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An extracellular domain of the accessory β1 subunit is required for modulating BK channel voltage sensor and gate

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A family of tissue-specific auxiliary β subunits modulates large conductance voltage- and calcium-activated potassium (BK) channel gating properties to suit their diverse functions. Paradoxically, β subunits both promote BK channel activation through a stabilization of voltage sensor activation and reduce BK channel openings through an increased energetic barrier of the closed-to-open transition. The molecular determinants underlying β subunit function, including the dual gating effects, remain unknown. In this study, we report the first identification of a β1 functional domain consisting of Y74, S104, Y105, and I106 residues located in the extracellular loop of β1. These amino acids reside within two regions of highest conservation among related β1, β2, and β4 subunits. Analysis in the context of the Horrigan-Aldrich gating model revealed that this domain functions to both promote voltage sensor activation and also reduce intrinsic gating. Free energy calculations suggest that the dual effects of the β1 Y74 and S104–I106 domains can be largely accounted for by a relative destabilization of channels in open states that have few voltage sensors activated. These results suggest a unique and novel mechanism for β subunit modulation of voltage-gated potassium channels wherein interactions between extracellular β subunit residues with the external portions of the gate and voltage sensor regulate channel opening.

INTRODUCTION

BK channels have an established role in regulating vascular smooth muscle tone by hyperpolarizing the membrane and deactivating voltage-dependent Ca\(^{2+}\) channels (Kaczorowski et al., 1996; Gribkoff et al., 1997; Calderone, 2002). Increased opening of smooth muscle BK channels is conferred by the β1 auxiliary subunit (Tanaka et al., 1997; Brenner et al., 2000b; Plüger et al., 2000). The important role of the β1 subunit has been demonstrated in β1 knockout mice, which display reduced BK channel opening, increased vascular tone, and hypertension (Brenner et al., 2000b; Plüger et al., 2000). In addition, two human β1 polymorphisms, each with a single amino acid change in the extracellular domain, have been associated with altered smooth muscle function. A gain-of-function polymorphism has been linked to a reduced incidence of hypertension (Fernández-Fernández et al., 2004). In addition, a polymorphism that moderately reduces channel opening has been associated with increased asthma severity (Seibold et al., 2008).

The BK channel pore-forming α subunit belongs to the six-transmembrane (TM) voltage-dependent K⁺ channel family (Atkinson et al., 1991). The β1 accessory subunit belongs to the two-TM BK channel auxiliary β subunit family consisting of four members, β1–β4, each having distinct tissue-specific expression (Knaus et al., 1994; Riazi et al., 1999; Wallner et al., 1999; Xia et al., 1999; Behrens et al., 2000; Brenner et al., 2000a). The β subunit structure contains a large extracellular loop, two TMs, and two small intracellular domains (Knaus et al., 1994; Orio et al., 2002). Recently, a distinct single TM protein, LRRC26 (leucine-rich repeat–containing protein 26), was also found to act as an accessory BK channel subunit in a prostate cancer cell line (Yan and Aldrich, 2010).

Mutagenesis of the α subunit has yielded a large number of insights into the structure of the pore-forming subunit of the BK channel (Lee and Cui, 2010). In contrast, the β structural determinants that modulate α subunit gating remain unclear. BK channel β subunits are apparently unrelated to other protein families. Therefore, identifying structural domains of β subunits by scanning mutagenesis is encumbered by the potentially large number of mutations that would be required. However, there is evidence that some modulatory effects of β subunit family members β1, β2, and β4 are conserved. These β subunits slow activation and deactivation gating kinetics (Behrens et al., 2000; Brenner et al., 2000a; Lippiat et al., 2003). In addition, these β subunits exert similar Ca\(^{2+}\)-dependent effects on steady-state opening (Behrens et al., 2000; Brenner et al., 2000a; Lippiat et al., 2003). They increase channel opening in high Ca\(^{2+}\), effects that are accounted for by...
β subunit modulation of voltage sensor and Ca\textsuperscript{2+} binding (Bao and Cox, 2005; Orio and Latorre, 2005; Wang and Brenner, 2006; Wang et al., 2006; Sweet and Cox, 2009). β subunits also reduce channel opening in low intracellular Ca\textsuperscript{2+} by a reduction of intrinsic gating (channel opening independent of voltage sensor activation and Ca\textsuperscript{2+} binding; Orio and Latorre, 2005; Wang and Brenner, 2006; Wang et al., 2006). These functional similarities suggest that structural determinants underlying β subunit function may also be conserved (Orio et al., 2006). Therefore, key residues and functional domains may be uncovered by alanine substitution of conserved amino acids. Using this approach, we report here the identification of a novel β1 extracellular domain that is critical for modulation of voltage sensor activation and intrinsic gating of BK channels.

**MATERIALS AND METHODS**

**Channel expression**

The mouse BK α cDNA (GenBank/EMBL/DDBJ accession no. MMU09383) was modified to include the extended amino-terminal sequence (beginning MANG) encoded by the KCNMA1 gene. We found that the extra sequence causes an ~20-mV larger negative G-V shift by mouse β1 as compared with the truncated α subunit (initiating translation at the internal MDAL residues) that has been most often used in the past. The extended aminoterminal sequence was also added to the F315Y construct (Wang and Brenner, 2006), which we call F380Y in this study. With mouse β1 cDNA (Wang and Brenner, 2006) as a template, mutant β1 constructs were generated with a Quick-Change XL Site-Directed Mutagenesis kit (Agilent Technologies) and confirmed by sequencing.

α and various β1 cDNAs were cotransfected into HEK-293 cells (American Type Culture Collection) and studied 1–2 d after transfection. β1 subunits were subcloned in the mammalian expression vector pIRE2-enhanced green fluorescent protein (Takara Bio Inc.), which contains the enhanced green fluorescent protein gene that fluorescently labels transfected cells. A expression vector pIRES2–enhanced green fluorescent protein (Takara Bio Inc.), which contains the enhanced green fluorescent protein gene that fluorescently labels transfected cells. A expression vector pIRES2–enhanced green fluorescent protein (Takara Bio Inc.), which contains the enhanced green fluorescent protein gene that fluorescently labels transfected cells. A expression vector pIRES2–enhanced green fluorescent protein (Takara Bio Inc.), which contains the enhanced green fluorescent protein gene that fluorescently labels transfected cells.

**Electrophysiology and data analysis**

Currents were recorded using the patch clamp technique in the inside-out configuration. The external recording solution contained 20 mM HEPES, 140 mM KMeSO\textsubscript{3}, 2 mM KCl, and 2 mM MgCl\textsubscript{2}, pH 7.2. Internal solutions contained 20 mM HEPES, 140 mM KMeSO\textsubscript{3}, and 2 mM KCl, pH 7.2. For the 60-mM free Ca\textsuperscript{2+} intracellular solution, Ca\textsuperscript{2+} was buffered with 5 mM nitro-tri-acetic acid. For nominally 0 Ca\textsuperscript{2+} (0.002 µM of free Ca\textsuperscript{2+}), intracellular Ca\textsuperscript{2+} was buffered with 2 mM EGTA. Free [Ca\textsuperscript{2+}] was measured using a Ca\textsuperscript{2+}-sensitive electrode (Orion Research).

Open probability (P\textsubscript{o}) was estimated by steady-state macroscopic recordings (when P\textsubscript{o} < 0.05) and single-channel recordings in the same patch. nP\textsubscript{o} was determined from all-points amplitude histograms by the sum of open levels (k) multiplied by fractional time spent (P\textsubscript{o}): nP\textsubscript{o} = ΣkP\textsubscript{o}. To estimate the number of channels in a patch (n), maximum macroscopic conductance (G\textsubscript{Max}) was divided by single-channel conductance (g\textsubscript{k}) at the same voltage for tail current measurements (~80 mV). n = G\textsubscript{Max}/g\textsubscript{k}.

The steady-state data in 0 Ca\textsuperscript{2+} were fit to the Horrigan-Aldrich model (Horrigan and Aldrich, 2002) based on least-squares criteria. The following equations were used to estimate energetic changes associated with mutations (Ma et al., 2006):

\[
C_{0-O} \Delta G = -kT \ln \frac{J_{o}^{M}}{J_{o}^{WT}}
\]

\[
C_{i-C} \Delta G = 4kT \ln \frac{J_{o}^{M}}{J_{o}^{WT}}
\]

\[
J_{0} = \frac{0.5}{\exp \left(l_0 V_{h}/kT\right)}
\]

\[
O_{0}-O \Delta G = -4kT \ln \frac{(J_{o}D)^{M}}{(J_{o}D)^{WT}}
\]

\[
J_{o}D = \frac{0.5}{\exp \left(l_0 V_{h}/kT\right)}
\]

\[
O_{i-C} \Delta G = kT \ln \frac{J_{o}^{M}D^{WT}}{J_{o}^{WT}D^{WT}}
\]

**RESULTS**

Identifying β1 residues critical for gating modulation

Despite different physiological roles, both β1 and neuron-specific β4 subunits slow BK channel gating and modulate steady-state properties (Fig. S1, A and B). Compared with α alone channels, both β1 and β4 increase steady-state P\textsubscript{o} at high Ca\textsuperscript{2+} but reduce it at low Ca\textsuperscript{2+} (Fig. S1 B). These steady-state effects are largely accounted for by two Ca\textsuperscript{2+}-independent mechanisms. These are a reduction in intrinsic gating and a negative shift of open-channel voltage sensor activation (Fig. S1 C; Bao and Cox, 2005; Wang and Brenner, 2006; Wang et al., 2006; Sweet and Cox, 2009). Similarly, data from a previous study suggest that the dual gating mechanisms also underlie β2 modulation of BK channel properties (Orio and Latorre, 2005).

These prior findings led us to hypothesize that residues mediating gating modulation are conserved among
residues reduce $\beta_1$-mediated G-V shifts, increasing $V_{1/2}$ by 28–44 mV (Fig. 3 A). These results are consistent with the hypothesis that segment A and B residues play an important role in gating modulation.

Segment A and B residues have nonadditive effects

Similar effects of alanine substitution of Y74, S104, Y105, and I106 suggest that these residues may contribute to common functional interactions. To test this hypothesis, we examined whether these mutations have additive effects. If the two residues contribute to a common interaction, effects of the double mutation should not exceed both single mutations.

We first examined segment B mutations Y105A and I106A (Fig. 4 A). Indeed, the steady-state effect of the double mutant Y105AI106A ($V_{1/2}$ of −48 ± 4 mV) is not significantly different from individual mutants Y105A ($V_{1/2}$ of −45 ± 4 mV, $P \approx 0.6$) or I106A ($V_{1/2}$ of −49 ± 4 mV, $P \approx 0.9$; Fig. 4 B, left). The nonadditive effects were also observed from activation kinetics. Compared with $\beta_1$WT, both single mutations slow activation, with I106A having a greater effect than Y105A. This kinetic effect of Y105AI106A is not significantly greater than I106A (Fig. 4 B, right). At −20 mV, time constants for I106A and Y105AI106A are 10.1 ± 1.5 ms and 14.2 ± 2.0 ms, respectively ($P \approx 0.1$). At −40 mV, they are 17.0 ± 1.3 ms and 21.4 ± 3.5 ms, respectively ($P \approx 0.3$).

β1, β2, and β4. To identify potential key gating residues and domains, we performed sequence alignment of $\beta_1$, $\beta_2$, $\beta_3$, and $\beta_4$ and identified two highly conserved segments (Fig. 1). Extracellular segment A consists of four identical residues (Q73YPC76), and segment B consists of five identical residues (C103SYIP107; Fig. 1). Segments A and B consist of the longest sequence of consecutive identical residues among these $\beta$ subunits. To test the hypothesis that these conserved segments have important roles in gating modulation, we performed an alanine substitution mutagenesis of 13 identical residues within or neighboring segments A and B.

Mutant $\beta_1$ ($\beta_1\text{MT}$) subunits were expressed at saturating concentrations with wild-type $\alpha$ subunits using transient transfection in HEK-293 cells. BK currents were recorded using the inside-out patch clamp configuration at 60 µM Ca²⁺ (Fig. 2 A). Averaged G-V relations of $\alpha_1\text{MT}$ were compared with $\alpha_1$ alone and $\alpha_1\beta_1\text{WT}$ channels (Fig. 2, B and C). Mutations such as V120A caused a small or no change in channel properties (Fig. 2, B and C). In contrast, mutations such as S104A reduced the size of the G-V shift (by −30 mV; Fig. 2 B) and slowed the activation time constants (Fig. 2 C).

In addition to S104, we have identified three other positions that are important in mediating gating modulation. These include segment B residues Y105 and I106 as well as segment A residue Y74 (Fig. 3). Alanine substitutions of these residues reduce $\beta_1$-mediated G-V shifts, increasing $V_{1/2}$ by 28–44 mV (Fig. 3 A). These results are consistent with the hypothesis that segment A and B residues play an important role in gating modulation.

Segment A and B residues have nonadditive effects

Similar effects of alanine substitution of Y74, S104, Y105, and I106 suggest that these residues may contribute to common functional interactions. To test this hypothesis, we examined whether these mutations have additive effects. If the two residues contribute to a common interaction, effects of the double mutation should not exceed both single mutations.

We first examined segment B mutations Y105A and I106A (Fig. 4 A). Indeed, the steady-state effect of the double mutant Y105AI106A ($V_{1/2}$ of −48 ± 4 mV) is not significantly different from individual mutants Y105A (−45 ± 5 mV, $P = 0.6$) or I106A (−49 ± 5 mV, $P = 0.9$; Fig. 4 B, left). The nonadditive effects were also observed from activation kinetics. Compared with $\beta_1\text{WT}$, both single mutants show faster activation, with I106A having a greater effect than Y105A. This kinetic effect of Y105AI106A is not significantly greater than I106A (Fig. 4 B, right). At −20 mV, time constants for I106A and Y105AI106A are 10.1 ± 1.5 ms and 14.2 ± 2.0 ms, respectively ($P = 0.1$). At −40 mV, they are 17.0 ± 1.3 ms and 21.4 ± 3.5 ms, respectively ($P = 0.3$).
Next, we tested whether mutant effects of segment A and B residues Y74A and Y105A are additive (Fig. 4 C). The steady-state effect of Y105AY74A (V_{1/2} of −48 ± 3 mV) is not significantly different from Y105A (−45 ± 5 mV, P = 0.6) even though it is greater than Y74A (−57 ± 3 mV, P = 0.03; Fig. 4 D, left). Similarly, kinetic effects of the double mutation are not greater than those of Y105A (Fig. 4 D, right). At −20 mV, the activation time constant for Y105AY74A (7.2 ± 0.9 ms) is similar to Y105A (8.8 ± 1.0 ms, P ≈ 0.3) and Y74A (8.3 ± 0.7 ms, P ≈ 0.4). Finally, deactivation time constants are also not significantly different between Y105AY74A and Y105A.

Figure 2. Alanine substitutions of key β1 residues affect the G-V relations and gating kinetics. (A) Families of currents recorded at 60 µM Ca^2+ from BK channels composed of α subunit alone (α), α subunit coassembled wild-type β1 (αβ1WT), or β1 with a single alanine substitution (αβ1S04A). α currents were evoked by 100-ms depolarization in 20-mV steps between −60 and 60 mV. αβ1 currents were evoked by 200-ms depolarization in 20-mV steps between −140 and 60 mV. The x and y scale bars represent 20 ms and 0.5 nA, respectively. (B) Averaged G-V relations of αβ1WT and αβ1WT largely overlap, indicating that the V120A mutation has little effect on steady-state modulation of the α subunit. In contrast, the S104A mutation shifts the G-V relations by ~30 mV, reducing β1 steady-state modulatory effects. α, n = 16; αβ1WT, n = 12; αβ1S04A, n = 32; αβ1V120A, n = 11. (C) S104A, but not V120A, alters β1 effects on activation kinetics. The averaged activation and deactivation time constants of α, αβ1WT, αβ1V120A, and αβ1S04A channels are shown, plotted as a function of voltage. α, n = 9–16; αβ1WT, n = 8–12; αβ1S04A, n = 17–32; αβ1V120A, n = 7–11. Error bars represent SEM.

Figure 3. Steady-state effects of mutations on β1 function. (A) Summarized steady-state effects of 13 alanine substitutions measured in 60 µM Ca^2+. Averaged V_{1/2} (top) and Q (bottom) for BK channels with no β1, wild-type β1, or mutant β1. α, n = 16; αβ1WT, n = 12; αβ1G73A, n = 16; αβ1P75A, n = 15; αβ1C76A, n = 16; αβ1C76A, n = 20; αβ1S104A, n = 16; αβ1D80A, n = 6; αβ1C103A, n = 8; αβ1S104A, n = 32; αβ1I106A, n = 26; αβ1T107A, n = 11; αβ1P107A, n = 14; αβ1V120A, n = 11. Error bars represent SEM. (B) Positions of key segments A and B on a schematic cartoon of β1. Residues mutated to alanine are labeled with their respective amino acids. Mutated residues having relatively large and small effects are represented by closed red and black circles, respectively. Identical and nonidentical but conserved residues in other positions are represented by closed black and gray circles, respectively.
Y105A. For example, at −200 mV, deactivation time constants are 1.3 ± 0.2 ms and 1.1 ± 0.1 ms, respectively (P = 0.1).

The aforementioned results show that at 60 µM Ca²⁺, steady-state and kinetic effects of segment A and B mutations are nonadditive, suggesting that these residues functionally interact in modulating BK channel gating. The BK channel dual allosteric gating model includes a gate, four independent voltage sensors, and four Ca²⁺ sensors (Rothberg and Magleby, 2000; Horrigan and

Figure 4. Alanine substitution of segment A and B residues displays nonadditive effects. (A) Both Y105 and I106 resides in segment B. (B) The effects of Y105A and I106A are nonadditive. (left) The positive shift of G-V by Y105AI106A (n = 10) is not greater than Y105A (n = 26) or I106A (n = 11). (right) Y105AI106A (n = 8–10) does not slow activation time constants more than both single mutations. αβ1Y105A, n = 13–24; αβ1I106A, n = 8–11. (C) Y105 and Y74 reside in segment B and A, respectively. (D) The effects of Y105A and Y74A are nonadditive. (left) The positive shift of G-V by Y105AY74A (n = 24) is not greater than Y105A. (right) Y105AY74A (n = 22–24) does not slow activation or speed deactivation time constants more than either single mutation. αβ1Y74A, n = 8–14; αβ1Y105A, n = 13–24. (B and D, left) The black and gray traces represent Boltzmann fits of averaged G-V relations of αβ1 and α channels, respectively. (right) The black and gray traces represent averaged activation and deactivation time constants of αβ1 and α channels, respectively. Error bars represent SEM.

Figure 5. Measurement of Po at 0 Ca²⁺ and the limiting slope using F380Y. (A) Representative single-channel recordings of αβ1F380Y channels at 0 Ca²⁺ and decreasing voltages. (B) Corresponding all-point amplitude histograms and estimates of Po. The estimated number of channels in the patch is 74.
Aldrich, 2002). Channel opening is allosterically coupled to voltage sensor activation and Ca$^{2+}$ binding. However, channels can open at a low probability independent of voltage sensor activation and Ca$^{2+}$ binding (intrinsic gating). Previously, β1 subunits have been shown to confer modulatory effects on intrinsic gating, voltage sensor activation, and Ca$^{2+}$ binding (Cox and Aldrich, 2000; Nimigean and Magleby, 2000; Bao and Cox, 2005; Orio and Latorre, 2005; Wang and Brenner, 2006; Sweet and Cox, 2009). Because segments A and B are located in the extracellular region, we focused on how these domains modulate Ca$^{2+}$-independent effects: intrinsic gating and voltage sensor activation.

**Segment A and B residues reduce intrinsic gating**

Intrinsic gating is weakly voltage dependent and can be described by two free parameters (Horrigan and Aldrich, 2002). $L_0$ represents the zero voltage value of the closed-to-open (C-O) equilibrium constant, and $z_L$ is its partial charge (Horrigan and Aldrich, 2002). To isolate mutant effects on intrinsic gating, we performed recordings using a modified α subunit F380Y. Previously, this modification has been shown to greatly increase channel opening in hslo1 (Lippiat et al., 2000) and mslol (Wang and Brenner, 2006). F380Y allows us to measure channel $P_o$ in ligand-unbound states (0 Ca$^{2+}$) with voltage sensors residing in the resting state (the limiting slope). In $\alpha_{WT}$ background, obtaining $P_o$ under these conditions is not technically feasible for the $\alpha\beta_1$ channels ($P_o < 10^{-8}$; Wang and Brenner, 2006).

An example recording of α subunit F380Y in 0 Ca$^{2+}$ over a range of voltages is shown in Fig. 5 A. The corresponding all-point histograms and estimated $P_o$ (Fig. 5 B) clearly show that the voltage dependence approaches a minimum between −120 and −220 mV. The estimated weak voltage dependence here (∼0.25 e$^0$) corresponds to the weak voltage dependence associated with intrinsic gating ($z_L$; Horrigan and Aldrich, 2002). Fitting 0 Ca$^{2+}$ limiting slope log$P_o$-$V$ relations to the Horrigan-Aldrich model (Horrigan and Aldrich, 2002), the two free parameters associated with intrinsic gating were estimated ($L_0$ of 6.6 ± 0.9 e$^{-0}$ and $z_L$ of 0.16 ± 0.01 e$^0$; Fig. 6 A and Table 1). For $\alpha_{F380Y}$, the effect of $\beta_1_{WT}$ on intrinsic gating is an ∼15-fold reduction in $L_0$ (4.3 ± 1.3 e$^{-3}$, $P < 0.001$). However, $z_L$ is not significantly altered (0.20 ± 0.05 e$^0$, $P = 0.5$; Table 1).

$L_0$ is significantly increased by segment A and B key mutations (Table 1). In the presence of Y105A, I106A, or Y74A, $L_0$ are 1.9 ± 0.4, 4.4 ± 1.1, or 2.5 ± 0.5 e$^0$ (Table 1). The ∼4-, 10-, and 6-fold increases of $L_0$ (relative to $\alpha_{F380Y}\beta_{1WT}$ channels) reflect a reduction in $\beta_1$’s ability to decrease intrinsic gating. The results suggest an important role that Y105, I106, and Y74 play in determining $L_0$. Double mutation Y105A-Y74A was also examined to test whether effects of Y105A and Y74A on intrinsic gating were additive. If effects of Y105A and Y74A on intrinsic gating were additive, a significant increase in $L_0$ would have been expected. Because the estimated $L_0$ for $\alpha_{F380Y}\beta_1$ Y105A-Y74A, 2.0 ± 0.3 e$^0$, is similar to the single mutations, the results suggest that segments A and B functionally interact in reducing intrinsic gating.

**Segment A and B residues modulate open-channel voltage sensor activation**

An increase in $L_0$ predicts an increase in steady-state opening at all Ca$^{2+}$. This is unlikely to be the sole effect of segment A and B mutants because the mutations displayed positive shifts of G-V relations. We therefore investigated whether these mutations also alter voltage sensor activation. Voltage sensor activation of unliganded channels is described by three free parameters: the partial charge associated with the resting-to-activated (R-A) transition ($z_L$) and the half-activation voltages for voltage

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**Figure 6.** Segment A and B residues contribute to reducing intrinsic opening. (A) A representative log$P_o$-$V$ relation of $\alpha_{F380Y}$ channel in which the limiting slope was fitted to the Horrigan-Aldrich model to estimate $z_L$ and $L_0$. The black curve represents the Boltzmann fit plotted in a log scale (right axis not shown, axis range 0.01–1). The red line represents the linear fits of log$P_o$-$V$ relations to the Horrigan-Aldrich model between −160 and −90 mV. Weak voltage dependence of $P_o$ at the limiting slope is apparent when compared with the Boltzmann fit. (B) Averaged estimates for $L_0$ and $z_L$ for different channel configurations. No $\beta_1$, $n = 5$; WT $\beta_1$, $n = 7$; Y105A, $n = 8$; I106A, $n = 8$; Y74A, $n = 9$; Y105AY74A, $n = 9$. Error bars represent SEM.
found that neither z_L nor z_J are altered by \( \beta_1 \) (Bao and Cox, 2005), suggesting that mutations likely do not alter z_L or z_J either.

The fits to the logP_o-V relations (Table 2) show that \( \beta_1 \) stabilizes voltage sensor activation in both open and closed channels. There is a \( -55 \text{ mV} \) shift of the half-activating voltage for the open-channel voltage sensor (V_ho), similar to the \( -61 \text{ mV} \) value estimated in the WT background (Bao and Cox, 2005). \( \alpha_{F380Y} \) reports a \( -37 \text{ mV} \) shift of the half-activating voltage for the closed-channel voltage sensor (V_hc), which is smaller than the \( -71 \text{ mV} \) value estimated in the WT background (Bao and Cox, 2005).

Y105A has little effect on closed-channel voltage sensor activation. This is evident from estimates of V_hc (76 vs. \( 73 \text{ mV} \), respectively; Horrigan and Aldrich, 2002). The R-A equilibrium constant of closed, unliganded channel J_0 (the zero voltage value of the R-A equilibrium constant) is a function of V_hc (\( J_0 = 0.5/\exp(z_JV_hc/kT) \)). The allosteric factor between the voltage sensor activation and gating (D) is a function of the difference between V_hc and V_ho (\( D = 0.5/\exp(z_J(V_hc - V_ho)/kT) \)); Horrigan and Aldrich, 2002).

We obtained \( 0 \text{ Ca}^{2+} P_o \) over a wide range of voltages. Averaged logP_o-V and P_o-V relations data were fit to the Horrigan-Aldrich model (Fig. 7 and Fig. S3; Horrigan and Aldrich, 2002). In all these fits, \( z_L \) and \( z_J \) were set as 0.2 e_0 and 0.58 e_0, respectively, to reduce the number of free parameters. The basis for our assumptions here are twofold. First, the finding that voltage sensors act as a source of gating charge for the opening transition (Ma et al., 2006) and the observation that the mutations do not alter z_L imply that these mutations do not significantly alter z_J as well. Second, prior gating current data

### Table 1

<table>
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<tr>
<th>Channels</th>
<th>( I_0 ) (approximately)</th>
<th>P-value</th>
<th>( z_L )</th>
<th>P-value</th>
<th>( z_J )</th>
<th>( \Delta \Delta G )</th>
<th>( n )</th>
</tr>
</thead>
<tbody>
<tr>
<td>No ( \beta_1 )</td>
<td>6.6 ( \pm ) 0.9</td>
<td>( &lt;0.001 )</td>
<td>0.16 ( \pm ) 0.01</td>
<td>0.5</td>
<td>( e_0 )</td>
<td>kcal/mol</td>
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<td>0.20 ( \pm ) 0.05</td>
<td>NA</td>
<td>1.62</td>
<td>7</td>
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<tr>
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<td>1.9 ( \pm ) 0.4</td>
<td>( &lt;0.01 )</td>
<td>0.21 ( \pm ) 0.02</td>
<td>0.8</td>
<td>0.74</td>
<td>8</td>
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</tr>
<tr>
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<td>4.4 ( \pm ) 1.1</td>
<td>( &lt;0.01 )</td>
<td>0.24 ( \pm ) 0.04</td>
<td>0.5</td>
<td>0.39</td>
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<tr>
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<td>( &lt;0.01 )</td>
<td>0.20 ( \pm ) 0.02</td>
<td>0.7</td>
<td>0.58</td>
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<tr>
<td>( \beta_1 )I106AY105A</td>
<td>2.0 ( \pm ) 0.3</td>
<td>( &lt;0.001 )</td>
<td>0.21 ( \pm ) 0.02</td>
<td>0.9</td>
<td>0.71</td>
<td>9</td>
<td></td>
</tr>
</tbody>
</table>

NA: not applicable.

\( e_0 \): \( \text{e}^{-1} \).

Figure 7. Segment A and B residues contribute to stabilization of open-channel voltage sensor activation. (A–F) Averaged logP_o-V relations (circles) and best fits to the Horrigan-Aldrich model (red curves). The black curves represent Boltzmann fits. No \( \beta_1 \), \( n = 5–13 \); WT \( \beta_1 \), \( n = 3–30 \); Y105A, \( n = 7–24 \); I106A, \( n = 4–8 \); Y74A, \( n = 5–12 \); Y105AY74A, \( n = 6–12 \). Error bars represent SEM.
of single mutations are nonadditive. Therefore, segments A and B may functionally interact in stabilizing voltage sensor activation in open channels.

**Discussion**

Our scan identified four residues critical for β1 gating modulation. Mutating these residues reduce β1-mediated negative G-V shift at 60 Ca²⁺, consistent with a partial disruption in β1 function. The experimental evidence does not suggest gain-of-function mutant effects. For example, gating parameters normally not altered by β1WT (such as zL) are also not altered by these mutations, and gating parameters normally altered by β1WT (such as L⁰ and Vho) are reduced by these mutations.

Previous studies suggest that β1 alters several gating parameters of BK channels (Cox and Aldrich, 2000; Bao and Cox, 2005; Orio and Latorre, 2005; Orio et al., 2006; Wang and Brenner, 2006; Sweet and Cox, 2009). Our observations that no single mutation eliminated β1-mediated negative G-V shift or free energy changes suggest that additional key gating residues remain to be uncovered. This is true even for the effects on intrinsic gating and open-channel voltage sensor activation that
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intrinsic opening and voltage sensor activation (Ma et al., 2006; Wang and Brenner, 2006), this coupling may reflect intrinsic properties of the pore-forming subunit rather than properties unique to β1 function.

The dual gating effects of the segment A-B domain likely contribute to the complex steady-state modulatory effects of β subunits (Fig. S1). Interactions involving the segment A-B domain transition the channel away from open states with relaxed voltage sensors (the OR state; Fig. 9). Thus, in the absence of calcium or membrane depolarization, β1 causes a relative destabilization of the OR state and “silences” the channel in the closed resting state (CR; Fig. 9). With sufficient depolarization and/or calcium, β1 promotes channel opening by facilitating transitions to the late opening states (the OA state; Fig. 9).

Our results suggest that the segment A-B domain is necessary for maintaining the regulatory effects of β1. The data do not exclude the possibility that this domain plays an indirect role, simply being required to maintain a particular β or α/β subunit structure necessary for β1 subunit gating effects. However, the fact that segments A and B are the most conserved domain led us to favor a simpler scenario that this domain mediates the modulatory effects directly.

Several studies suggest that the extracellular segments A and B may be positioned near the external vestibule of BK channels (Fig. 9). Extracellular residues of β subunits have been shown to affect charybdotoxin binding and instantaneous I-V relations (Hanner et al., 1998; Zeng et al., 2003; Chen et al., 2008). Interestingly, hβ2 lysine residues flanking segment B (Fig. 1) have been shown to confer outward rectification of BK currents.

<table>
<thead>
<tr>
<th>Channels</th>
<th>ΔΔG (kcal/mol)</th>
</tr>
</thead>
<tbody>
<tr>
<td>No β1</td>
<td>-1.96 2.96 0.99 -1.99</td>
</tr>
<tr>
<td>β1R105A</td>
<td>-1.02 1.24 -0.38 0.16</td>
</tr>
<tr>
<td>β1I106A</td>
<td>-1.45 1.56 0.10 -0.22</td>
</tr>
<tr>
<td>β1Y74A</td>
<td>-0.97 1.29 -0.37 0.05</td>
</tr>
<tr>
<td>β1Y105A</td>
<td>-1.16 1.13 -0.18 0.22</td>
</tr>
</tbody>
</table>

Table 3: Free energy change relative to wild-type β1.

Our finding that mutations of β1 disrupt effects on intrinsic gating and voltage sensor activation is consistent with previous studies ascribing β1 modulation of BK channels to these two effects (Nimigean and Magleby, 2000; Bao and Cox, 2005; Orio and Latorre, 2005; Wang and Brenner, 2006). Indeed, mutations of the BK channel voltage sensor have been suggested to specifically occlude the β1-mediated negative G-V shift (Yang et al., 2008). Based on the observation that single-residue mutations both reduce the energetic barrier for intrinsic opening and increase the energetic barrier for open-channel voltage sensor activation, a novel insight provided by the current study is that β1 modulation of these two aspects of gating is likely coupled. Because previous studies have identified single α mutations that alter both intrinsic opening and voltage sensor activation (Ma et al., 2006; Wang and Brenner, 2006), this coupling may reflect intrinsic properties of the pore-forming subunit rather than properties unique to β1 function.

The dual gating effects of the segment A-B domain likely contribute to the complex steady-state modulatory effects of β subunits (Fig. S1). Interactions involving the segment A-B domain transition the channel away from open states with relaxed voltage sensors (the OR state; Fig. 9). Thus, in the absence of calcium or membrane depolarization, β1 causes a relative destabilization of the OR state and “silences” the channel in the closed resting state (CR; Fig. 9). With sufficient depolarization and/or calcium, β1 promotes channel opening by facilitating transitions to the late opening states (the OA state; Fig. 9).

Our results suggest that the segment A-B domain is necessary for maintaining the regulatory effects of β1. The data do not exclude the possibility that this domain plays an indirect role, simply being required to maintain a particular β or α/β subunit structure necessary for β1 subunit gating effects. However, the fact that segments A and B are the most conserved domain led us to favor a simpler scenario that this domain mediates the modulatory effects directly.

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Figure 9. A hypothetical mechanism for β1 function. Cartoons illustrate how the β1 extracellular domain may affect both intrinsic gating and voltage sensor (VS) activation. The pore and voltage sensor domains are depicted in four combinations: closed resting (CR), open resting (OR), open activated (OA), and closed activated (CA). The segment A-B domain promotes transitions away from open states with no or few voltage sensors activated (OR). At rest, the domain reduces opening by causing a relative destabilization of the OR state and promoting transitions to the CR states. With sufficient depolarization, the domain stabilizes channel opening by promoting transitions to the late opening states (OA). The segment A-B domain may selectively interact with the open but not closed channels.
(Chen et al., 2008). These results suggest close approximation between the outer vestibule of the channel and the β subunit extracellular domain (including the segment B region). Assuming the role of the segment A-B domain is direct, how do β subunit residues near the external mouth of BK channels modulate gating? There has been compelling evidence that suggests the selectivity filter is the activation gate for BK channels (Li and Aldrich, 2004; Piskorowski and Aldrich, 2006; Wilkens and Aldrich, 2006; Chen and Aldrich, 2011; Cox and Hoshi, 2011; Geng et al., 2011; Zhou et al., 2011). Proximity between the outer vestibule and the selectivity filter suggests that the segment A-B domain may be positioned near the gate. Our finding that the segment A and B mutations affect both voltage sensing and intrinsic gating presents the intriguing possibility that this domain may interact at a nexus for coupling open-channel voltage sensor to the selectivity filter/channel gate. Indeed, it will be interesting to determine whether physical interactions between the segment A-B domain and the pore-forming subunits are state dependent (i.e., interactions only occurring in open states; Fig. 9).

Alternatively, the segment A-B domain may interact with the pore-forming subunits indirectly, via other extracellular residues or other parts of the channel. Using chimera and deletion approaches, the intracellular domains of β2 have also been shown to be important for β1 function (Orio et al., 2006; Wang and Brenner, 2006). In addition, β1 TMs have been shown to lay in a position between voltage sensor domains of adjacent α subunits (Liu et al., 2010), suggesting a role these domains play in modulating voltage sensor activation. The relations among various β subunit domains and their relevant contributions remain an important question to be addressed in future studies.

In summary, this study identifies an important domain of β1 subunits that underlies, in part, modulation of BK/β1 channels. The reduced intrinsic opening conferred by the segment A-B domain and increased voltage sensor activation created a steeper activation response of BK channels associated with β1. The fact that human β1 polymorphisms with more moderate gating effects have been linked to decreased prevalence of hypertension and reduced pulmonary function (Fernández-Fernández et al., 2004; Seibold et al., 2008) highlights the potential physiological importance of the segment A-B domain in β1 and perhaps other β subunits.

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