (Fig. 1, Supporting Information Table S1). Across all sites and times, the ratio of belowground to aboveground NPP was 5.5 ± 0.6 (mean \pm standard error). The relative investment of NPP to structures responsible for resource acquisition was higher below ground than above ground, with 72% of belowground productivity occurring in fine roots compared with 45% of aboveground productivity occurring in leaves. The responses of fine roots to $CO₂$ varied over time, with increases (e.g. 1997– 1998, 2005) followed by sharp declines (1999–2000, 2006, Table S1). Compared with belowground NPP, aboveground NPP was less dynamic (Fig. 1).

The three oak species responded differently to $CO₂$ exposure. Relative to *Q. geminata*, the aboveground production of Q. myrtifolia and Q. chapmanii was more responsive to elevated

 $CO₂$, with many (although not all) components of aboveground NPP exhibiting significant positive responses (Tables 2, 3). By contrast, no productivity component of Q. geminata responded significantly to elevated $CO₂$ (Table 2), nor were any interactions between $CO₂$ and year significant for this species. There was no evidence that the positive growth responses of Q. myrtifolia and Q. chapmanii reduced the growth of Q. geminata through competition: increased growth of the two responsive species was not associated with any reduced growth in Q. geminata.

Nitrogen

Elevated $CO₂$ increased total plant N uptake during some years, in concert with the effects of elevated $CO₂$ on NPP (Tables 4, S2). $CO₂$ increased N uptake strongly early in the experiment (year 2) and after the 2004 hurricane (Fig. 2). $CO₂$ increased total aboveground N uptake, especially by the subdominant Q. chapmanii (Tables 4, S2). N uptake by Q. myrtifolia did not increase significantly in response to elevated $CO₂$, but tended to

Table 4 P values from repeated-measures analysis of variance of nitrogen uptake and translocation

N retranslocation	CO ₂	Year	Year \times CO ₂
Aboveground	0.040	< 0.001	0.085
Quercus myrtifolia	0.420	< 0.001	0.180
Q. geminata	0.892	< 0.001	0.646
Q. chapmanii	0.126	< 0.001	0.102
N uptake			
Aboveground	0.019	< 0.001	0.259
Q. myrtifolia	0.110	< 0.001	0.128
Q. geminata	0.810	0.013	0.439
Q. chapmanii	0.035	0.066	0.680
Belowground	0.486	< 0.001	0.520
Coarse root	0.612	0.306	0.137
Litter production	0.947	< 0.001	0.303
increment	0.519	0.415	0.141
Fine roots	0.531	< 0.001	0.587
Litter production	0.442	< 0.001	0.023
Increment	0.887	< 0.001	0.668
Total	0.079	< 0.001	0.550

Bold indicates significant effects ($P < 0.05$).

Fig. 2 (a) Nitrogen (N) retranslocation and (b) N uptake during 11 yr of CO₂ enrichment of a subtropical oak woodland. Symbols are means \pm 2 SE (n = 8). Open circles, ambient; closed circles, elevated.

be higher and, combined with the significant increase shown by Q. chapmanii, contributed to the significant increase in total aboveground N uptake by all the oaks combined. Total belowground N uptake was insensitive to elevated CO_2 ; the only component of belowground N uptake to respond significantly was fine root litter production, for which responses varied over time (Tables 4, S2).

Elevated $CO₂$ increased total N retranslocation during leaf senescence (Table 4, Fig. 2), but did not consistently increase the

proportion of N retranslocated, which averaged c. 30% in both treatments (data not shown). Thus, the effect of elevated $CO₂$ on N translocation was not a function of increased retranslocation efficiency, but rather reflects the $CO₂$ stimulation of leaf production and thus a greater pool of leaf N from which reabsorption could occur.

Elevated CO_2 caused a more rapid decline in the ^{15}N content of plants (Fig. 3), indicating more rapid N turnover. Part of this effect was driven by increased dilution of $15N$ in plant tissues $(P< 0.05)$, indicating that plants in the elevated $CO₂$ treatment accessed N not available to plants in the ambient $CO₂$ treatment.

Elevated $CO₂$ increased N productivity (Fig. 4); the 90% CI for the difference between treatments $(E - A)$ was positive and did not overlap zero for 1996, 1997, 2004 and 2005. Across all years, N productivity was 12.6 g biomass $yr^{-1} g^{-1} N$ higher in the elevated relative to the ambient $CO₂$ treatment (90% CI, -1.3 to 35.6 g biomass yr⁻¹ g⁻¹ N). Elevated CO₂ had no effect on N residence time in plant tissues (Fig. 4), with a mean difference between the elevated and ambient treatments across all years of -0.4 yr (90% CI, -2.4 to 2.3 yr). Elevated CO₂ increased NUE, the product of N productivity and N residence time (Fig. 4). Averaged across all years, NUE was 164 ± 8 g biomass g^{-1} N for the ambient CO₂ treatment and 188 \pm 11 g biomass g^{-1} N for the elevated CO₂ treatment ($P = 0.054$).

Increased NUE played a minor role in explaining the N economy underpinning positive NPP responses to elevated CO₂. The effect of elevated $CO₂$ on N uptake explained nearly all of the observed large stimulation of NPP found in the years 1997 and 2005 (Fig. 5). Thus, acquisition of N from the environment was associated with (and perhaps necessary for) the strong responses of NPP to elevated $CO₂$. In 2005, the year with the strongest NPP response after application of the $15N$ tracer, the dilution of $15N$ in plant tissues was less apparent relative to other years (Fig. 3). Thus, the large CO_2 stimulation of NPP and N uptake for this year involved increased uptake of N from the pools of soil N labeled by the tracer, which are probably the rapidly cycling pools of soil organic matter.

Fig. 3 15 N content in plant biomass over time (mg 15 N m⁻²). Symbols are means \pm 2 SE (n = 8). Open circles, ambient; closed circles, elevated.

Fig. 4 Effect of the $CO₂$ treatment on components of nitrogen use efficiency. (a) The absolute effect of elevated $CO₂$ on nitrogen productivity per unit N in plant tissue and (b) the absolute effect of elevated $CO₂$ on N residence time, which is plant N content divided by N uptake. Symbols for (a) and (b) show the mean effect size, elevated – ambient, and bars show the 90% confidence intervals (CIs) determined by resampling. (c) Net primary productivity (NPP) as a function of N uptake. The slopes of the relationships are nitrogen use efficiency (g biomass g^{-1} N) for the ambient (solid) and elevated (dashed) treatments. Open circles, ambient; closed circles, elevated.

Disturbance

The strongest effects of elevated CO₂ on NPP occurred following disturbance, whether by fire at the initiation of the experiment, or by hurricane after 8 yr of exposure to the $CO₂$ treatment (Fig. 6). Within the time scale of our annual resolution of NPP, responses of NPP to elevated $CO₂$ exhibited a similar time constant across the two disturbance events, with the maximum response 1–2 yr after the disturbance (Fig. 6). After these peak

Fig. 5 Observed effect of elevated $CO₂$ on net primary productivity (NPP; diamonds, observed), and effects explained by altered N uptake (squares, N uptake only) or by altered nitrogen use efficiency (circles, NUE only). Asterisks indicate years in which elevated $CO₂$ altered significantly aboveground, belowground or total NPP.

Fig. 6 Temporal pattern of the effect of elevated $CO₂$ on total net primary productivity (NPP) (a) and %N in stems (b) as a function of time since disturbance, highlighting the roles of the initial fire disturbance before the experiment was established, and disturbance by hurricane in September 2004, 7.4 yr later. Responses are plotted as absolute effect sizes, elevated $CO₂ treatment – ambient CO₂ treatment. Asterisks indicate years in which$ the effect of $CO₂$ on NPP was statistically significant. Closed circles, fire; open circles, hurricane.

responses, the effect of elevated CO₂ on NPP declined over time at a rate of $77 \text{ g C m}^{-2} \text{ yr}^{-1}$ (bootstrapped 90% CI, 1– $148 \text{ g C m}^{-2} \text{ yr}^{-1}$). Nutrient availability also peaked and then declined following disturbance: total extractable soil N was high after fire disturbance and subsequently declined, and ortho-P captured on resin lysimeters exhibited a similar pattern (Fig. 7;

although this was not the case for inorganic N captured on resin lysimeters; Johnson et al., 2003). Soil extractable micronutrients, including molybdenum, vanadium and iron, were high in 2005, the year following hurricane disturbance, and declined in 2006 and 2007 (B. A. Hungate, unpublished). There was some evidence that the pulse of nutrient availability post-disturbance influenced the response of the oaks to elevated CO_2 : the effect of elevated $CO₂$ on N concentration in oak stems became increasingly negative over time after disturbance (Fig. 6), although this relationship was not as clear for leaves (data not shown). The slope of the effect of elevated $CO₂$ on stem %N vs time was negative $(-0.010 \text{ %N yr}^{-1}$; CI, -0.005 to $-0.014 \text{ %N yr}^{-1}$), and the intercept was not significantly different from zero (CI, -0.025 to 0.018 %N). Thus, in the first several years after disturbance, elevated $CO₂$ had little effect on the N concentration of oak stems, but, with increasing time since disturbance, the reduction in N concentration in oak stems became more pronounced.

Discussion

In this experiment, elevated CO₂ (+350 μ l l $^{-1}$) caused an average stimulation of NPP of 24%, comparable with that observed in other experiments on trees (Norby et al., 2005). Yet, this average response was far less striking than its year-to-year variation, where negligible responses to the elevated $CO₂$ treatment in most years (averaging 5%) were punctuated by a few years with very strong $CO₂$ enhancements of NPP ($>$ 100%). Such extreme interannual variation in the response of NPP to elevated $CO₂$ is unusual: most experiments show less variable responses (Dukes et al., 2005; Norby et al., 2005; Reich et al., 2006b). However, episodic responses have been observed previously. For example, a strong $CO₂$ stimulation of productivity by an invasive annual grass and the dominant shrubs in a desert ecosystem were associated with high rainfall years (Smith et al., 2000; Housman et al., 2006), whereas overall growth responses in this ecosystem were absent (Newingham *et al.*, 2013). In the scrub-oak experiment described here, years with high precipitation also promoted a stronger aboveground growth response to elevated $CO₂$, particularly in the more responsive species, Q. myrtifolia and Q. chapmanii (Seiler et al., 2009). Thus, climate partially contributed to the temporal variability in the $CO₂$ response observed here.

Although climate played a role, the episodic responses of NPP to elevated CO₂ were more strongly associated with disturbance (Fig. 6). Disturbance can cause resource pulses, including pulses of soil nutrients (Matson & Vitousek, 1981), and disturbance is a strong selective agent, often favoring plants with high allocation below ground and plant strategies enabling recovery, such as the ability to resprout (Clarke et al., 2013). The observed pattern of NPP responses to elevated $CO₂$ following disturbance may be explained by the plant strategies that have evolved in response to episodic disturbance regimes and by the effects of disturbance on nutrient availability.

Resprouting occurred immediately after fire (Dijkstra et al., 2002) and refoliation immediately after hurricane (Li et al., 2007); yet, the maximum response of NPP to elevated $CO₂$ did not occur immediately, but rather 18–24 months after disturbance (Fig. 6). This was also apparent when analyzed at a finer temporal resolution. For example, root growth responses to elevated $CO₂$ for the first 7 months after fire were not as large as those observed during the first 24 months (Dilustro et al., 2002). Similarly, the effects of $CO₂$ on leaf area and net ecosystem C assimilation, which were measured independently from the components of NPP reported here, were present, but not unusually large during the 4 months immediately following the hurricane (Li et al., 2007). The effect of elevated $CO₂$ was most strongly manifested after the plants had begun to re-establish the capacity for C gain through canopy regrowth, and the effect was most pronounced as increased fine root growth.

Belowground NPP was five times greater than NPP above ground in this subtropical woodland, and therefore belowground NPP was the major driver of the year-to-year variation in total NPP and of the $CO₂$ response. This may reflect strong relative limitation of productivity by belowground resources, which is probably pronounced in the sandy, low-nutrient soils of this system. High belowground allocation may also occur because of plant strategies favoring resource allocation to organs protected from fire (Clarke et al., 2013). Increased allocation to roots following aboveground disturbance has been observed (Di Iorio et al., 2011) and may be adaptive in scavenging and retaining soil nutrients (Langley et al., 2002), especially if retranslocatable C stored in belowground plant tissues is sufficient to alleviate C limitation after defoliation. Elevated $CO₂$ enhanced instantaneous net ecosystem C uptake in this ecosystem (Hymus et al.,

Fig. 7 Soil extractable nitrogen (a) and phosphorus (b) as a function of time since disturbance. Open squares, ambient; closed squares, elevated CO₂. Plots are means \pm 2 SE.

The transient effect of disturbance on nutrient availability also probably contributed to the strong response of NPP to elevated $CO₂$, as well as the subsequent decline in the response. Hurricanes and fires cause organic matter inputs to soil through leaf fall (hurricane) and ash from combustion of leaves, stems and surface litter. At our site, NMR analysis of the soil in a chronosequence after fire suggests that substantial organic matter inputs from fine root death also occur after fire (Alexis et al., 2012). Rapid mineralization of these organic matter sources probably contributes to the pulse of nutrient availability observed (Fig. 7), and the corresponding NPP response to elevated $CO₂$. The peak in the $CO₂$ stimulation of productivity was coincident with the high availability of multiple soil nutrients, including N, P and micronutrients (Fig. 7; B. A. Hungate, unpublished), and the response of NPP to elevated $CO₂$ declined, together with the availability of these nutrients in soil. The temporal pattern of the CO_2 -mediated reduction in stem N concentration is also consistent with increasing N limitation of the $CO₂$ response (Fig. 6). Together, these results indicate that nutrient limitation can cause a progressive restriction in the response of NPP to elevated $CO₂$ (Field, 1999; Luo et al., 2004; Reich et al., 2006a,b; Norby et al., 2010), and that interactions between disturbance and nutrient availability can strongly affect productivity responses to elevated $CO₂$. This interpretation is also complementary to the notion described earlier that fine root regrowth following disturbance is adaptive for capturing the pulse of soil nutrients (Langley et al., 2002), and is consistent with past observations that growth responses to elevated $CO₂$ are more pronounced when nutrient availability is high (de Graaff et al., 2006; Reich & Hobbie, 2012).

The maximum responses of NPP to elevated $CO₂$ reported here (>100%) are larger than those observed in most other experiments, even in experiments with fertilizer additions (Reich et al., 2006b). Because of the dominance of belowground production to total NPP, and the importance of fine root production to the belowground total, our conclusions rely on scaling minirhizotron observations to biomass and calculating biomass increments to estimate productivity, a challenge in any ecosystem, and an approach that usually yields productivity estimates higher than those obtained from sequential coring or other approaches (Strand et al., 2008). However, the scaling problem applies equally to both ambient and elevated $CO₂$ treatments, and so is unlikely to explain the large responses to elevated $CO₂$ observed here. Remobilization of stored C after disturbance could also skew productivity estimates. We have demonstrated previously that stored C is used to support new root growth in this ecosystem (Langley et al., 2002), creating a multi-year time lag between photosynthesis and root growth. The use of stored C to produce new fine roots will cause an overestimate of belowground productivity if replenishment of stored reserves in belowground storage organs does not occur simultaneously to remobilization, or if the decrement in coarse root C caused by mobilization of stored reserves is difficult to detect. Over multiple years, the influence of this error, if it occurs, will diminish, as what is mistakenly counted as fine root NPP in 1 yr will

be missed in coarse root increment in subsequent years. This phenomenon could dampen the temporal pattern observed after disturbance (Fig. 6), but is unlikely to change it qualitatively. Thus, the large responses of NPP to elevated $CO₂$ immediately following disturbance probably reflect real properties of the system rather than measurement challenges.

Increased N uptake (Finzi et al., 2007) and increased NUE (Calfapietra et al., 2007; Norby et al., 2010) contribute to the response of NPP to elevated $CO₂$, consistent with the findings reported here that elevated CO₂ increased both NUE and N uptake (Fig. 4). Our finding that increased N uptake was the more important mechanism, explaining > 90% of the response in the years immediately post-disturbance (Fig. 5), underscores the facilitating role of N availability (and of other nutrients) for positive NPP responses to elevated $CO₂$ (Reich et al., 2006a,b), and is consistent with the finding that nutrient availability increased after disturbance (Fig. 7).

Increased total N uptake and faster turnover of tracer ¹⁵N through plants indicate that elevated $CO₂$ increased the rates of N cycling through the plant–soil system, enhancing the availability of unlabeled N via priming (Carney et al., 2007; Langley et al., 2009) or access to N deep in the soil profile (McKinley et al., 2009), or both. Both sources could contribute to the increased N uptake and observed $15N$ dilution. Yet, the implications of these sources differ. The priming mechanism would involve a net redistribution of N from soil organic matter to plants, from a reservoir with a narrow C to N ratio to one with a wider C to N ratio. This N could become more susceptible to loss from the system through leaching and gaseous pathways, as found for scrub-oak, in which total system $15N$ recovery was lower in the high $CO₂$ treatment (Hungate et al., 2013). Although there is the potential for priming and redistribution of N to enhance C uptake and storage (Drake et al., 2011), this outcome is far from certain. The other mechanism, increased N uptake from the water table, raises issues of experimental design and scale. N inputs from groundwater originate in part from N originally contained in the vertical column defined by the experimental chambers, but also from N flowing laterally from adjacent areas via the water table. In other words, the water table provides hydrological connectivity between the chambers and the surrounding landscape, an avenue for nutrient subsidies to the experimental plots. Such subsidies do not represent viable sources of new nutrients when atmospheric $CO₂$ concentration is increasing globally, but rather represent an artifact of the spatial scale of the design in which plants can access nutrient subsidies from areas outside the experiment. At this point, we cannot quantify the relative importance of these mechanisms, but we note that there is experimental evidence for both (Carney et al., 2007; Langley et al., 2009; McKinley et al., 2009). To the extent that the latter operates, our results overestimate the $CO₂$ effect on N uptake and availability.

Conclusions

The subtropical woodland studied here has a short mean fire return interval, and thus the responses shown here may not be apparent in experiments of comparable duration in ecosystems

with longer times between disturbance events, or with plants that lack certain adaptive strategies, such as the ability to resprout using C stored in belowground tissues. Nevertheless, given the widespread occurrence and influence of episodic disturbance on terrestrial ecosystems modulated by changes in resource availability, the phenomena observed here may illustrate general interactions between disturbance and ecosystem responses to the changing atmosphere.

Our work shows that disturbance modulates ecosystem responses to chronic global environmental change. Disturbance can reset the biogeochemical trajectories and trigger plant allocation strategies that shape responses of NPP to elevated $CO₂$. Transient pulses of nutrient availability, such as those caused by disturbance, may be especially important in explaining the strong NPP responses in some years to elevated $CO₂$. Our results caution against the interpretation of short-term responses as representative, because they may dissipate over time. Disturbance can influence ecosystem responses to global environmental change, and there is value in incorporating disturbances and other extreme events into the design of global change experiments.

Acknowledgements

The $CO₂$ project was supported by the US Department of Energy (DE-FG-02-95ER61993 and subcontract 95-59, MPOOO02) and by the National Science Foundation (DEB 0092642 and 0445324). The National Aeronautics and Space Administration at the Kennedy Space Center and the US Fish and Wildlife Service at Merritt Island National Wildlife Refuge provided generous support throughout the $CO₂$ project. The authors thank David Johnson, Graham Hymus, Jiahong Li, Debra Colavito, Hans Anderson, Dan Stover, Alisha Brown, Rachel Schroeder, John Butnor, John Dilustro and Troy Seiler for fieldwork.

References

- Alexis MA, Rasse DP, Knicker H, Anquetil C, Rumpel C. 2012. Evolution of soil organic matter after prescribed fire: a 20-year chronosequence. Geoderma 189–190: 98–107.
- Block CE, Knoepp JD, Fraterrigo JM. 2013. Interactive effects of disturbance and nitrogen availability on phosphorus dynamics of southern Appalachian forests. Biogeochemistry 112: 329–342.
- Brown ALP, Day FP, Hungate BA, Drake BG, Hinkle CR. 2007. Root biomass and nutrient dynamics in a scrub-oak ecosystem under the influence of elevated atmospheric $CO₂$. Plant and Soil 292: 219-232.
- Brown ALP, Day FP, Stover DB. 2009. Fine root biomass estimates from minirhizotron imagery in a shrub ecosystem exposed to elevated $CO₂$. Plant and Soil 317: 145–153.
- Brown JR, Blankinship JC, Niboyet A, van Groenigen KJ, Dijkstra P, Le Roux X, Leadley PW, Hungate BA. 2012. Effects of multiple global change treatments on soil N₂O fluxes. Biogeochemistry 109: 85-100.
- Calfapietra C, De Angelis P, Gielen B, Lukac M, Moscatelli MC, Avino G, Lagomarsino A, Polle A, Ceulemans R, Mugnozza GS et al. 2007. Increased nitrogen-use efficiency of a short-rotation poplar plantation in elevated $CO₂$ concentration. Tree Physiology 27: 1153–1163.
- Carney KM, Hungate BA, Drake BG, Megonigal JP. 2007. Altered soil microbial community at elevated $CO₂$ leads to loss of soil carbon. Proceedings of the National Academy of Sciences, USA 104: 4990–4995.
- Clarke PJ, Lawes MJ, Midgley JJ, Lamont BB, Ojeda F, Burrows GE, Enright NJ, Knox KJE. 2013. Resprouting as a key functional trait: how buds, protection and resources drive persistence after fire. New Phytologist 197: 19–35.
- Day FP, Schroeder RE, Stover DB, Brown ALP, Butnor JR, Dilustro J, Hungate BA, Dijkstra P, Duval BD, Seiler TJ et al. 2013. The effects of 11 yr of CO₂ enrichment on roots in a Florida scrub-oak ecosystem. New Phytologist. doi: 10.1111/nph.12246.
- Day FP, Stover DB, Pagel AL, Hungate BA, Dilustro JJ, Herbert BT, Drake BG, Hinkle CR. 2006. Rapid root closure after fire limits fine root responses to elevated atmospheric CO₂ in a scrub oak ecosystem in central Florida, USA. Global Change Biology 12: 1047–1053.
- Di Iorio A, Montagnoli A, Scippa GS, Chiatante D. 2011. Fine root growth of Quercus pubescens seedlings after drought stress and fire disturbance. Environmental and Experimental Botany 74: 272–279.
- Dijkstra P, Hymus G, Colavito D, Vieglais DA, Cundari CM, Johnson DP, Hungate BA, Hinkle CR, Drake BG. 2002. Elevated atmospheric CO₂ stimulates aboveground biomass in a fire-regenerated scrub-oak ecosystem. Global Change Biology 8: 90–103.
- Dilustro JJ, Day FP, Drake BG, Hinkle CR. 2002. Abundance, production and mortality of fine roots under elevated atmospheric $CO₂$ in an oak-scrub ecosystem. Environmental and Experimental Botany 48: 149–159.
- Drake JE, Gallet-Budynek A, Hofmockel KS, Bernhardt ES, Billings SA, Jackson RB, Johnsen KS, Lichter J, McCarthy HR, McCormack ML et al. 2011. Increases in the flux of carbon belowground stimulate nitrogen uptake and sustain the long-term enhancement of forest productivity under elevated CO₂. Ecology Letters 14: 349-357.
- Dukes JS, Chiariello NR, Cleland EE, Moore LA, Shaw MR, Thayer S, Tobeck T, Mooney HA, Field CB. 2005. Responses of grassland production to single and multiple global environmental changes. PLoS Biology 3: 1829–1837.
- Field CB. 1999. Diverse controls on carbon storage under elevated CO₂: toward a synthesis. In: Luo YM, Mooney HA, eds. Carbon dioxide and environmental stress. San Diego, CA, USA: Academic Press, 373–391.
- Field CB, Chapin FS, Matson PA, Mooney HA. 1992. Responses of terrestrial ecosystems to the changing atmosphere - a resource-based approach. Annual Review of Ecology and Systematics 23: 201–235.
- Finzi AC, Norby RJ, Calfapietra C, Gallet-Budynek A, Gielen B, Holmes WE, Hoosbeek MR, Iversen CM, Jackson RB, Kubiske ME et al. 2007. Increases in nitrogen uptake rather than nitrogen-use efficiency support higher rates of temperate forest productivity under elevated CO₂. Proceedings of the National Academy of Sciences, USA 104: 14014-14019.
- de Graaff M-A, van Groenigen K-J, Six J, Hungate B, van Kessel C. 2006. Interactions between plant growth and soil nutrient cycling under elevated CO2: a meta-analysis. Global Change Biology 12: 2077–2091.
- Guerin DN. 1993. Oak dome clonal structure and fire ecology in a Florida longleaf pine dominated community. Bulletin of the Torrey Botanical Club 120: 107–114.
- Hagedorn F, Maurer S, Bucher JB, Siegwolf RTW. 2005. Immobilization, stabilization and remobilization of nitrogen in forest soils at elevated CO₂: a ¹⁵N and ¹³C tracer study. *Global Change Biology* 11: 1816–1827.
- Henry HAL, Chiariello NR, Vitousek PM, Mooney HA, Field CB. 2006. Interactive effects of fire, elevated carbon dioxide, nitrogen deposition, and precipitation on a California annual grassland. Ecosystems 9: 1066-1075.
- Housman DC, Naumburg E, Huxman TE, Charlet TN, Nowak RS, Smith SD. 2006. Increases in desert shrub productivity under elevated carbon dioxide vary with water availability. Ecosystems 9: 374-385.
- Hungate BA, Dijkstra P, Wu Z, Duval BD, Day FP, Johnson DW, Megonigal JP, Brown ALP, Garland JL. 2013. Cumulative response of ecosystem carbon and nitrogen stocks to chronic $CO₂$ exposure in a subtropical oak woodland. New Phytologist. doi: 10.1111/nph.12333.
- Hungate BA, Johnson DW, Dijkstra P, Hymus G, Stiling P, Megonigal JP, Pagel AL, Moan JL, Day F, Li JH et al. 2006. Nitrogen cycling during seven years of atmospheric CO₂ enrichment in a scrub oak woodland. Ecology 87: 26–40.
- Hymus GJ, Johnson DP, Dore S, Anderson HP, Hinkle CR, Drake BG. 2003. Effects of elevated atmospheric $CO₂$ on net ecosystem $CO₂$ exchange of a scrub-oak ecosystem. Global Change Biology 9: 1802-1812.
- Phytologist Research 777
- Johnson DW, Hoylman AM, Ball JT, Walker RF. 2006. Ponderosa pine responses to elevated $CO₂$ and nitrogen fertilization. Biogeochemistry 77: 157– 175.
- Johnson DW, Hungate BA, Dijkstra P, Hymus G, Hinkle CR, Stiling P, Drake BG. 2003. The effects of elevated $CO₂$ on nutrient distribution in a fire-adapted scrub oak forest. Ecological Applications 13: 1388–1399.
- Körner C. 2006. Plant CO₂ responses: an issue of definition, time and resource supply. New Phytologist 172: 393-411.
- Langley JA, Dijkstra P, Drake BG, Hungate BA. 2003. Ectomycorrhizal colonization, biomass, and production in a regenerating scrub oak forest in response to elevated CO₂. Ecosystems 6: 424-430.
- Langley JA, Drake B, Hungate BA. 2002. Extensive belowground carbon storage supports roots and mycorrhizae in regenerating scrub oaks. Oecologia 131: 542-548.
- Langley JA, McKinley DC, Wolf AA, Hungate BA, Drake BG, Megonigal JP. 2009. Priming depletes soil carbon and releases nitrogen in a scrub-oak ecosystem exposed to elevated CO₂. Soil Biology and Biochemistry 41: 54-60.
- Leuzinger S, Luo Y, Beier C, Dieleman W, Vicca S, Koerner C. 2011. Do global change experiments overestimate impacts on terrestrial ecosystems? Trends in Ecology & Evolution 26: 236–241.
- Li J, Powell TL, Seiler TJ, Johnson DP, Anderson HP, Bracho R, Hungate BA, Hinkle CR, Drake BG. 2007. Impacts of Hurricane Frances on Florida scrub-oak ecosystem processes: defoliation, net $CO₂$ exchange and interactions with elevated CO₂. Global Change Biology 13: 1101-1113.
- Luo Y, Su B, Currie WS, Dukes JS, Finzi AC, Hartwig U, Hungate B, McMurtrie RE, Oren R, Parton WJ et al. 2004. Progressive nitrogen limitation of ecosystem responses to rising atmospheric carbon dioxide. BioScience 54: 731–739.
- Mailander JL. 1990. Climate of the Kennedy Space Center and Vicinity US Aeronautics and Space Administration Technical Memorandum 103498.
- Martin A, Diaz-Ravina M, Carballas T. 2012. Short- and medium-term evolution of soil properties in Atlantic forest ecosystems affected by wildfires. Land Degradation & Development 23: 427-439.
- Matson PA, Vitousek PM. 1981. Nitrogen mineralization and nitrification potentials following clearcutting in the Hoosier-National Forest, Indiana. Forest Science 27: 781–791.
- McKinley DC, Romero JC, Hungate BA, Drake BG, Megonigal JP. 2009. Does deep soil N availability sustain long-term ecosystem responses to elevated CO₂? Global Change Biology 15: 2035–2048.
- Michalzik B, Martin S. 2013. Effects of experimental duff fires on C, N and P fluxes into the mineral soil at a coniferous and broadleaf forest site. Geoderma 197: 169–176.
- Newingham BA, Vanier C, Charlet T, Ogle K, Smith SD, Nowak RS. 2013. No cumulative effect of ten years of elevated $CO₂$ on perennial plant biomass components in the Mojave Desert. Global Change Biology 19: 2168-2181.
- Niboyet A, Brown JR, Dijkstra P, Blankinship JC, Leadley PW, Le Roux X, Barthes L, Barnard RL, Field CB, Hungate BA. 2011. Global change could amplify fire effects on soil greenhouse gas emissions. PLoS ONE 6: e20105.
- Niklaus PA, Körner C. 2004. Synthesis of 6 year $CO₂$ enrichment study: interactions between nutrient and hydrological cycles and soil processes. Ecological Society of America Annual Meeting Abstracts 89: 373–374.
- Norby RJ, Delucia EH, Gielen B, Calfapietra C, Giardina CP, King JS, Ledford J, McCarthy HR, Moore DJ, Ceulemans R et al. 2005. Forest response to elevated CO₂ is conserved across a broad range of productivity. Proceedings of the National Academy of Sciences, USA 102: 18052–18056.
- Norby RJ, Warren JM, Iversen CM, Medlyn BE, McMurtrie RE. 2010. CO₂ enhancement of forest productivity constrained by limited nitrogen availability. Proceedings of the National Academy of Sciences, USA 107: 19368–19373.
- Owensby CE, Ham JM, Knapp AK, Auen LM. 1999. Biomass production and species composition change in a tallgrass prairie ecosystem after long-term exposure to elevated atmospheric CO₂. Global Change Biology 5: 497-506.
- Rasse DP, Peresta G, Drake BG. 2005. Seventeen years of elevated CO₂ exposure in a Chesapeake Bay Wetland: sustained but contrasting responses of plant growth and CO₂ uptake. Global Change Biology 11: 369-377.
- Reich PB, Hobbie SE, Lee T, Ellsworth DS, West JB, Tilman D, Knops JMH, Naeem S, Trost J. 2006a. Nitrogen limitation constrains sustainability of ecosystem response to CO₂. Nature 440: 922-925.
- Reich PB, Hungate BA, Luo Y. 2006b. Carbon–nitrogen interactions in terrestrial ecosystems in response to rising atmospheric carbon dioxide. Annual Review of Ecology, Evolution, and Systematics 37: 611–636.
- Reich PBH, Hobbie SE. 2012. Decade-long soil nitrogen constraint on the CO₂ fertilization of plant biomass. Nature Climate Change 1: 346–347.
- Schäppi B, Körner C. 1996. Growth responses of an alpine grassland to elevated CO₂. Oecologia 105: 43-52.
- Schmalzer PA, Hinkle CR. 1992. Recovery of oak-saw palmetto scrub after fire. Castanea 57: 158–173.
- Seiler TJ, Rasse DP, Li J, Dijkstra P, Anderson HP, Johnson DP, Powell TL, Hungate BA, Hinkle CR, Drake BG. 2009. Disturbance, rainfall and contrasting species responses mediated aboveground biomass response to 11 years of CO₂ enrichment in a Florida scrub-oak ecosystem. Global Change Biology 15: 356–367.
- Smith E. 1999. Atlantic and East Coast hurricanes 1900–98: a frequency and intensity study for the twenty-first century. Bulletin of the American Meteorological Society 80: 2717–2720.
- Smith SD, Huxman TE, Zitzer SF, Charlet TN, Housman DC, Coleman JS, Fenstermaker LK, Seemann JR, Nowak RS. 2000. Elevated CO₂ increases productivity and invasive species success in an arid ecosystem. Nature 408: 79–82.
- Sousa WP. 1984. The role of disturbance in natural communities. Annual Review of Ecology and Systematics 15: 353–391.
- Stiling P, Moon D, Rossi A, Hungate BA, Drake B. 2009. Seeing the forest for the trees: long-term exposure to elevated CO₂ increases some herbivore densities. Global Change Biology 15: 1895-1902.
- Stover DB, Day FP, Butnor JR, Drake BG. 2007. Effect of elevated CO₂ on coarse-root biomass in Florida scrub detected by ground-penetrating radar. Ecology 88: 1328–1334.
- Stover DB, Day FP, Drake BG, Hinkle CR. 2010. The long-term effects of CO₂ enrichment on fine root productivity, mortality, and survivorship in a scrub-oak ecosystem at Kennedy Space Center, Florida, USA. Environmental and Experimental Botany 69: 214–222.
- Strand AE, Pritchard SG, McCormack ML, Davis MA, Oren R. 2008. Irreconcilable differences: fine-root life spans and soil carbon persistence. Science 319: 456–458.
- Wang YP, Houlton BZ, Field CB. 2007.A model of biogeochemical cycles of carbon, nitrogen, and phosphorus including symbiotic nitrogen fixation and phosphatase production. Global Biogeochemical Cycles 21, GB1018, doi: 10.1029/2006GB002797.
- Webber HJ. 1935. The Florida scrub, a fire-fighting association. American Journal of Botany 22: 344–361.
- Zak DR, Pregitzer KS, Kubiske ME, Burton AJ. 2011. Forest productivity under elevated $CO₂$ and $O₃$: positive feedbacks to soil N cycling sustain decade-long net primary productivity enhancement by CO₂. Ecology Letters 14: 1220-1226.
- Zanetti S, Hartwig UA. 1997. Symbiotic N_2 fixation increases under elevated atmospheric pCO_2 in the field. Acta Oecologica-International Journal of Ecology 18: 285–290.

Supporting Information

Additional supporting information may be found in the online version of this article.

Table S1 Net primary production 1996–2007 in response to increased atmospheric $CO₂$ concentrations in a subtropical woodland

Table S2 Nitrogen uptake above- and belowground

Please note: Wiley-Blackwell is not responsible for the content or functionality of any supporting information supplied by the authors. Any queries (other than missing material) should be directed to the New Phytologist Central Office.